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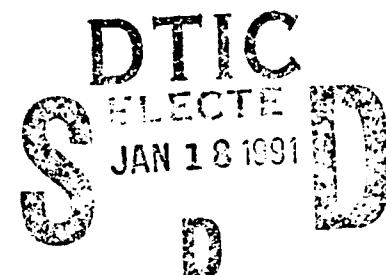
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Towards Wound Closure Optimization: Final Report

Larry I. Sanders



LaserSurge, Inc.

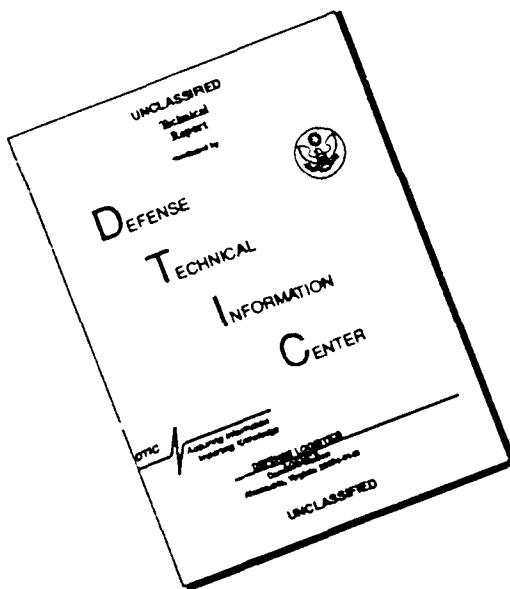
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Summary

Extensive experimentation with many combinations of lasers and chromophores was conducted to determine the optimal parameters for laser tissue welding. New Zealand White rabbit small bowel was used as an *in vitro* model for this study. The analysis indicates that particular chromophores can be utilized to enhance the effectiveness of certain lasers. For example, India ink is an effective chromophore for the 1.06 micron Nd:YAG laser, while blood is most effective for the 0.514 micron Ar⁺⁺ laser. The 2.1 micron Ho:YAG laser produced strong chromophore-free welds, and also produced strong welds in the presence of India ink and blood. The laser welding literature indicates that Indocyanine Green is an adjuvant to laser welding with the 0.808 micron Ga:Al:As diode laser. The diode lasers that were examined in this study did not provide sufficient energy to weld tissue and evaluate these statements. Indocyanine Green did prove to be an effective chromophore for the 0.532 micron KTP laser. The apparatus designed and developed for this study provides an unique method to determine the laser tissue welding parameters which will produce the strongest welds.

Introduction

LaserSurge, Inc, of Rochester, NY was established to develop, demonstrate, and facilitate the clinical use of automated surgical laser delivery systems. The speed, effectiveness, and reliability of using laser energy to connect or weld living tissue makes laser tissue welding one of the most promising areas of medical research today. The controlled application of laser energy thermally induces intrinsic tissue changes which lead to both immediately strong bonds between tissue and rapid restoration of tissue function. Conventional surgical techniques typically require sutures and staples to close tissue wounds or to construct anastomoses (i.e., surgical connections that provide functional communication between hollow organs such as bowel). A surgeon's bonding of tissue using laser tissue welding can be compared to a metal worker's handling of steel. Whereas the conventional suturing or stapling is like the use of screws or rivets in steel, laser tissue welding, like welding steel, uses an outside energy source to cause intrinsic material changes which alone provide for the strong bond. A fundamental requirement to weld either material is that the surfaces to be welded be in intimate contact with each other. To fully realize the enormous potential of this revolutionary wound closure modality, an enhanced theoretical understanding of laser tissue welding is essential. In an effort to better recognize and evaluate the many factors that determine a successful laser tissue weld, LaserSurge has developed a laser tissue welding equation which describes laser tissue welding as a function of its component parameters; lasing parameters, chromophores, and tissue parameters. Using this structured approach, an investigator can readily dissect and compare the effects of changes in individual or multiple variables. This study was designed to identify the most favorable laser tissue welding conditions. To accomplish this goal, an integrated laser tissue welding and tensiometry (weld strength measurement) system was designed, constructed, and tested. This system permits the user to selectively vary several parameters of interest including welding temperature, welding time, aperture size, laser type, power, and chromophores. Experimental trials using different combinations of the aforementioned parameters were performed in this study.

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Methods, Assumptions, and Procedures

Methods

Device Refinement and Evaluation

In order to examine the effects of multiple combinations of lasers and chromophores, a new device was developed to quantify the strength of laser welds. The laser welding/tensiometry apparatus permits any laser which can be transmitted through an optical fiber to be coupled to the beam alignment system. The beam alignment system configured the beam so that it slightly overfilled the aperture. The beam passed through a high-speed shutter, which opened and closed in response to the thermocouple temperature reading. The aperture size was fixed at either 0.5×4.0 mm or 2.0×4.0 mm, and the aperture was placed just above the surface of the tissue strips to be welded. The beam impinged perpendicularly upon the tissue strips at the site of apposition. A thermocouple was placed between the strips so that it was positioned 1 mm beneath the surface of the weld. The strips were then brought into contact by a piezoelectric motor under computer control. The laser welding temperature control program allowed the operator to designate the parameters of interest, including the welding control temperature. Following welding, the welded tissue was pulled apart and the load required to rupture the weld was measured as a function of the distance moved. Graphs of load versus distance were generated to determine maximum weld strength.

The tensiometry software was seamlessly integrated into the laser welding control program by Mr. William Kraft. This new program eliminated the need to reboot the computer between experimental trials. This major improvement permitted more rapid data acquisition and reduced the average time between trials to approximately five minutes.

The thermocouple placement system has also been modified to provide more reproducible and accurate temperature measurements. A single thermocouple was mounted on a specially-designed plexiglas holder. This thermocouple was placed 1 mm below the surface of the apposed tissue at the site of welding, as depicted in Figure 1 in Appendix C. The temperature recorded from the thermocouple controlled the opening and closing of the shutter mechanism, thereby maintaining the designated tissue welding temperature. The new thermocouple holder permits easier, faster, and more reliable thermocouple placement.

A new problem surfaced during the experimentation. The stages began to stall and resist free movement because of dried bowel and saline which had entered the stage. The stages were removed, cleaned, and remounted. Periodic cleaning and maintenance have rectified this problem.

Assumptions

An implicit assumption in the original design of this experimental protocol is that the freshly harvested *in vitro* bowel very closely resembles native *in vivo* bowel. The time from resection to welding was minimized by sequentially harvesting small sections of bowel from intravenously anesthetized rabbits. The resected bowel was placed in physiological saline to maintain its integrity, while the remaining intact bowel was returned to the

abdomen for later use. Tissue processing and handling were minimized to the extent possible. Tissue trauma was minimized by handling and manipulating the bowel gently. We have assumed that the *in vitro* tissue would not be significantly altered by the removal and processing steps. The veracity of this assumption is important since the intent of this study is to generate data which will be readily transferrable to clinical applications.

One set of experiments suggests that the *in vitro* bowel model may not be wholly representative of the *in vivo* state. When more than one hour elapsed between bowel resection and welding, the welds produced were clearly inferior to welds produced using freshly harvested bowel under the same conditions. This lack of success may be attributable to changes in tissue turgidity and perhaps also to other biological effects (platelet aggregation and adhesion, etc.). The bowel turgidity appears to change as a result of storage in physiological saline. It is unclear whether this change might influence the laser welding of bowel. Ischemia and oxygen deprivation may also cause changes which are responsible for the difference seen between welding *in vitro* and *in vivo*.

As a consequence of the *in vitro* alterations to bowel, the weld strengths obtained in this study were much lower than anticipated based upon our earlier *in vivo* data. In order to obtain recognizable welds, the highest laser power and temperature settings were employed. Several of the lasers, namely the diode and Argon⁺⁺, were unable to provide the high power output required to weld tissue. Thus, the results for the diode and Argon lasers are inconclusive, and few or no welds were performed using these lasers.

Experimental Procedure

The experimental protocol required the resection of small bowel from New Zealand White rabbits. As discussed in the third quarter 1990 report, weld strength appears to diminish as the length of time between bowel harvesting and welding increases. In order to more closely emulate the clinical setting, fresh bowel was sequentially resected from an intravenously anesthetized rabbit (see Figure 2 in Appendix C for a complete description of the experimental protocol). The harvested tissue was cut into strips measuring 4 mm x 20 mm. The strips were carefully positioned on the laser welding/tensiometry apparatus. The thermocouple was positioned 1 mm deep at the site to be welded and the tissue strips were brought into apposition. Next, the laser was fired and, following the laser exposure, a welded tissue seam was created where the tissue strips were in contact. Thus, the thermocouple was imbedded in the welded tissue. The laser welding temperature control software directed the opening and closing of the shutter to maintain the designated welding temperature. Immediately following laser welding, the tissue was pulled apart and the load required to break the bond (in grams) was measured as a function of the distance (in millimeters) moved. Load versus distance plots were generated to assess the strength of each weld. Maximum strength was determined for each weld.

The 0.808 micron diode lasers used in this study were not powerful enough to use for laser welding using the apparatus as designed. Both LaserSurge's Spectra Diode Laboratory 3 W laser diode array and Columbia-Presbyterian's 1.5 W IRIS Medical OcuLight SL diode were coupled to the laser welding/tensiometry apparatus. The power delivered to the weld site, as measured by a power meter, was insufficient to weld tissue, since less than 0.15 W was transmitted through the optical delivery system. No appreciable temperature rise was observed, even with the Indocyanine Green chromophore. Thus, further studies of the diode laser and its interaction with chromophores were not performed.

Another laser which was initially included in this study was the 1.32 micron Nd:YAG. Rochester General Hospital has one 1.32 micron laser, and it had not been used in more than one year. When the laser was turned on, the laser's power setting mechanism did not function. This laser was unable to be repaired prior to the conclusion of this study. Thus, the 1.32 micron Nd:YAG laser was not studied further.

The Rochester General Hospital 0.514 micron Ar⁺⁺ laser was studied. This laser, a 7.5 W Lexel Aurora Model 150 Pump Laser and a Model 600 Dye Laser, is a tremendously large research unit. The delivered power was quite low, approximately 0.7 W, and this precluded testing of some chromophore and temperature combinations.

The 100 W, CW, Sharplan 2100 1.06 micron Nd:YAG laser was examined using all chromophores and both aperture sizes. The maximum power of 5.0 W (delivered to the site of welding) was selected for use throughout since, at the lower temperatures even with this high power, weld strength was low. Thus, the lower power settings were not examined.

The Columbia-Presbyterian Medical Center in New York, NY has a Coherent Two Point One Holmium laser, with a maximum output power of 15 W operating in pulsed mode. The 2.1 micron Ho:YAG was examined at approximately 1 and 5 W with the small and large apertures.

The 20 W, CW, LaserScope 0.532 KTP laser was also examined in this study. Both aperture sizes were examined at the highest obtainable delivered power of 4 W.

The power delivered to the weld site through the optical system was occasionally not sufficient to attain some of the higher temperatures. These results are indicated in the data section that follows.

Results and Discussion

Results

The data collected and included in the subsequent analysis are limited in number. Most of the early data collected using the ASYSTANT data acquisition program has been excluded due to problems with the tensiometry apparatus, as will be explained in the Discussion (see Appendix B for these force versus distance plots). A great deal of time was devoted to software development and testing, and therefore, the results reported herein generally come from one or two samples. A thorough statistical analysis was not performed; rather, averages were obtained for each laser and chromophore combination. These averages were assessed to determine the most promising combinations. The averages are presented in Table 1.

TABLE 1- MAXIMUM WELD STRENGTH (g)

0.514 Micron Argon (Small Aperture, 0.7-0.8W delivered)

<u>TEMP</u>	<u>BLOOD</u>	<u>ICG</u>	<u>INDIA INK</u>
UNWELDED	0.9	0.6	0.2
50° C	1.2	0.7	1.1
60° C	1.4	1.0	0.9
70° C	-	0.7	1.3
80° C	3.8	-	-
100° C	-	-	-

0.532 Micron KTP (Small Aperture, 4.0 W delivered)

<u>TEMP</u>	<u>BLOOD</u>	<u>ICG</u>	<u>INDIA INK</u>
UNWELDED	0.7	0.1	0.5
50° C	1.7	1.0	0.4
60° C	1.1	2.2	0.5
70° C	1.7	2.7	0.9
80° C	1.3	1.3	1.0
100° C	2.1	3.4	0.6

0.532 Micron KTP (Large Aperture, 4.0 W delivered)

<u>TEMP</u>	<u>BLOOD</u>	<u>ICG</u>	<u>INDIA INK</u>
UNWELDED	-	0.1	-
50° C	-	0.7	-
60° C	-	0.8	-
70° C	-	1.0	-
80° C	-	1.0	-
100° C	-	3.7	-

1.06 Micron Nd:YAG (Small Aperture, 5.0 W delivered)

<u>TEMP</u>	<u>BLOOD</u>	<u>ICG</u>	<u>INDIA INK</u>
UNWELDED	0.2	0.1	0.5
50° C	0.9	0.9	1.0
60° C	1.5	1.1	0.7
70° C	1.6	1.2	2.8
80° C	1.9	1.0	2.0
100° C	1.8	1.1	18.6

1.06 Micron Nd:YAG (Large Aperture, 5.0 W delivered)

<u>TEMP</u>	<u>BLOOD</u>	<u>ICG</u>	<u>INDIA INK</u>
UNWELDED	-	-	1.0
50° C	-	-	1.0
60° C	-	-	1.0
70° C	-	-	1.0
80° C	-	-	1.25
100° C	-	-	2.0

2.1 Micron Ho:YAG (Small Aperture, 5.0 W delivered)

<u>TEMP</u>	<u>BLOOD</u>	<u>ICG</u>	<u>INDIA INK</u>	<u>NO CHROMO</u>
UNWELDED	0.6	1.7	0.6	0.7
50°	1.8	3.4	5.6	4.0
60°	3.7	2.9	9.4	5.8
70°	4.8	9.6	7.0	7.9
80°	13.6	5.9	21.0	8.9
100°	29.4	10.9	21.0	20.8

2.1 Micron Ho:YAG (Large Aperture, 5.0 W delivered)

<u>TEMP</u>	<u>BLOOD</u>	<u>ICG</u>	<u>INDIA INK</u>	<u>NO CHROMO</u>
UNWELDED	0.6	0.9	1.0	0.8
50° C	0.6	4.0	5.4	13.9
60° C	1.5	11.4	2.4	1.1
70° C	2.8	2.8	15.3	2.8
80° C	31.3	45.0	14.2	34.0
100° C	36.5	50.8	2.1	39.0

2.1 Micron Ho:YAG (Small Aperture, 1.0 W delivered)

<u>TEMP</u>	<u>BLOOD</u>	<u>ICG</u>	<u>INDIA INK</u>	<u>NO CHROMO</u>
UNWELDED	1.3	0.8	1.6	1.2
50° C	2.1	2.1	1.7	1.7
60° C	2.8	0.4	2.4	1.4
70° C	2.4	4.8	3.4	1.5
80° C	1.5	3.6	7.0	2.1
100° C	9.5	5.9	17.9	2.9

2.1 Micron Ho:YAG (Large Aperture, 1.0 W delivered)

<u>TEMP</u>	<u>BLOOD</u>	<u>ICG</u>	<u>INDIA INK</u>	<u>NO CHROMO</u>
UNWELDED	1.0	0.3	0.2	0.2
50° C	6.7	2.7	1.3	1.5
60° C	6.1	6.8	0.9	-
70° C	5.5	24.5	2.0	4.4
80° C	4.9	4.1	0.8	1.4
100° C	11.3	5.3	3.1	4.1

NOTES:

BLOOD = fresh whole blood collected in a Vacutainer blood collection tube with heparin.

ICG = prepared as a 2X solution (25 mg Indocyanine Green per 10 ml aqueous solvent).

- = unable to perform.

Discussion

Thermocouple Placement

The earlier observation that thermocouple placement is crucial continued to be supported by experimentation. If the thermocouple is placed above the weld site, the recorded temperature is an artefact which corresponds to the heating of the thermocouple and not to the heating of the tissue at the weld site. If the thermocouple is placed too deeply, it again fails to accurately represent the temperature increase at the site of welding since the recorded temperatures are lower than those at the weld. Since the laser welding temperature control program controls the temperature at the weld site based upon the thermocouple measurements, proper thermocouple placement is imperative. The final thermocouple holder design provides adequate reproducibility. When thermocouple placement was incorrect, as evidenced by either an abnormally short or a protracted time to attain the stipulated control temperature, the trial was repeated with fresh tissue strips.

Prior to the design and construction of the new thermocouple holder, there were major problems with thermocouple placement. The thermocouple was difficult to position reproducibly, and thus, much of the early data is erroneous. Since the temperature control program utilizes feedback from the thermocouple, and since the thermocouple was incorrectly positioned, the time of laser exposure as well as the actual temperature at the weld site were incorrect as well.

Chromophore Delivery and Placement

Precise chromophore placement was another important factor in this study. Each of the chromophores tested was a liquid. Attempts were made to deposit the chromophore at the weld site using a micropipettor. This allowed a fixed amount of chromophore to be deposited, but the application was not evenly distributed along the weld seam. A cotton-tipped applicator was used to apply a barely visible coat of chromophore to both tissue strips prior to bringing the strips into apposition. The strips were then brought into contact by the computer-controlled piezoelectric motor. When the requisite apposition was obtained, the top surface of the weld site was gently wiped with a clean cotton-tipped applicator to remove any chromophore which oozed upwards when the tissue was compressed. When too much chromophore was placed at the top of the weld site, excessive energy was absorbed at the surface, rather than in the deeper areas of the weld site, and smoking, charring, and diminished weld strength were noted. Chromophore placement in the deeper half of the weld site resulted in stronger, more homogeneous welds.

Maintenance of equipment, software package, and its effect on data.

Maintenance of the laser welding/tensiometry apparatus was crucial. The stages upon which the tissue is clamped are highly susceptible to stalling if they become contaminated with dried bowel and saline. Stalling of the motor due to a contaminated stage was readily apparent on the first control run of the day, as the tensiometry plots of force versus distance indicated the resistance of the stage to movement. Periodic removal of the stages for cleaning and maintenance was a necessity. The outside surfaces of the laser welding/tensiometry apparatus were carefully cleaned after each use. These steps prevented corrosion of the apparatus.

Initially, however, the problem of stalling was not so readily apparent. Using the original ASYSTANT data acquisition software package, we were unable to obtain plots of the data in either real time or immediately after the completion of the trial. To plot data in ASYSTANT required a lengthy process of file conversion and computer rebooting. Thus, data was not analyzed until after the animal had been sacrificed. This software problem resulted in the generation of tremendous amounts of erroneous data (see Appendix B), since it was impossible to interpret the effect of stage resistance on the measured weld strengths.

Since there were substantial problems with the software, several different software programs were utilized during this study. ASYSTANT was utilized for the earliest data acquisition, but it was cumbersome and inefficient. The time between welding and tensiometry was often more than five minutes. The final version of the temperature control and tensiometry program is written in BASIC and can switch from the welding temperature control program to tensiometry in a matter of seconds. The data acquired using the BASIC programs was imported into either Lotus 1-2-3 version 2.1 or Lotus Symphony release 2.0 for subsequent analysis and graphing.

All data were collected at the laboratory and were stored on 3.5 inch high density floppy disks to permit data analysis at the office. Additional backup copies of the data have been made for the LaserSurge software archives in case of loss or damage to the working disks. These disks are stored off site.

Overall results- difference in aperture size and its effect, power delivered and its effect, chromophores and their effect, time of welding and its effect

Aperture size was one parameter examined in this study. Two aperture sizes were utilized, 0.5 x 4.0 mm and 2.0 x 4.0 mm. The small aperture, with its attendant higher power density, generally produced stronger welds. This was true for all lasers examined, except for the Ho:YAG. For the 5.0 W Ho:YAG trials, the large aperture produced better welds with either Indocyanine Green or in the absence of chromophore. For the 1.5 W trials, the large aperture again produced stronger welds with Indocyanine Green, blood, and without a chromophore.

Power densities can also be calculated for the two aperture sizes used. The power densities, in W/cm^2 , were:

- Small Aperture, 1 W- 50
- Small Aperture, 5 W- 250
- Large Aperture, 1 W- 12.5
- Large Aperture, 5 W- 62.5.

Since the large aperture appeared to weld better than the small aperture for the Ho:YAG laser, perhaps the increased surface area of welding was responsible for the strong welds seen.

High and low power settings were also examined. The higher power of 5 W produced welds of superior strength when compared to the lower power welds. This is true for equal welding temperatures, and seems surprising since the lower power setting often required a longer total laser exposure to reach the desired welding temperature.

The effect of the length of exposure to laser energy on weld strength was not assessed in this study. Most of the temperature settings used required between 15 and 30 seconds to reach the designated control temperature. When the control temperature was attained, feedback from the thermocouple to the shutter control maintained the desired temperature for 10 s. Since there was significant variation in the amount of time required to reach the predetermined welding temperature, the differences due to the variation of controlled exposure would be very difficult to interpret. Thus, a single, fixed 10 s exposure was consistently used throughout the study.

The relationship between maximum temperature and weld strength is also unclear. The highest temperature examined, 100 degrees C, usually elicited smoking as well as tissue blanching and desiccation. Occasional charring was noted in some samples. As mentioned above, the time required to reach the desired temperature varied from sample to sample. Thus, the conclusion that weld strength appears to increase with increasing temperature must be tempered with caution. This is particularly true because the 100 degree welds were often slightly weaker than the 80 degree welds. We speculate that this might be due to increased thermal injury at the higher temperature. Many samples that were welded at 100 degrees became charred or carbonized, and the tissue was dry and brittle. This type of extreme thermal injury may compromise the strength of the weld, and may explain why the 100 degree welds were weaker than the 80 degree welds. Many of the 100 degree welds that were mildly desiccated and blanched seemed to be strongly welded. The overall pattern seemed to indicate that weld strength did increase with increasing temperature.

Conclusions

Laser tissue welding is successful when the correct combinations of laser wavelength, chromophore, and aperture size are utilized. Based upon the *in vitro* results obtained in this study, the following laser/chromophore combinations appear to show promise for laser tissue welding:

- 0.514 micron Ar⁺⁺- blood or ICG chromophore
- 0.532 micron KTP- blood or ICG chromophore
- 1.06 micron Nd:YAG- India ink chromophore
- 2.1 micron Ho:YAG- blood, India ink, or no chromophore.

In comparing the weld strength produced by each of these lasers, the Ho:YAG laser clearly produced the strongest welds. Many of the Ho:YAG welds appeared to be full thickness welds. The optimal welding temperature for any of the tested laser and chromophore combinations appeared to be between 80 and 100 degrees C. The higher power levels seemed to produce stronger welds, at least in the range of 1 to 5 W. A certain minimum power is required to increase the tissue temperature to the desired range (80 to 100 degrees C). Whether additional power above this threshold actually improves the strength of the welds cannot be asserted from the data collected.

Recommendations

The laser welding/tensiometry apparatus developed for this study provides an unique and ideal method to analyze the effects of various parameters on weld strength. The *in vitro* model has some limitations, and an *in vivo* corroboration of these results is needed. An *in vivo* model would permit an assessment of the effects of thermal injury and healing, two clinically relevant issues.

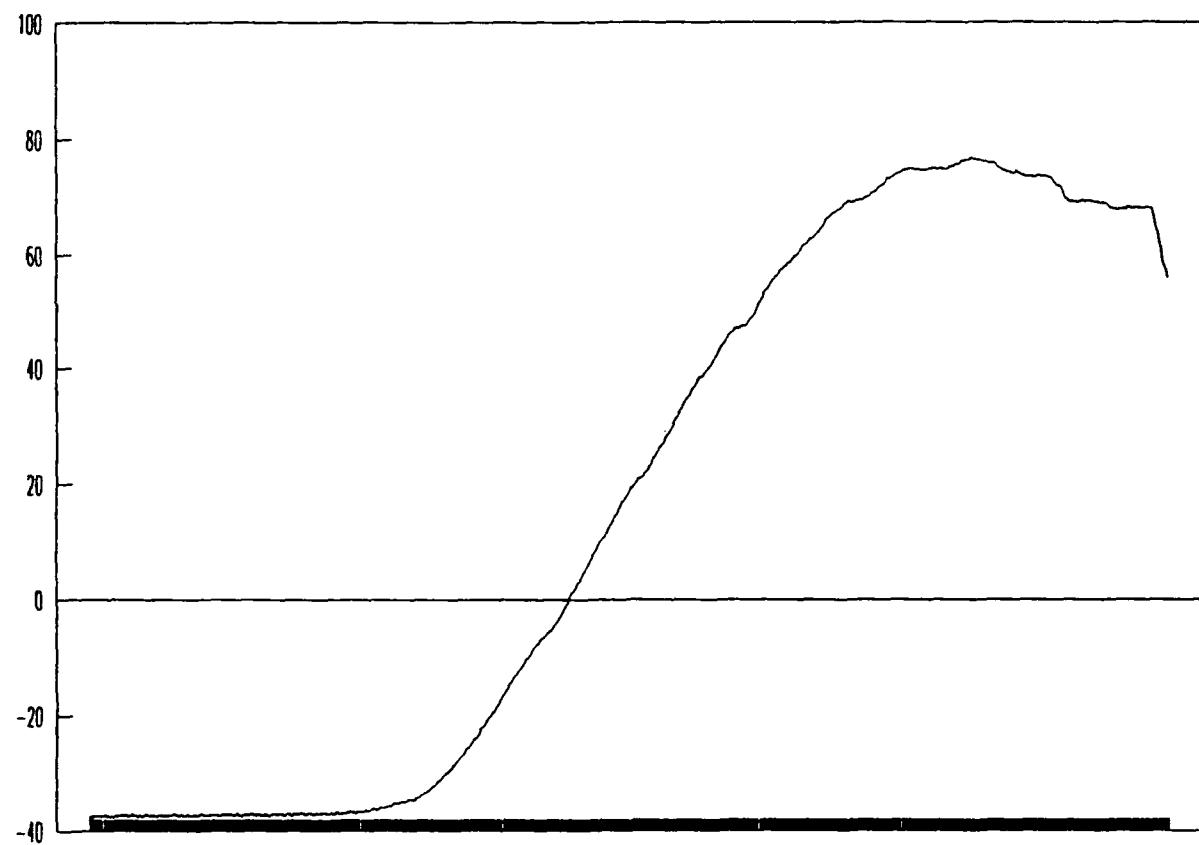
Laser tissue welding continues to show great promise for clinical application, and the data generated in this study provide a solid foundation for future pre-clinical and clinical investigations of this new modality.

Appendix A- Included Data

~~1.06 μm Nd:YAG kTP~~

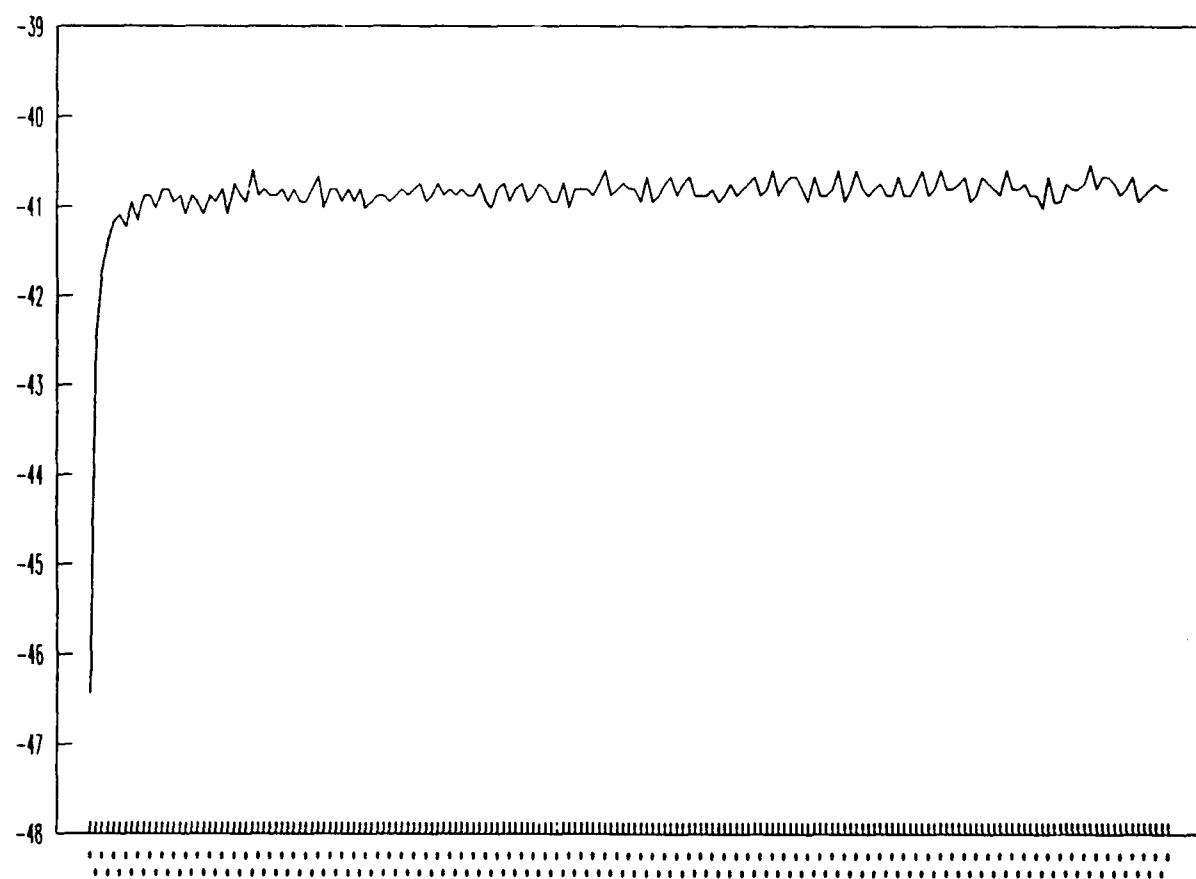
ONE STRIP UNWELDED CONTROL SEP27A

small aperture



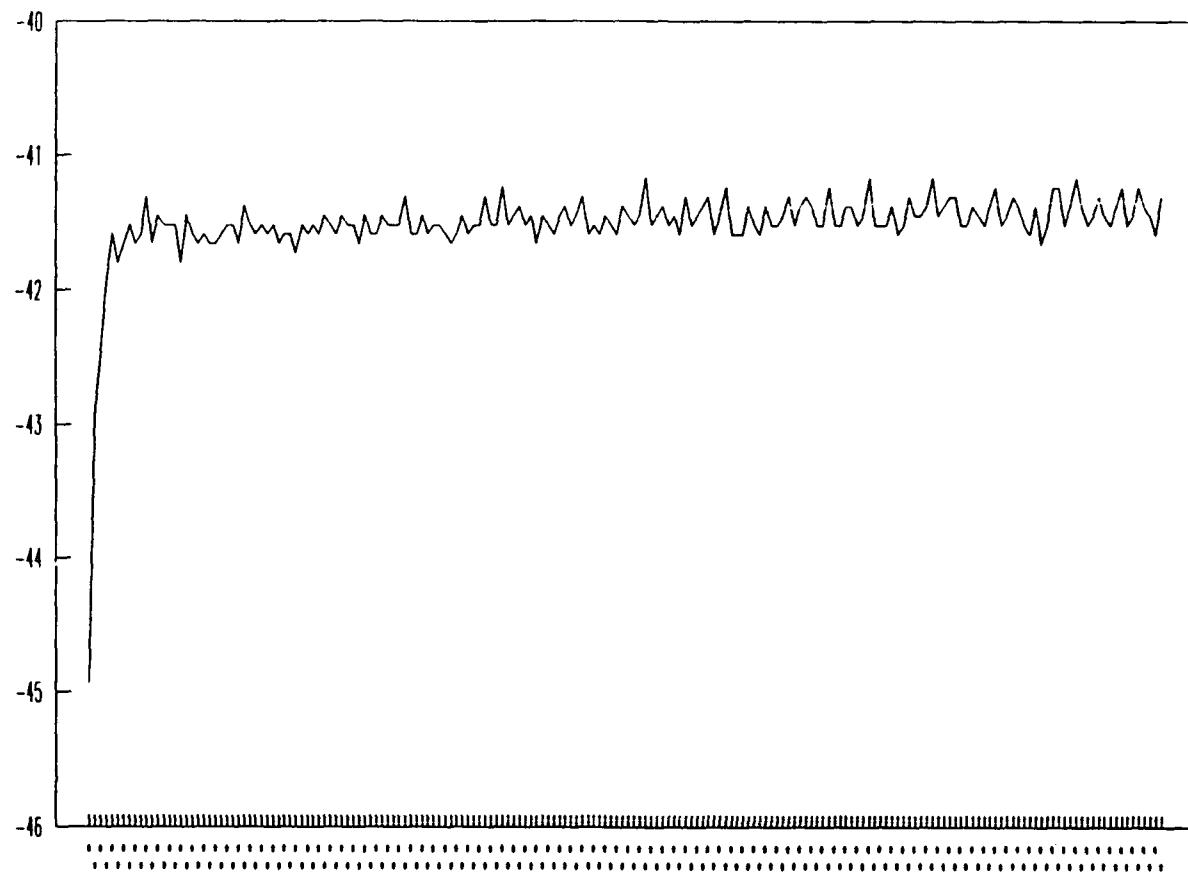
Two Strip Unwelded Control
Small aperture

SEP27B



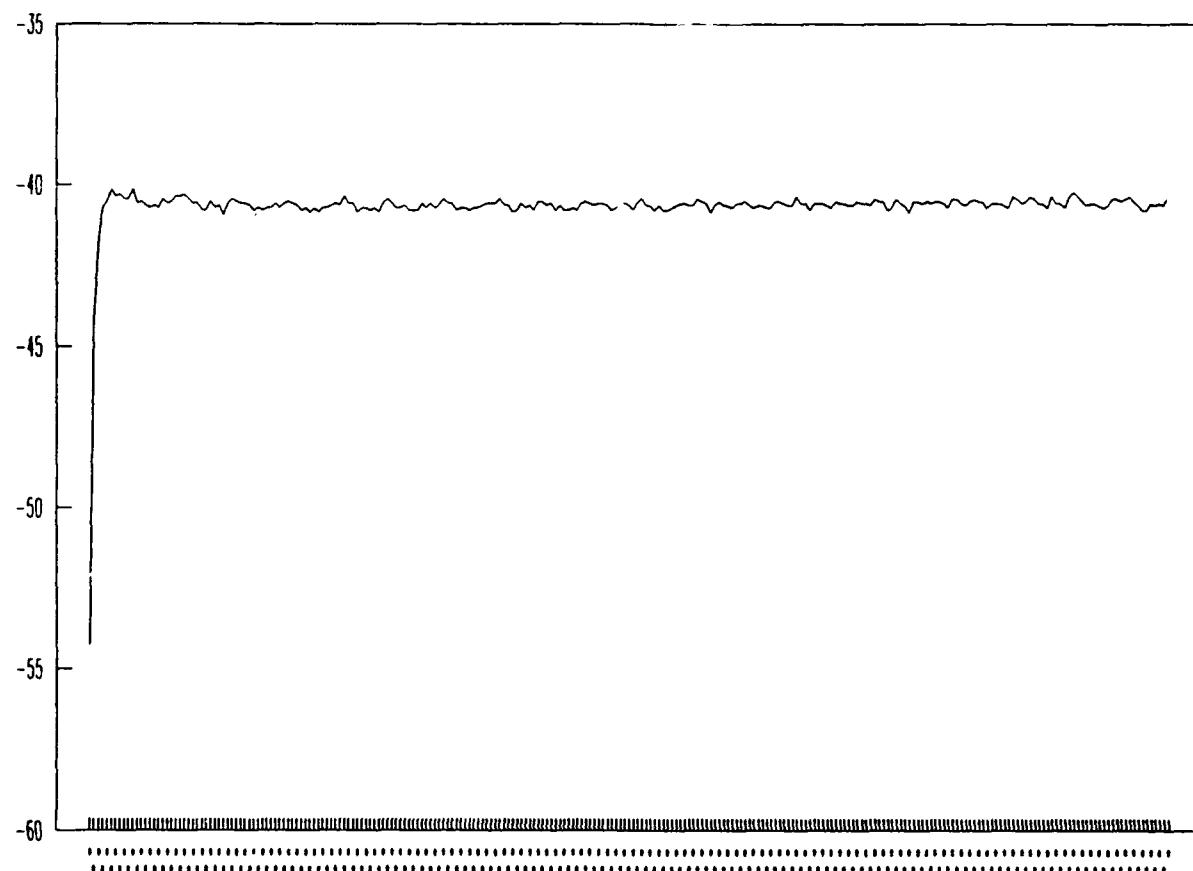
~~DATA~~ TWO STRIP INDIA IN 5 UNWELDED
small aperture CONTROL

SEP 27C



~~10.6 μm Nd:YAG~~ KTP
small aperture

INDIA INK - CONTROLLED AT 50°C SEP 27 D

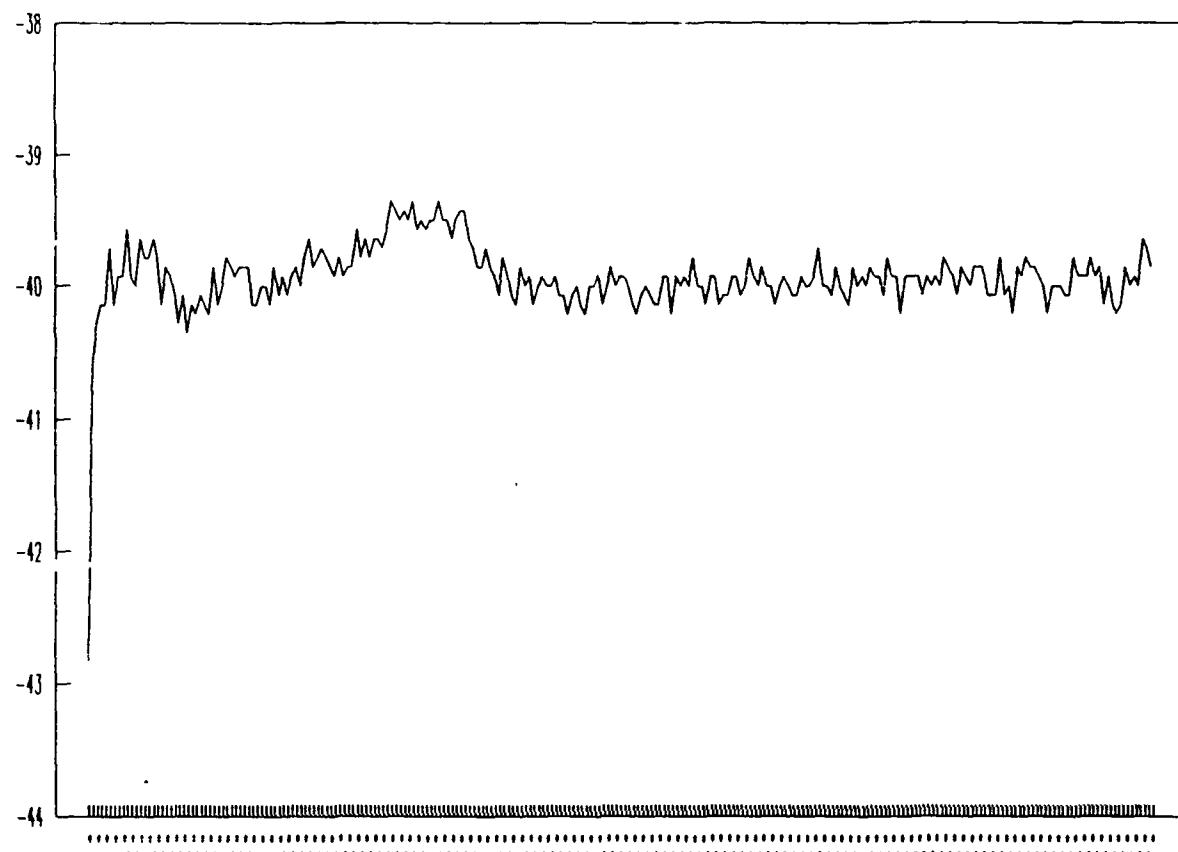


~~1064nm Nd:YAG KTP~~

small aperture

INDIA INK - CONTROLLED AT 70°C

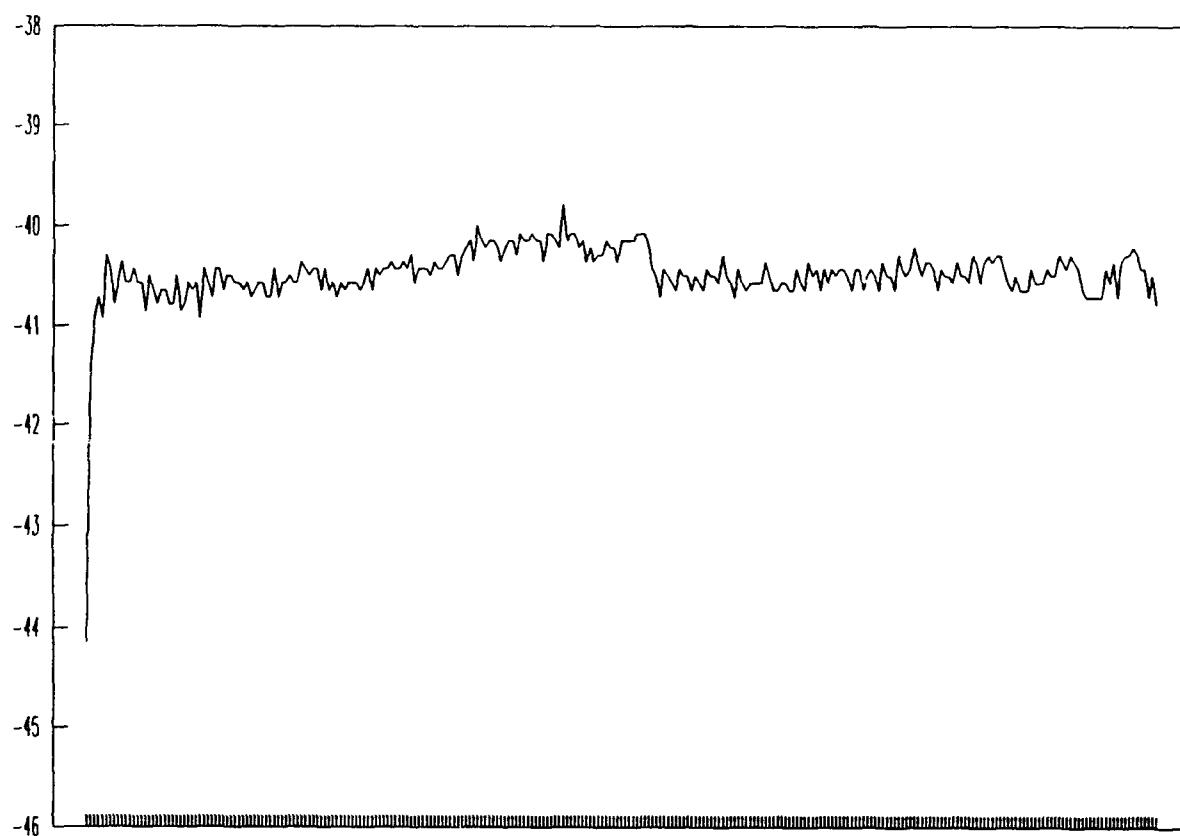
SEP 27F



~~56 μm ND: YAG K1~~
Small aperture

SEP 27 G

INDIA INK CONTROLLED AT 80°C

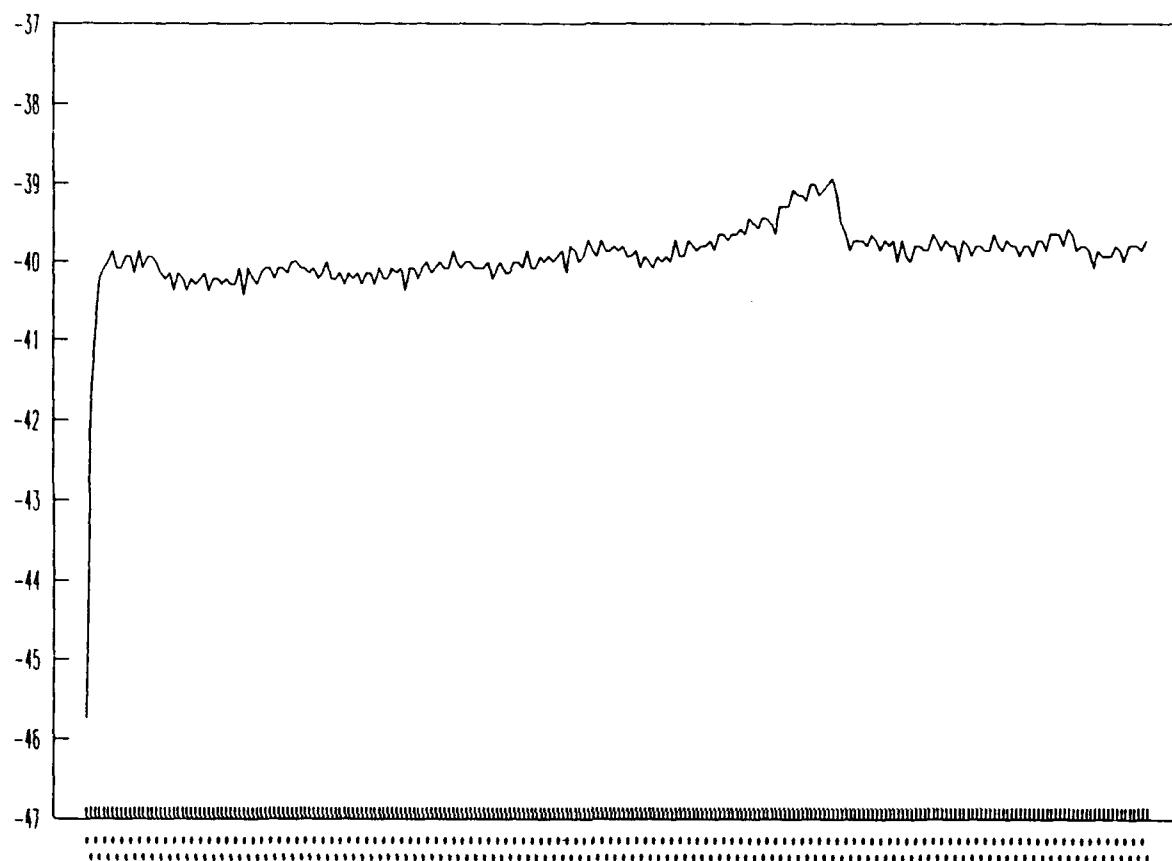


~~System Native~~ KTP

Small aperture

INDIA INK CONTROLLED AT 100°C

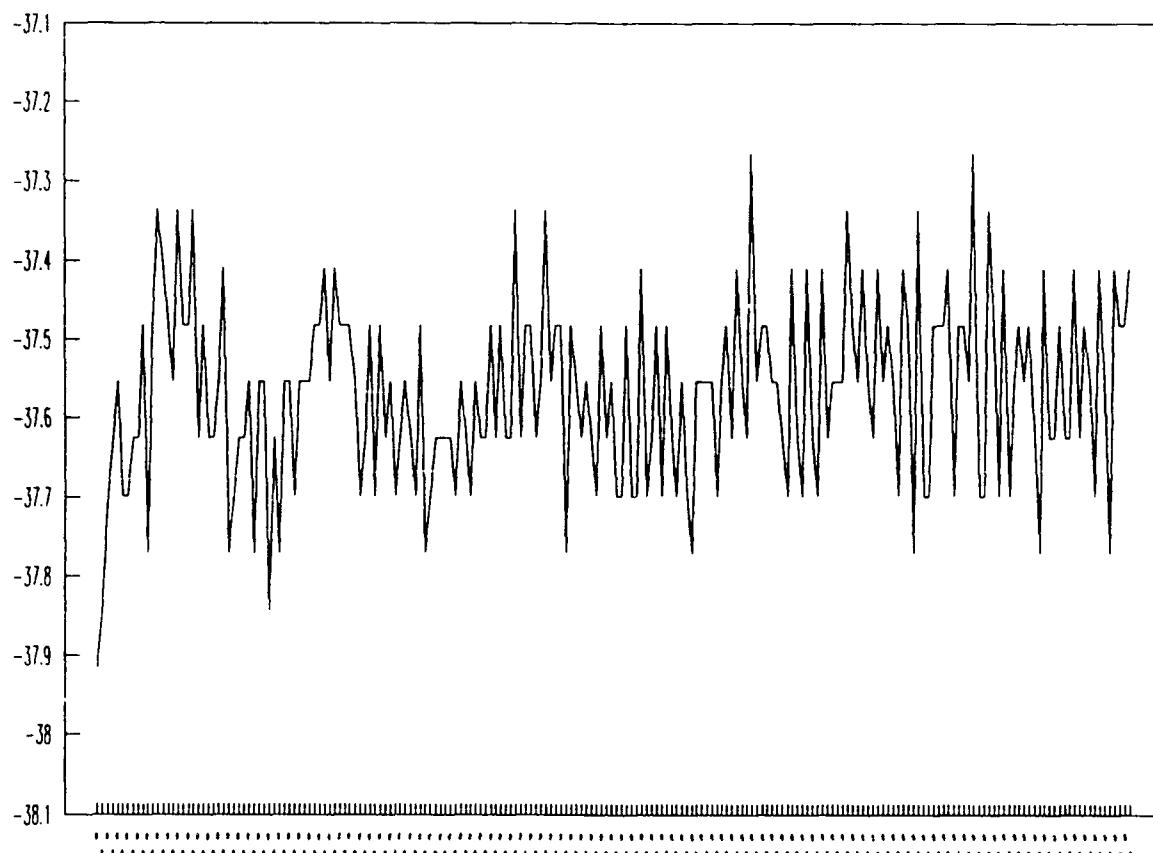
SCP 27 H



~~66pm Nd:YAG~~ KTP
Small aperture

SEP 27 0

BLOOD UNWELDED CONTROL



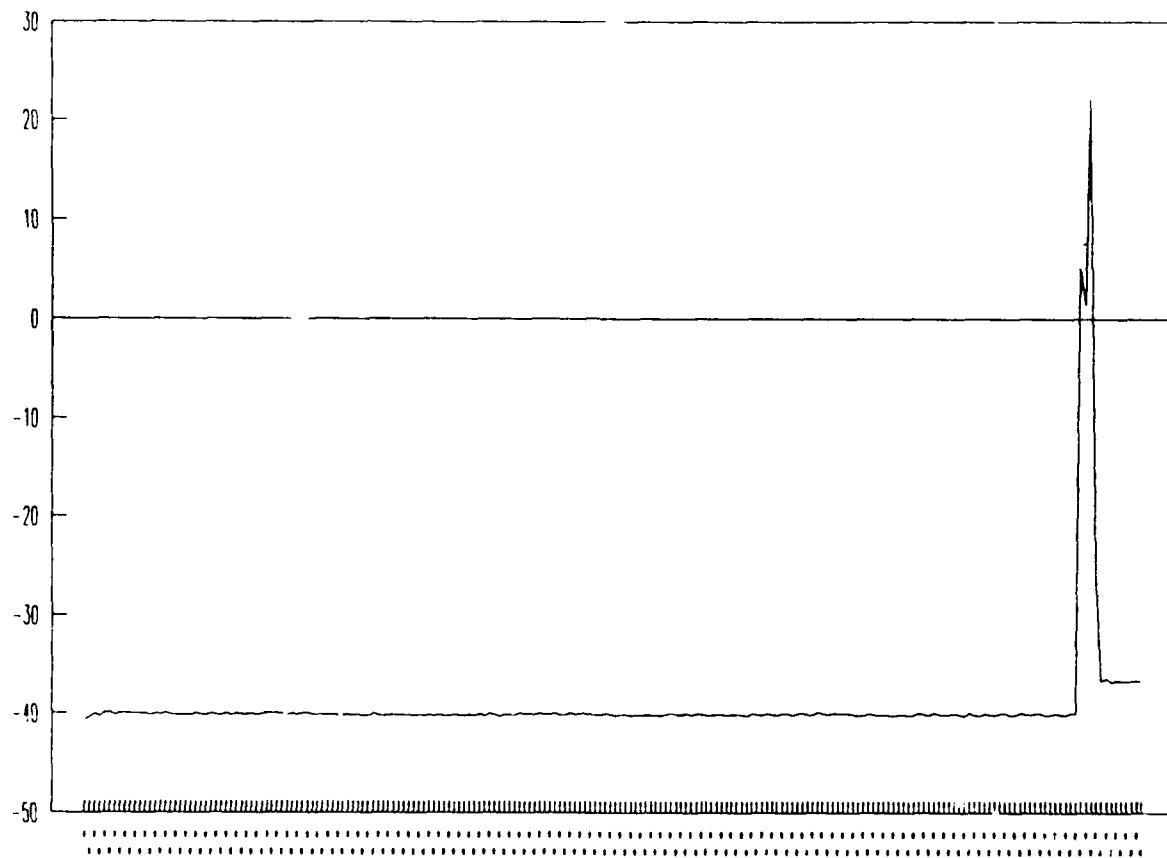
~~136 μm ND YAG KTP~~

small aperture

SEP 277

BLOOD CONTROLLED AT 50°

(touched table at end of run)

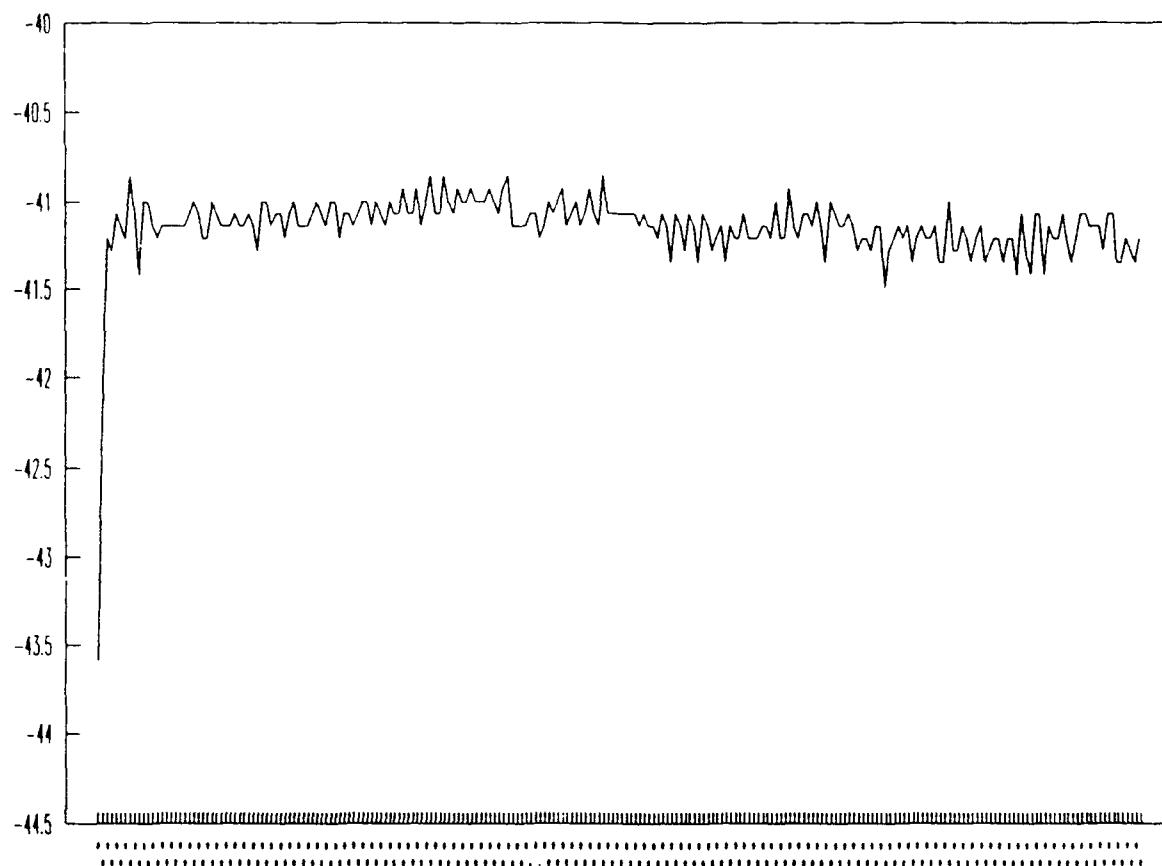


1064nm Nd:YAG KTP

small aperture

SEP 27Q

BLOOD CONTROLLED AT 60°C

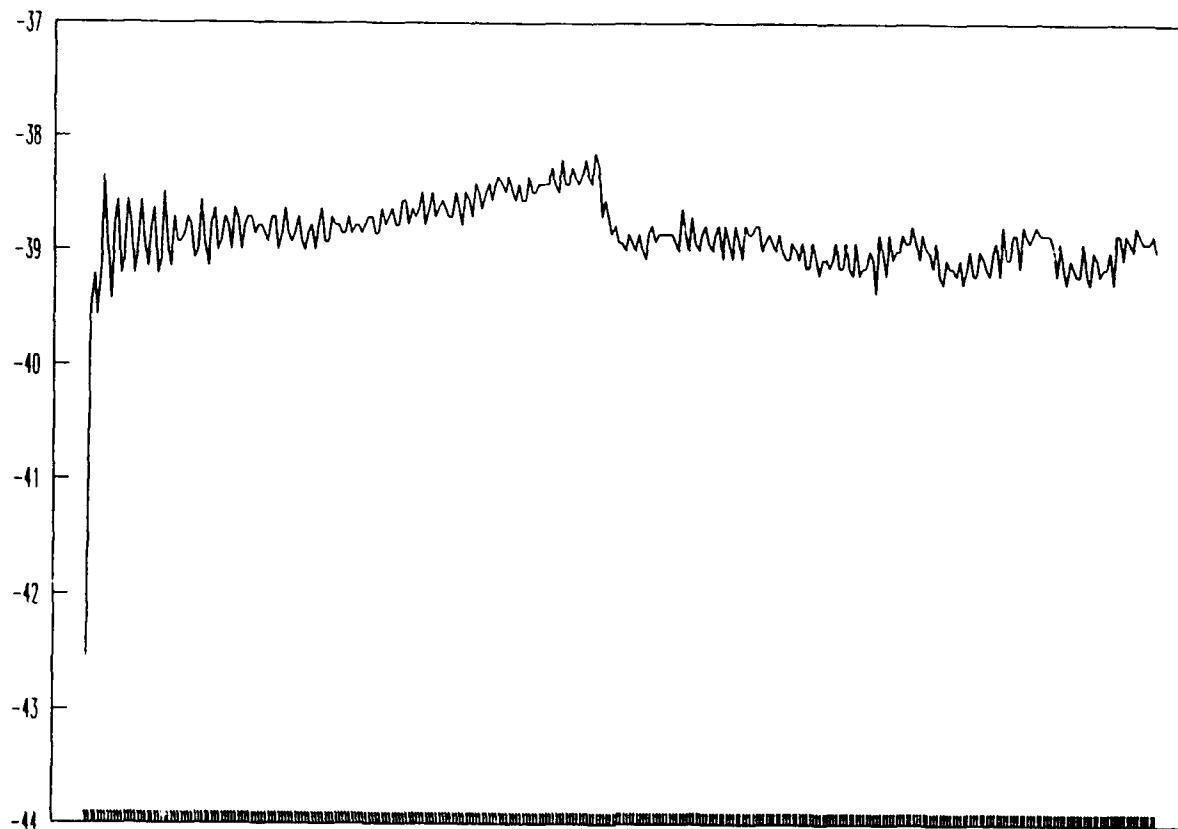


~~0.6 μm Nd-YAG KTP~~

Small aperture

BLOOD CONTROLLED AT 70°C

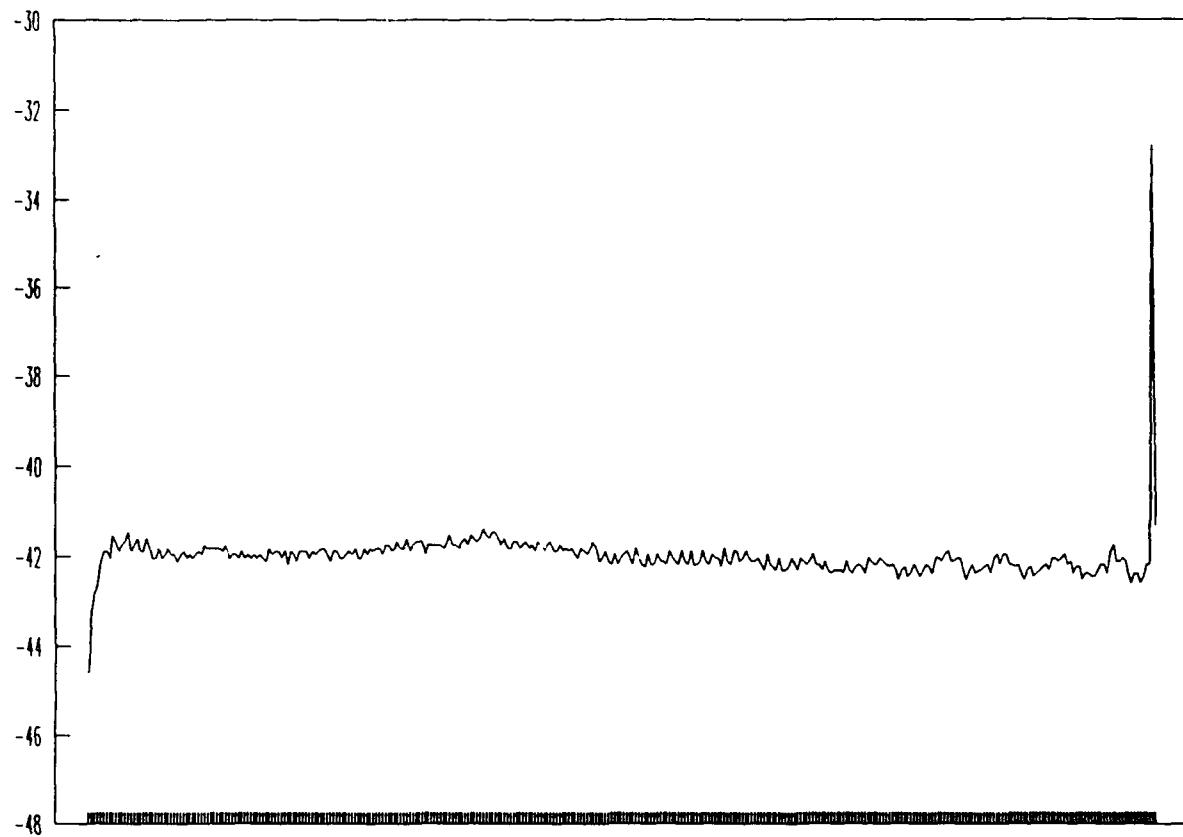
SEP 27 R



1064nm Nd:YAG KTP
small aperture

BLUOD CONTROLLED AT 80°

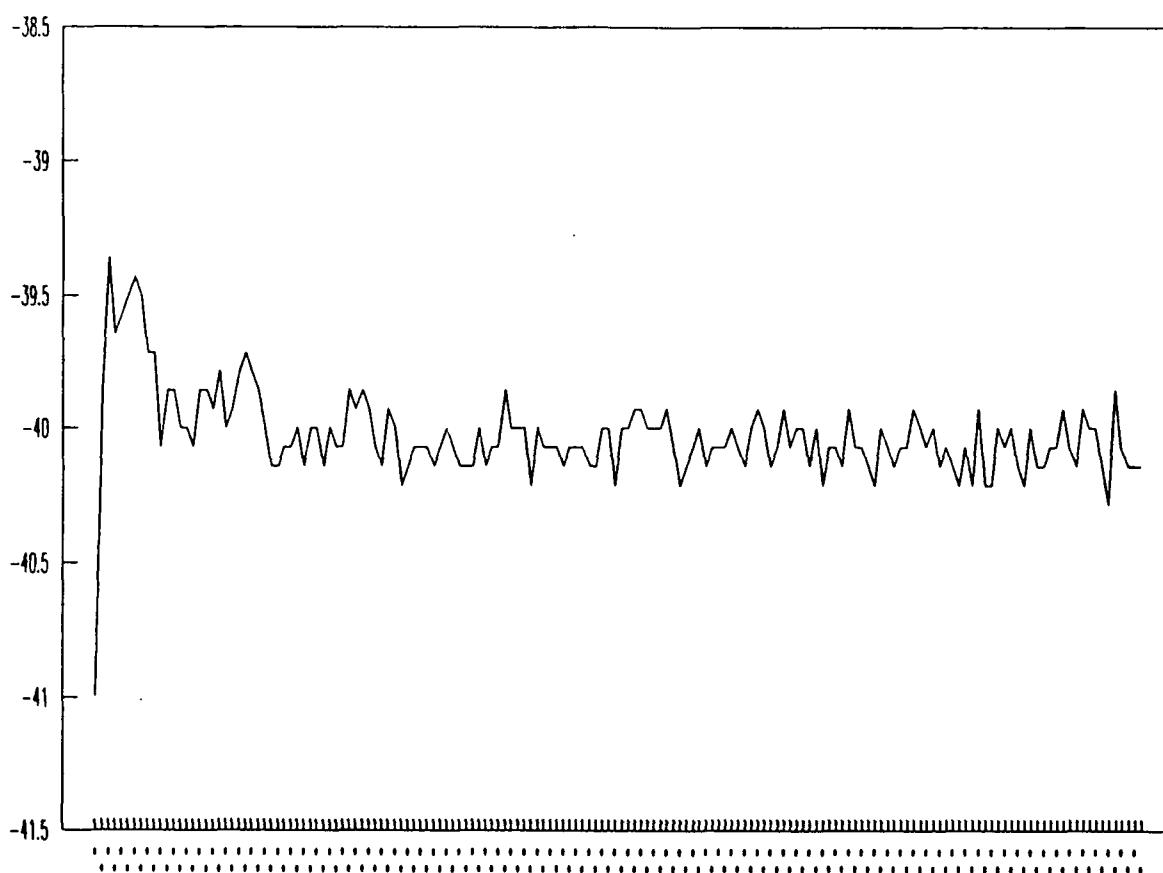
SEP 275



~~0.6mm Na+ 1A6~~ KTP
small aperture

BLOOD CONTROLLED AT 100°C

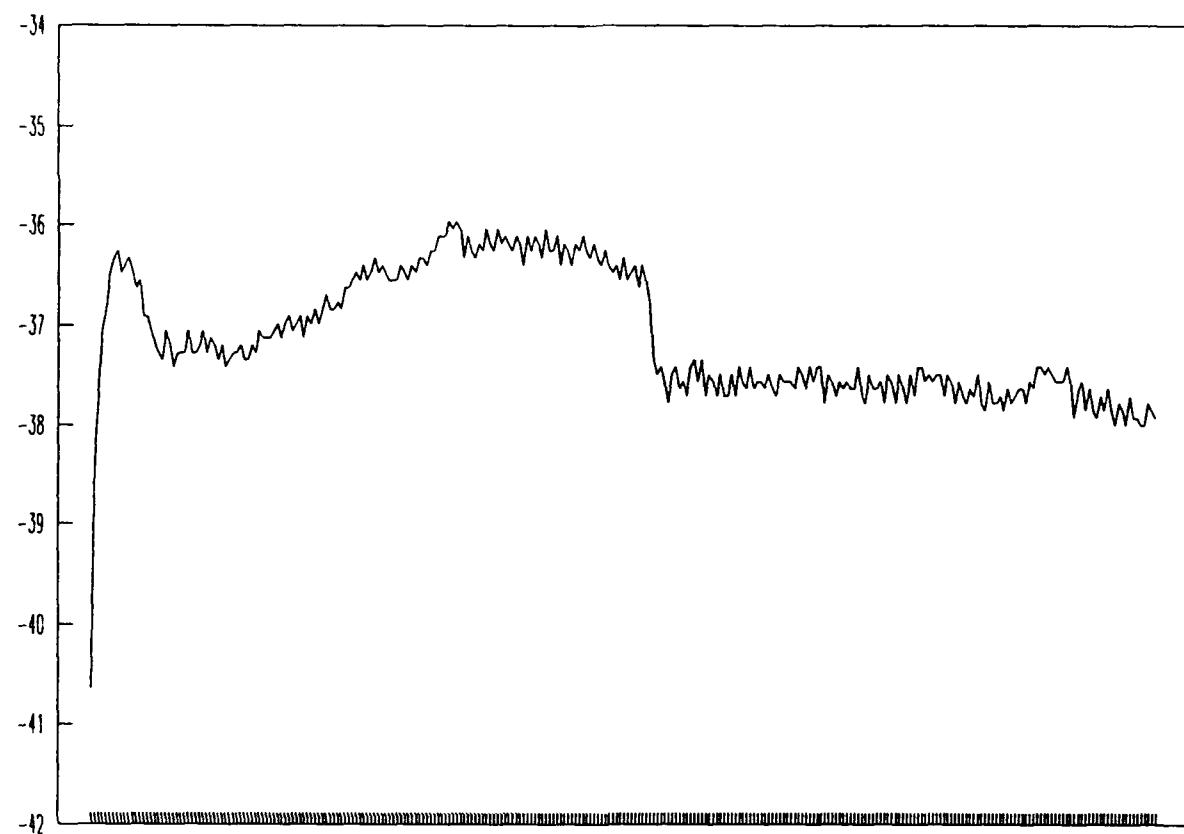
SEP 27T



~~160 μm ND VAG~~ KTP
small aperture

SEP 27U

BLOOD CONTROLLED AT 100°C

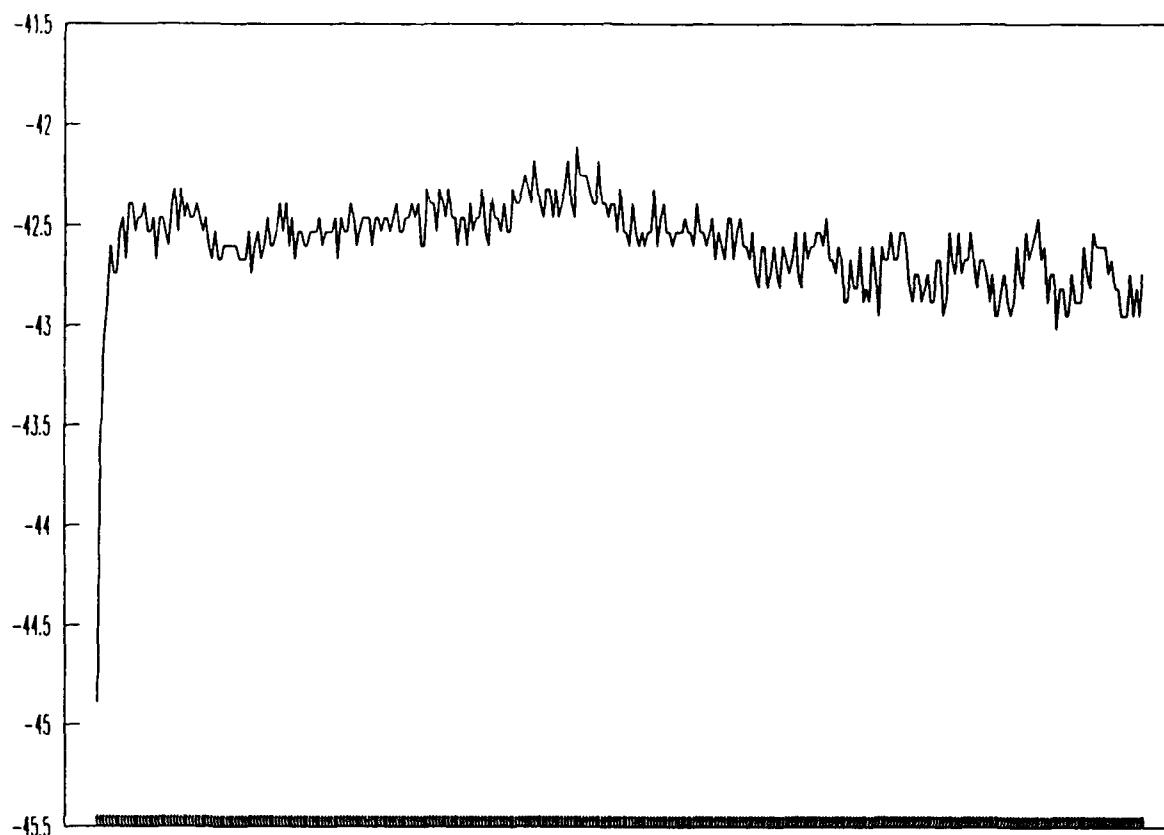


~~H bottom Not YAF~~ KTP

SEP 27 V

small aperture

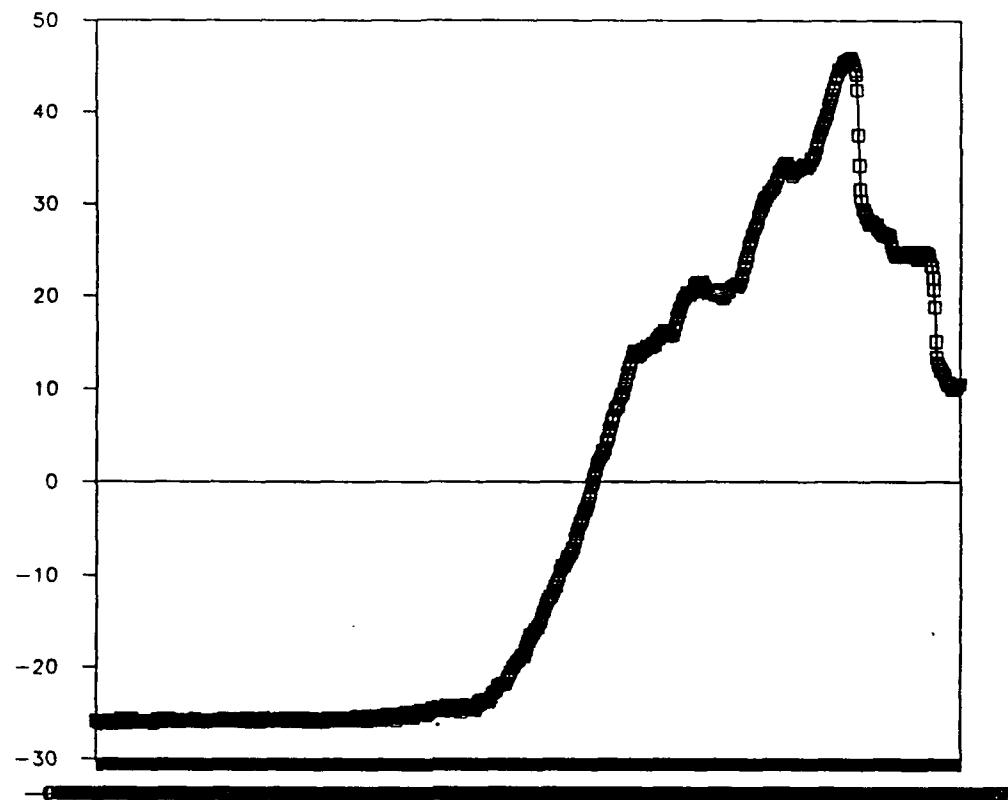
BLOOD CONTROLLED AT 100°C



.06 μm Nd:VAG
small aperture

ONE STRIP
~~TEST~~ UNWELDED CONTROL

SEP 28 I



06μm Nd:YAG
Small aperture

ONE STRIP
~~TCG~~ UNWELDED CONTROL

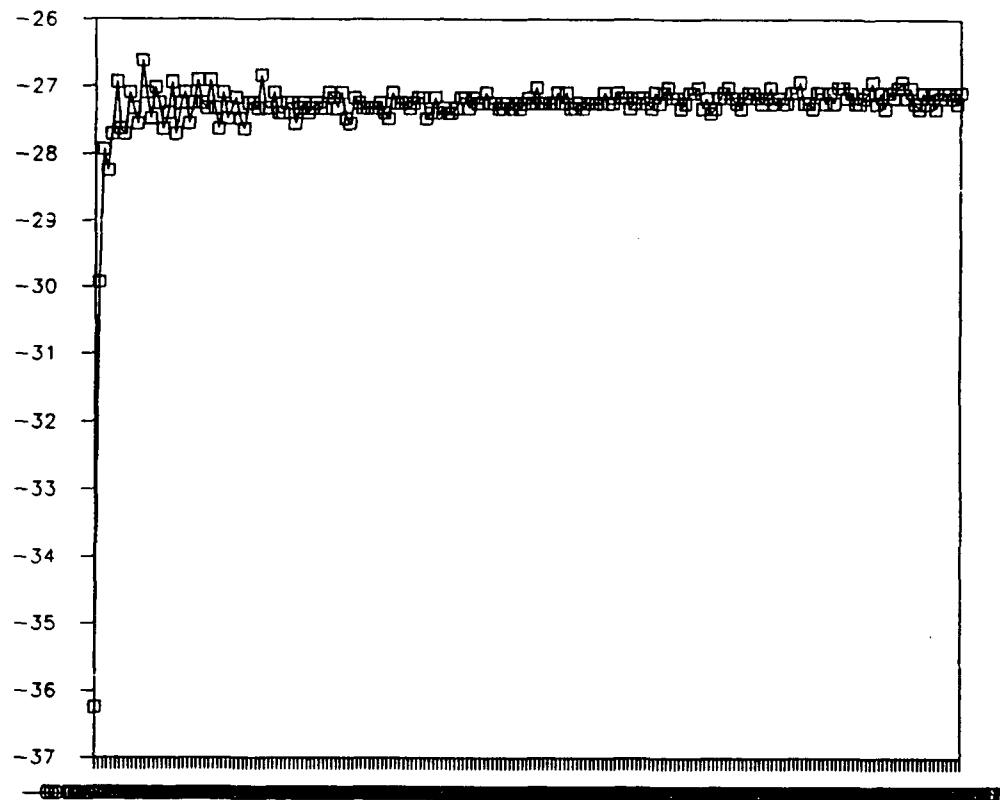
SEP 28 J



0.6 μm Nd:YAG
Small aperture

TWO STRIP UNWELDED CONTROL

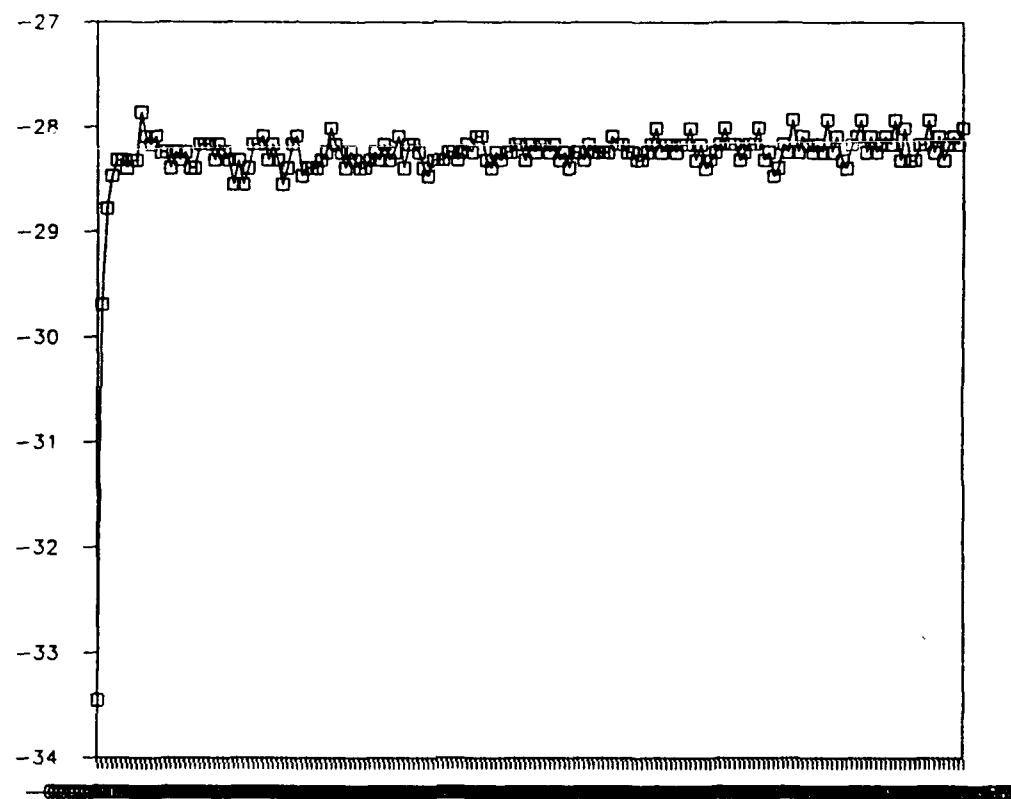
SEP 28 K



0.6μm Nd:YAG
small aperture

TWO STRIPS ICG UNWELDED CONTROL

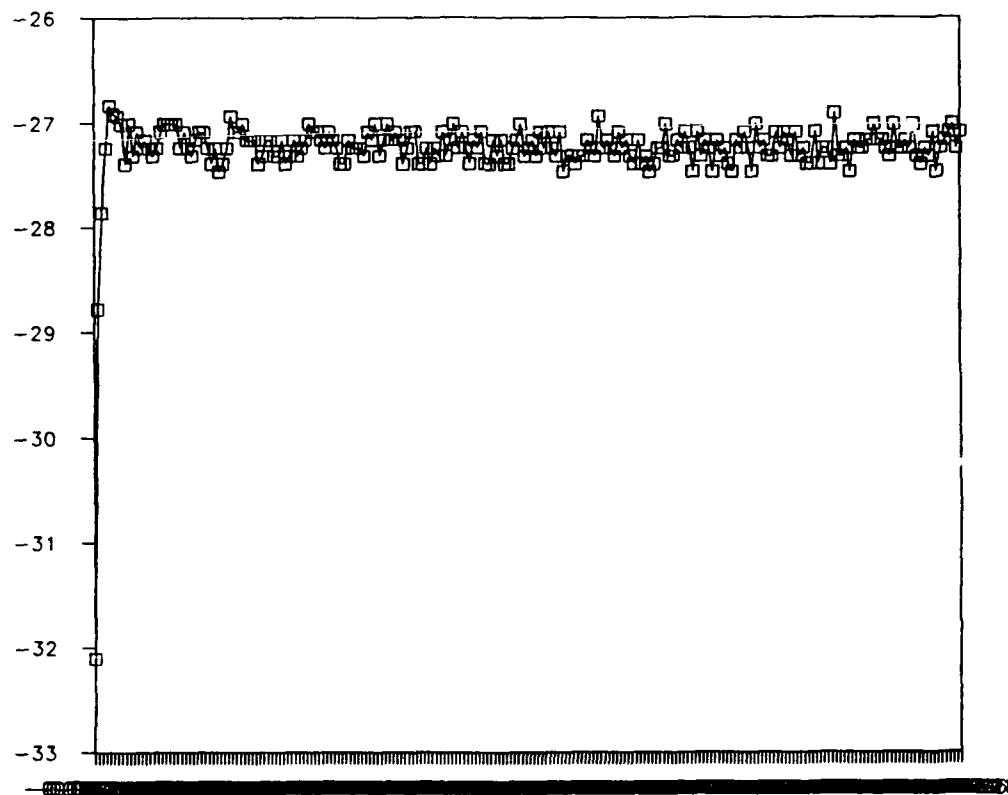
SEP 28 L



$0.6 \mu\text{m}$ Nd:YAG
small aperture

SEP 28 M

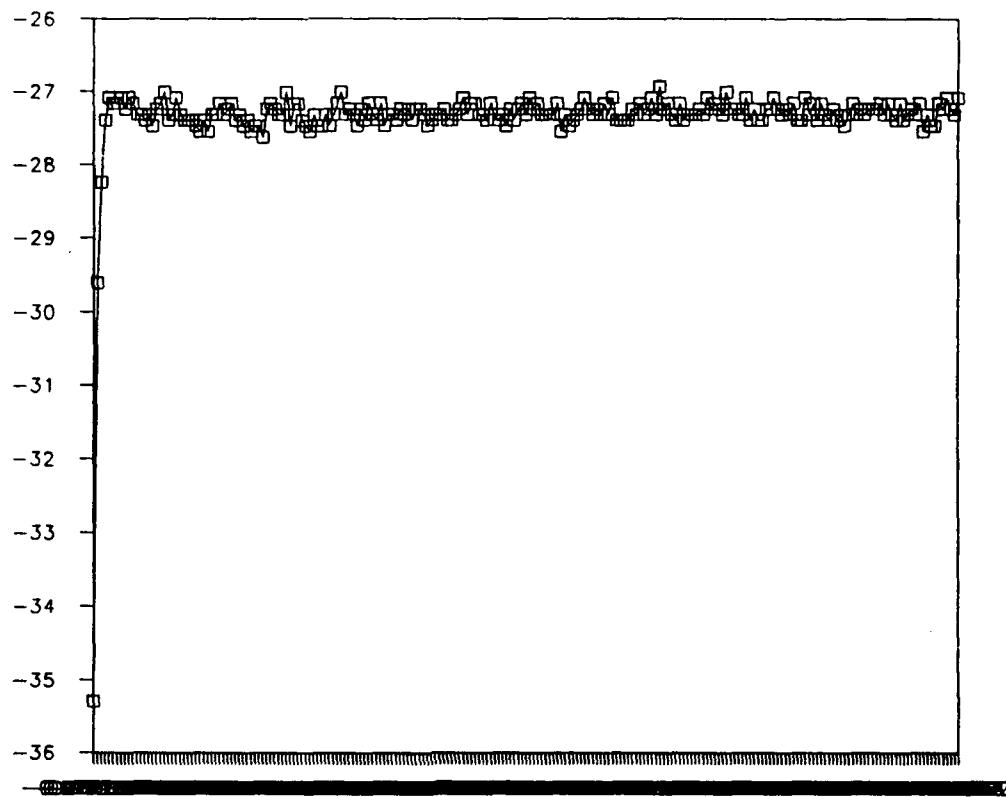
ICG CONTROLLED AT 50°C



0.6 μm Nd:YAG
small aperture

SEP 28 N

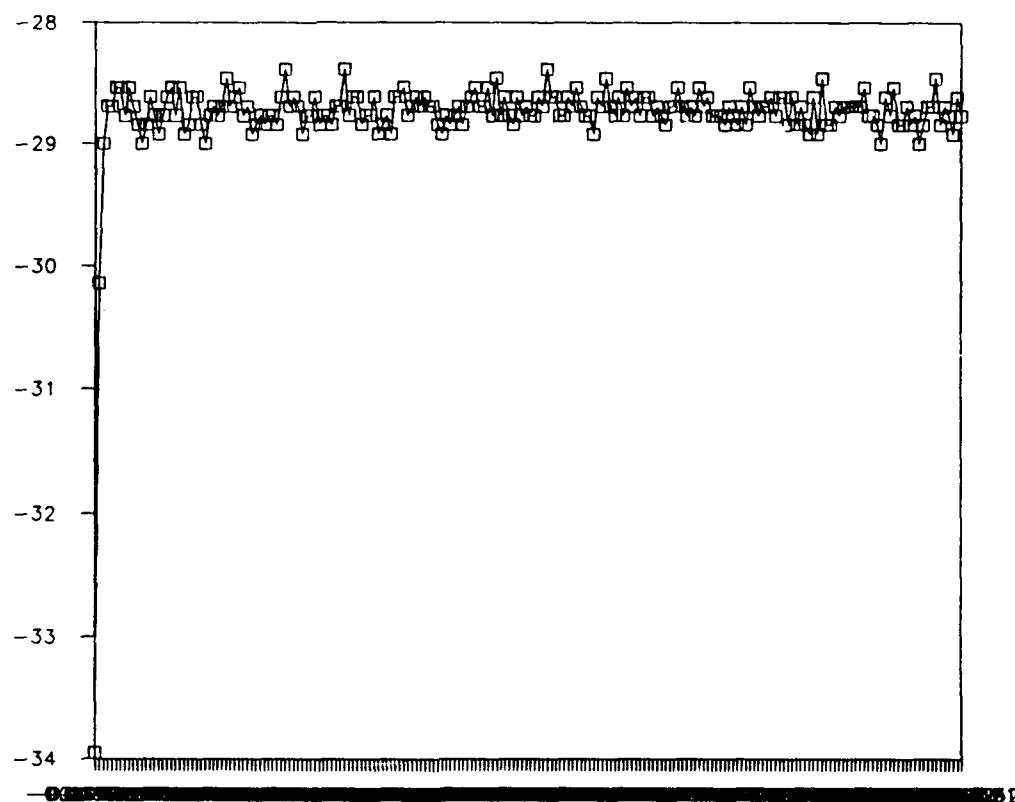
ICG CONTROLLED AT ~~50°C~~ 60°C



56 μm Nd: YAG
Small aperture

ICG CONTROLLED AT 70°C

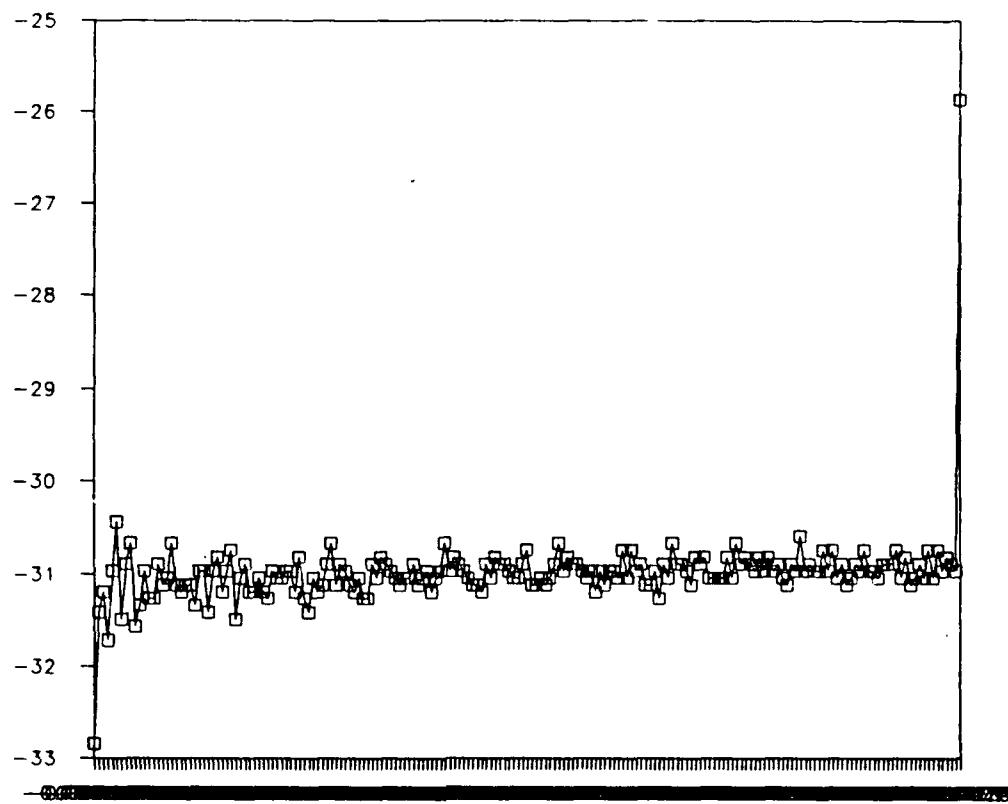
SEP 28 0



$0.6\mu\text{m}$ Nd:YAG
small aperture

ICG CONTROLLED AT 80°C

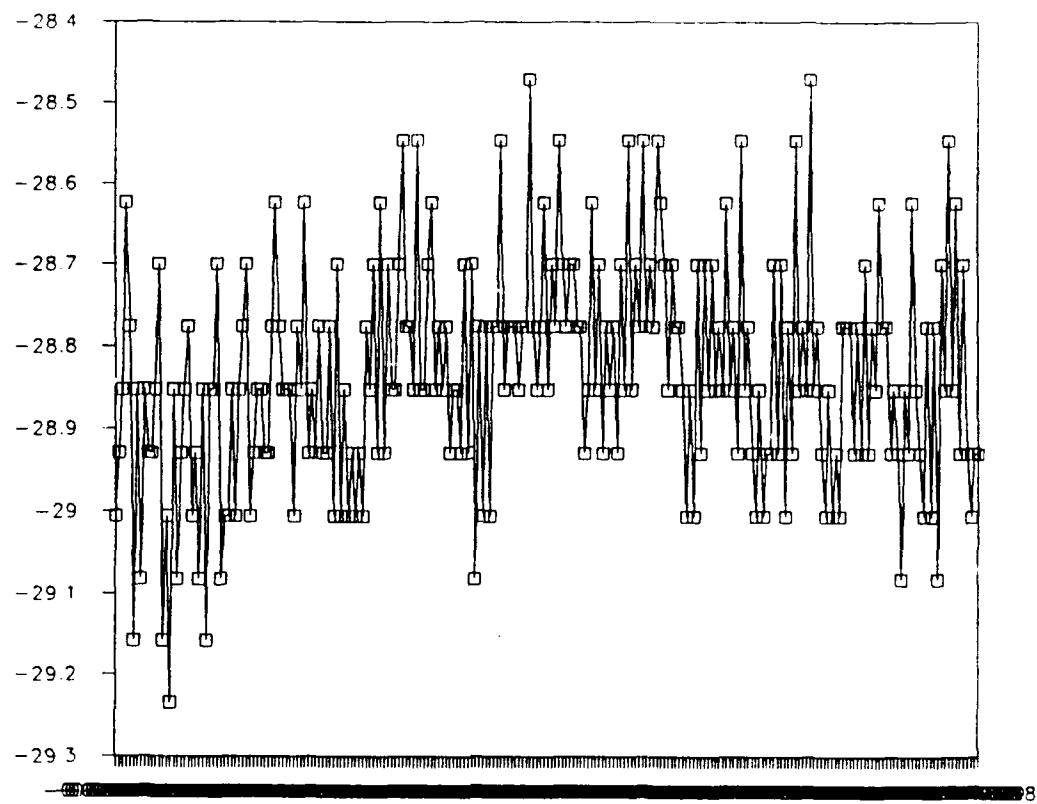
SEP 28 P



16 μ m Nd:YAG
small aperture

ICG CONTROLLED AT 1000C

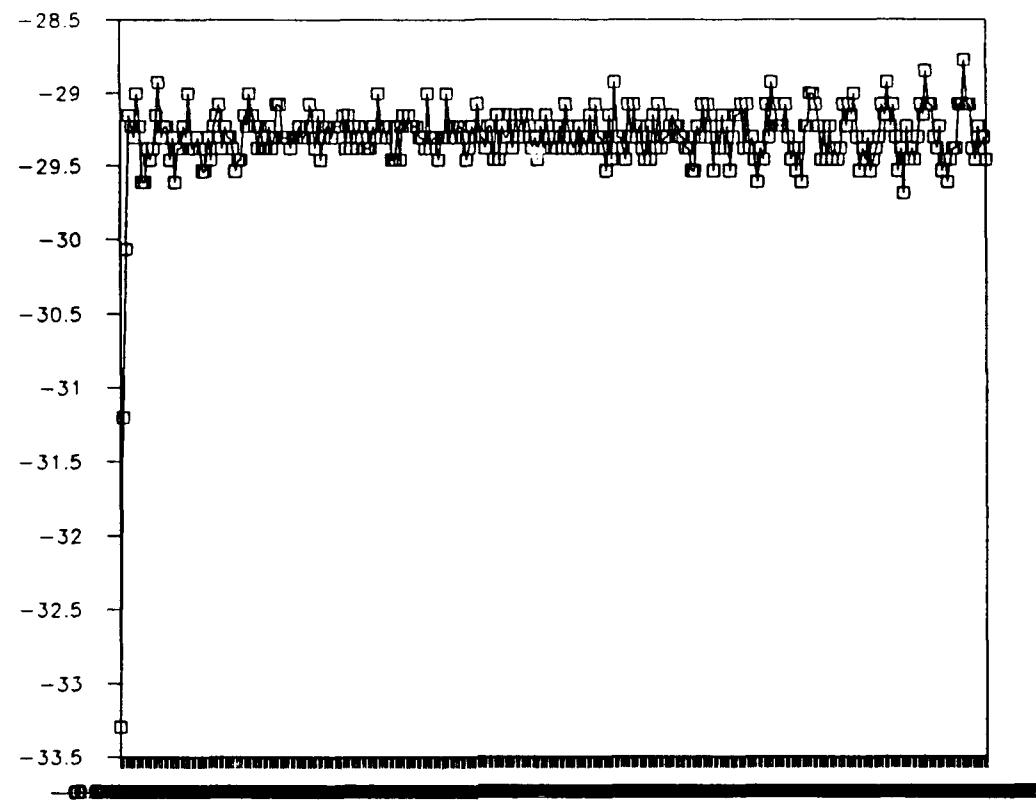
SEP 23 Q1



106 μm Nd:YAG
small aperture

ICG CONTROLLED AT 100°C

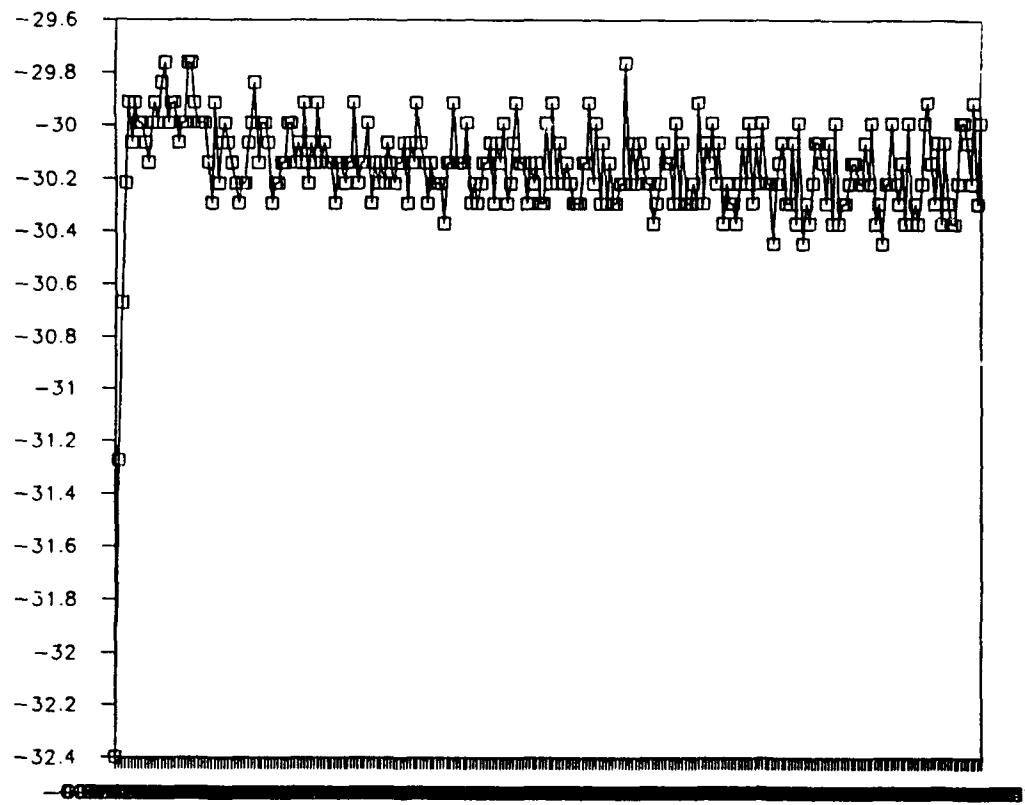
SEP 28 R



0.6 μm Nd:YAG
small aperture

SEP 28 5

INDIA INK CONTROLLED AT 80°C



$0.6\mu\text{m}$ Nd:YAG
small aperture

SEP 28 1

ONE STRIP CONTROL



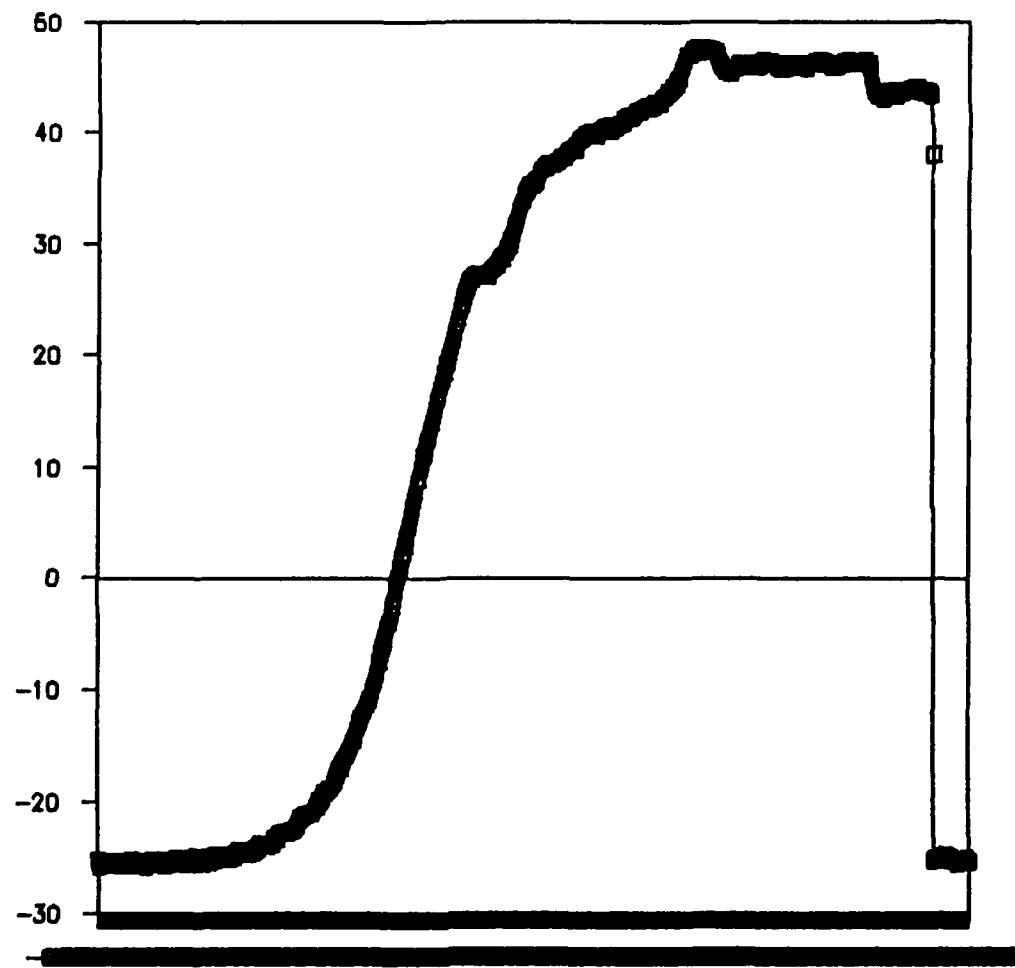
OCT 9, 1990

KTP laser → 20W out of laser → 4.0 W delivered
Small Aperture

Ferronyl - $\frac{1\text{g Ferronyl}}{3\text{ml } 0.9\% \text{ saline}}$

Ferronyl + Blood - $\frac{1\text{g Ferronyl}}{3\text{ml whole blood (in EDTA)}}$

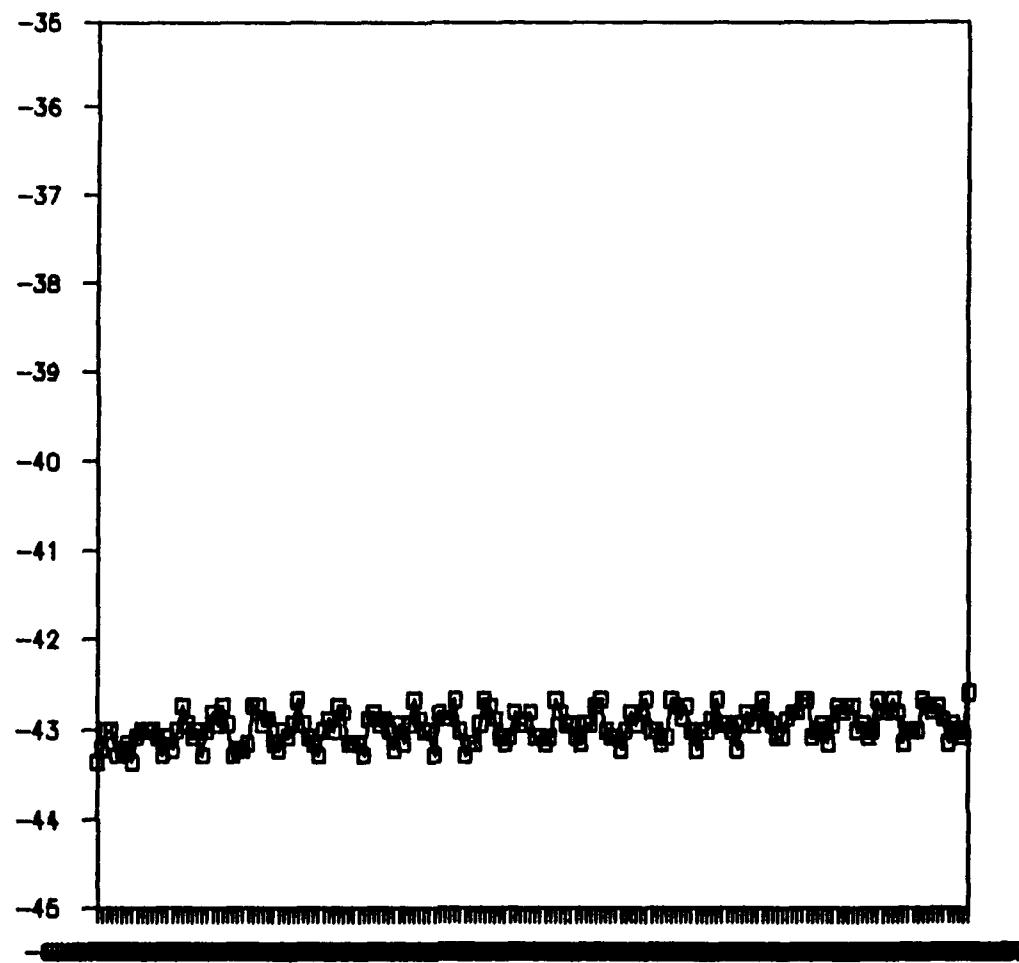
OCT9A



KTP - one step unwelded
control

small aperture

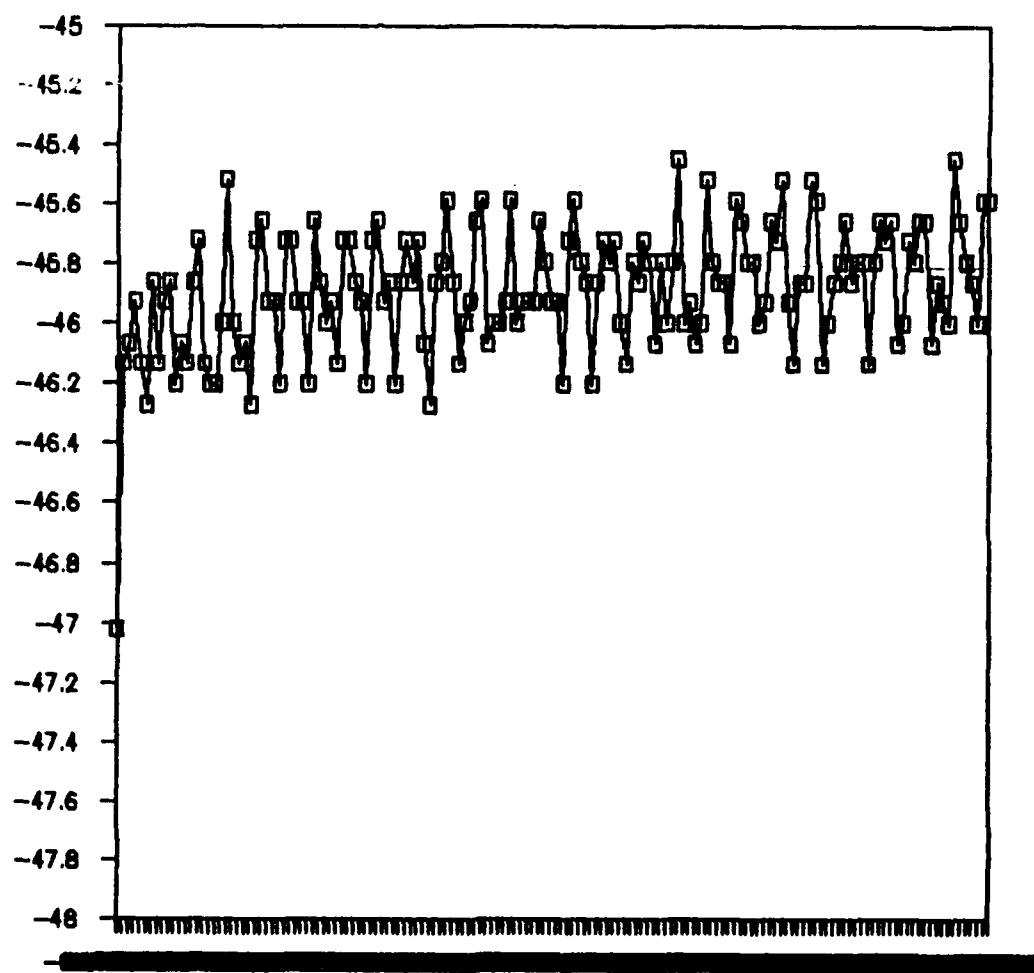
OCT9B



KTP - two strip unwelded
control

small aperture

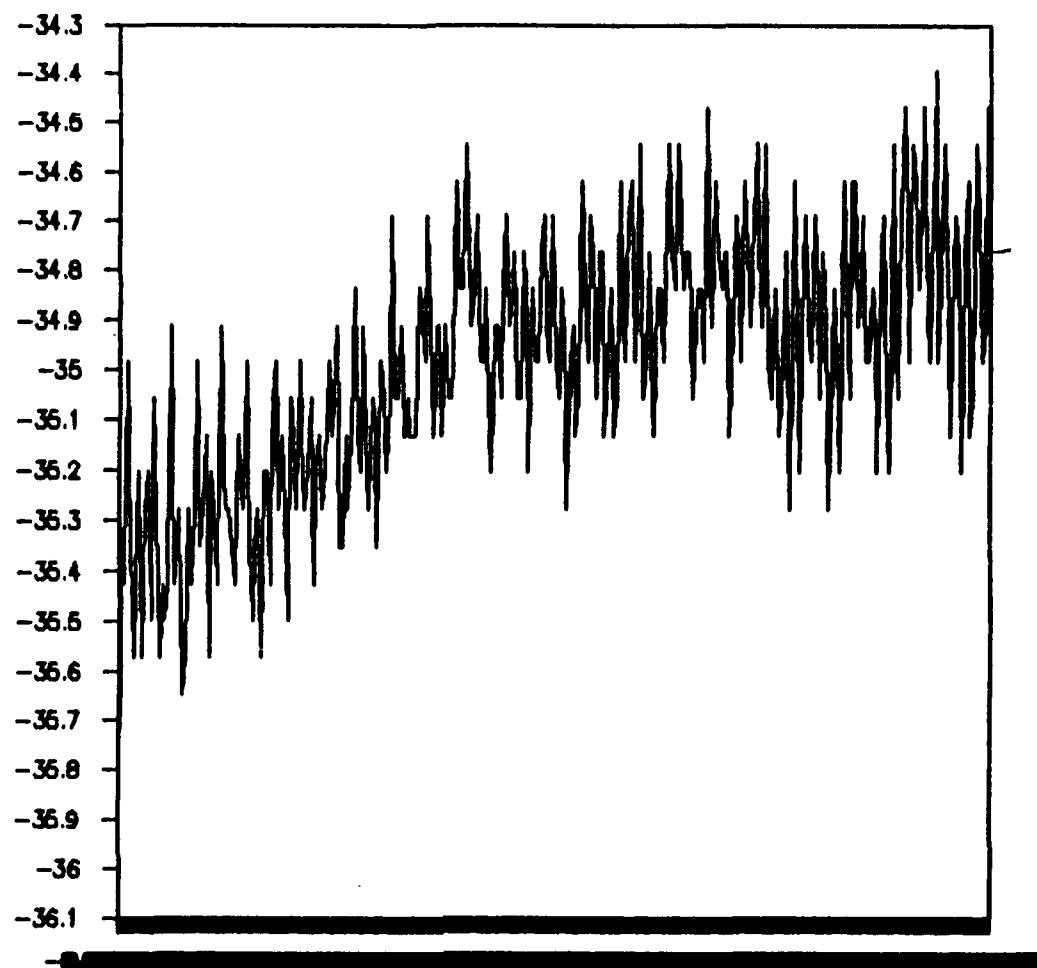
OCT9C



KTP - two step India ink
unwelded control
small aperture

OCT9D

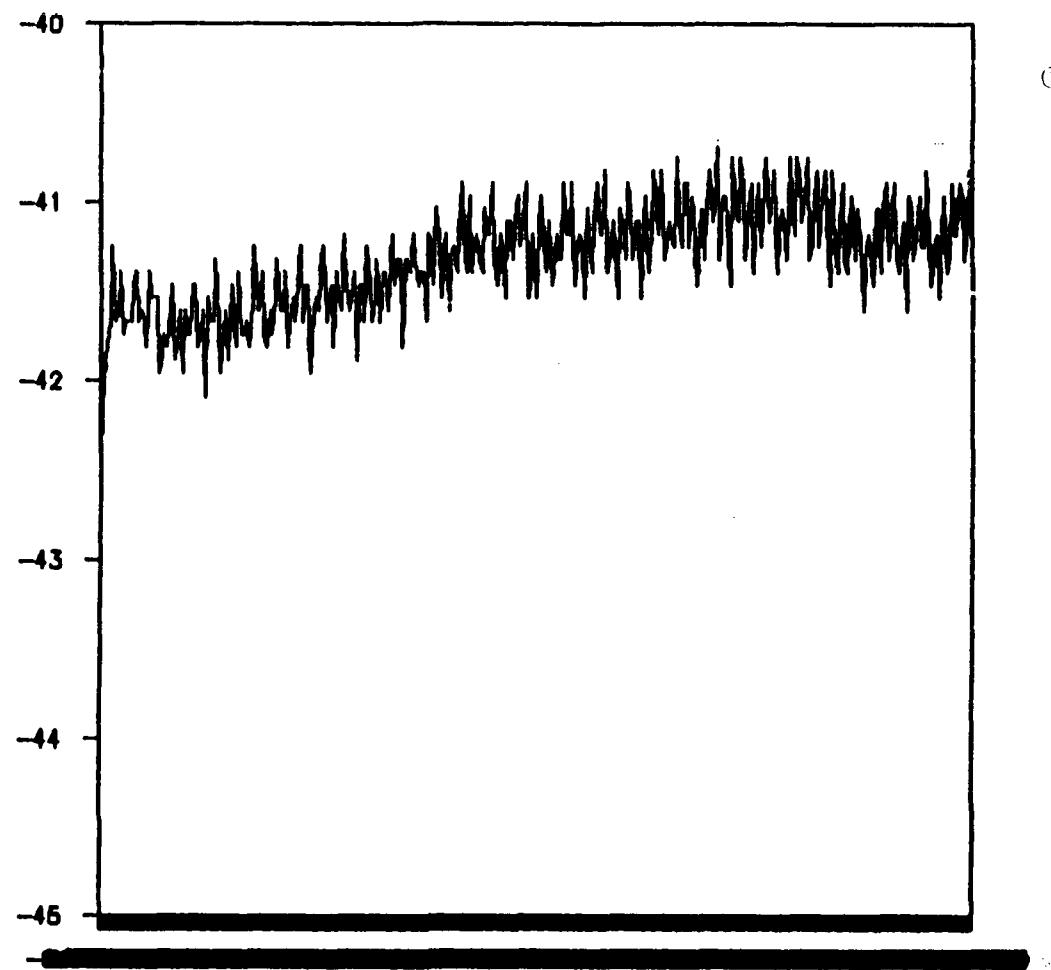
06g



KTP - Andea cut - 50°C

small aperture

OCT9E

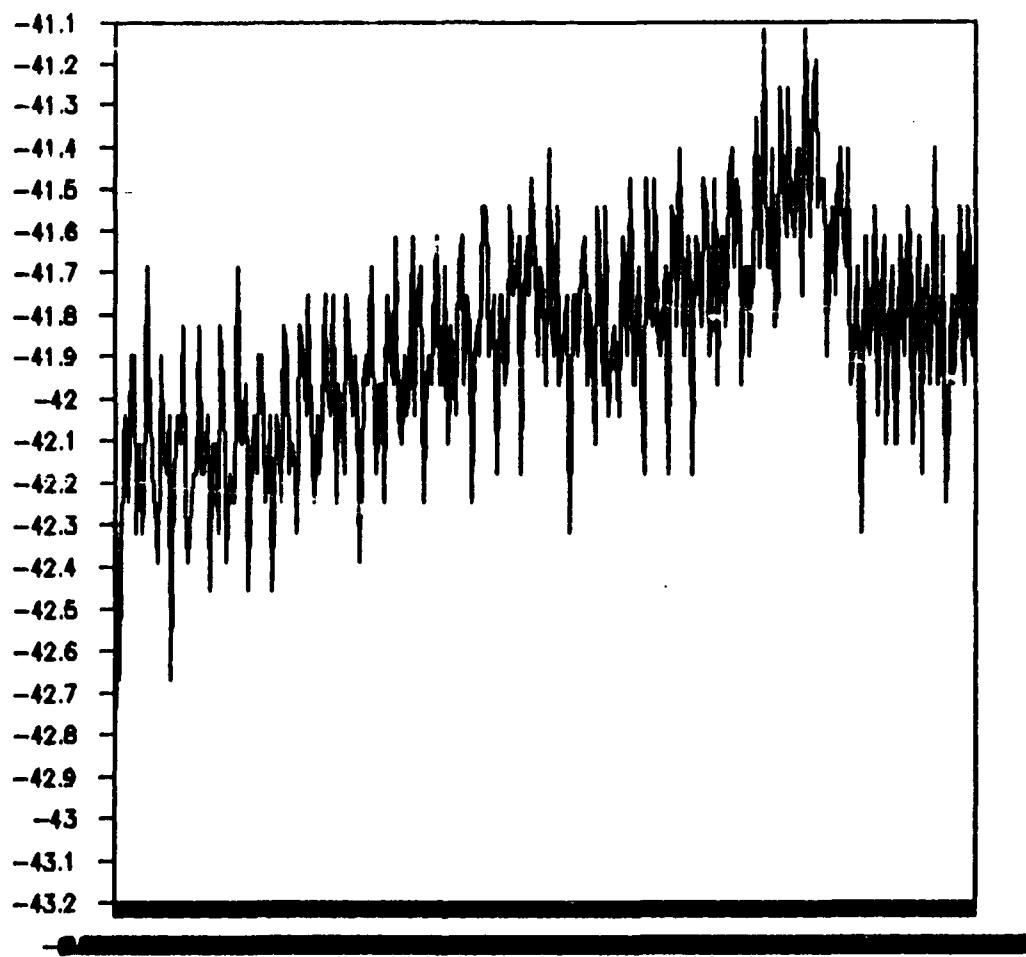


KTP - India ink - 60°C

small aperture

OCT9F

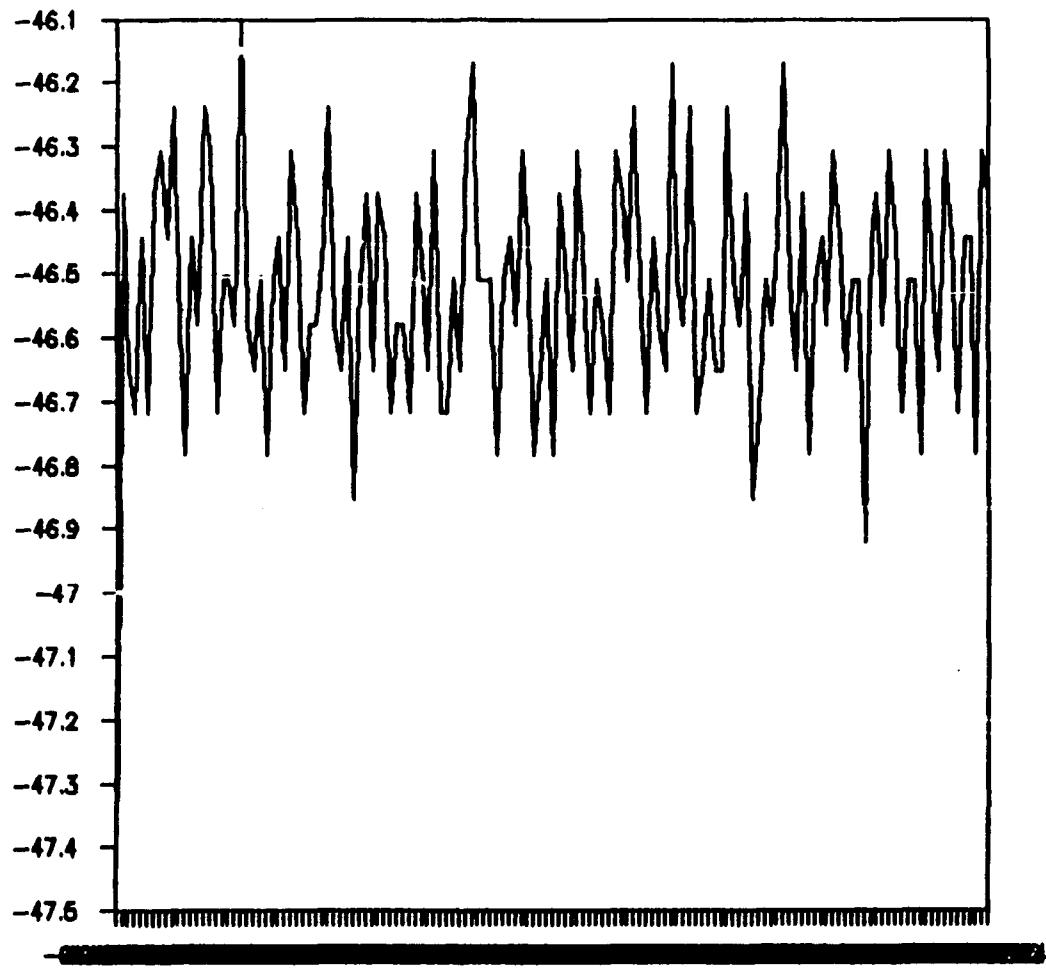
0.79



KTP- India ink - 70°C

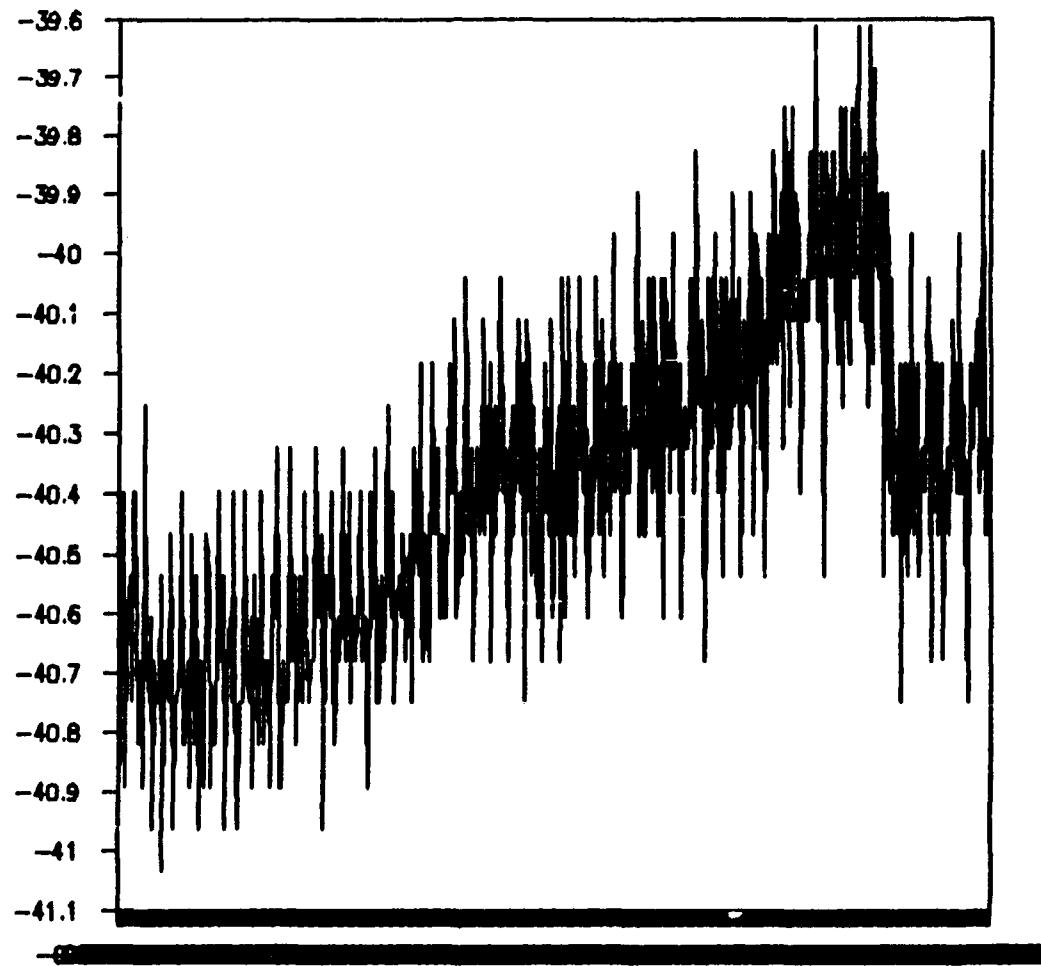
small aperture

OCT9G



KTP - India unk - 80°C
T_c too superficial ?
small aperture

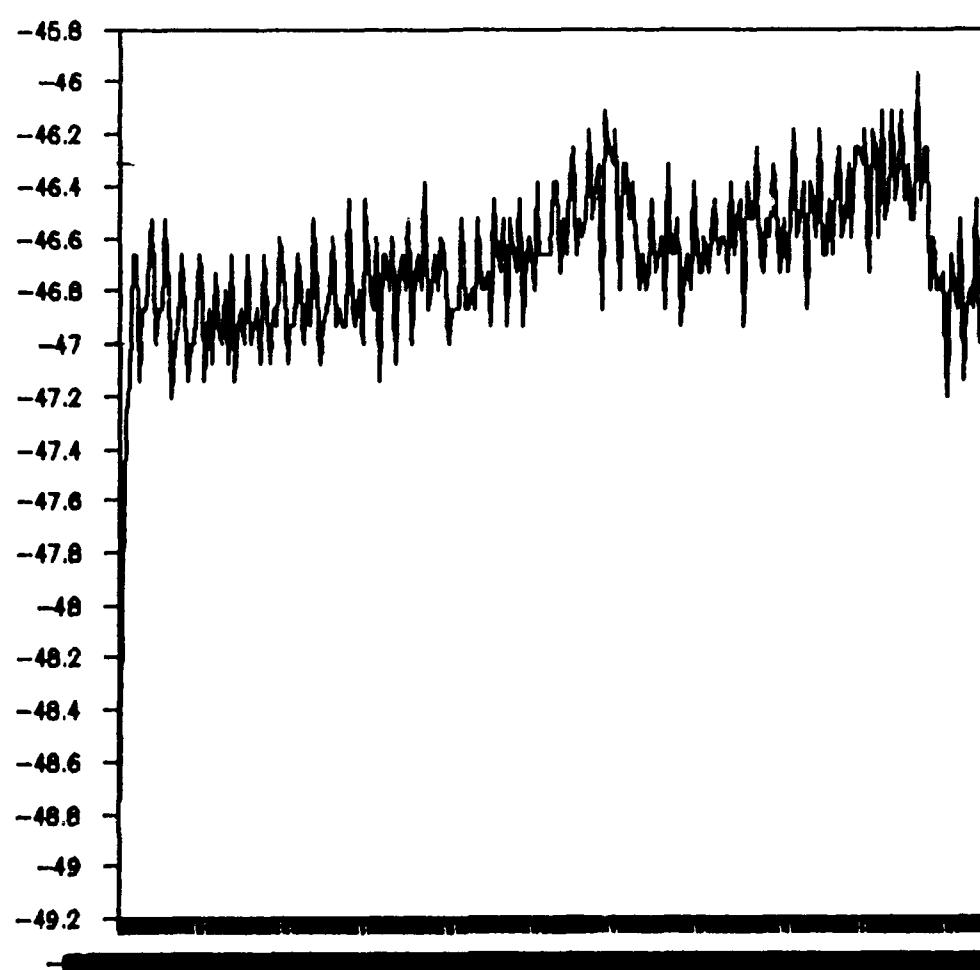
OCT9H



KTP - Nd:YAG - 80°C

small aperture

OCT91

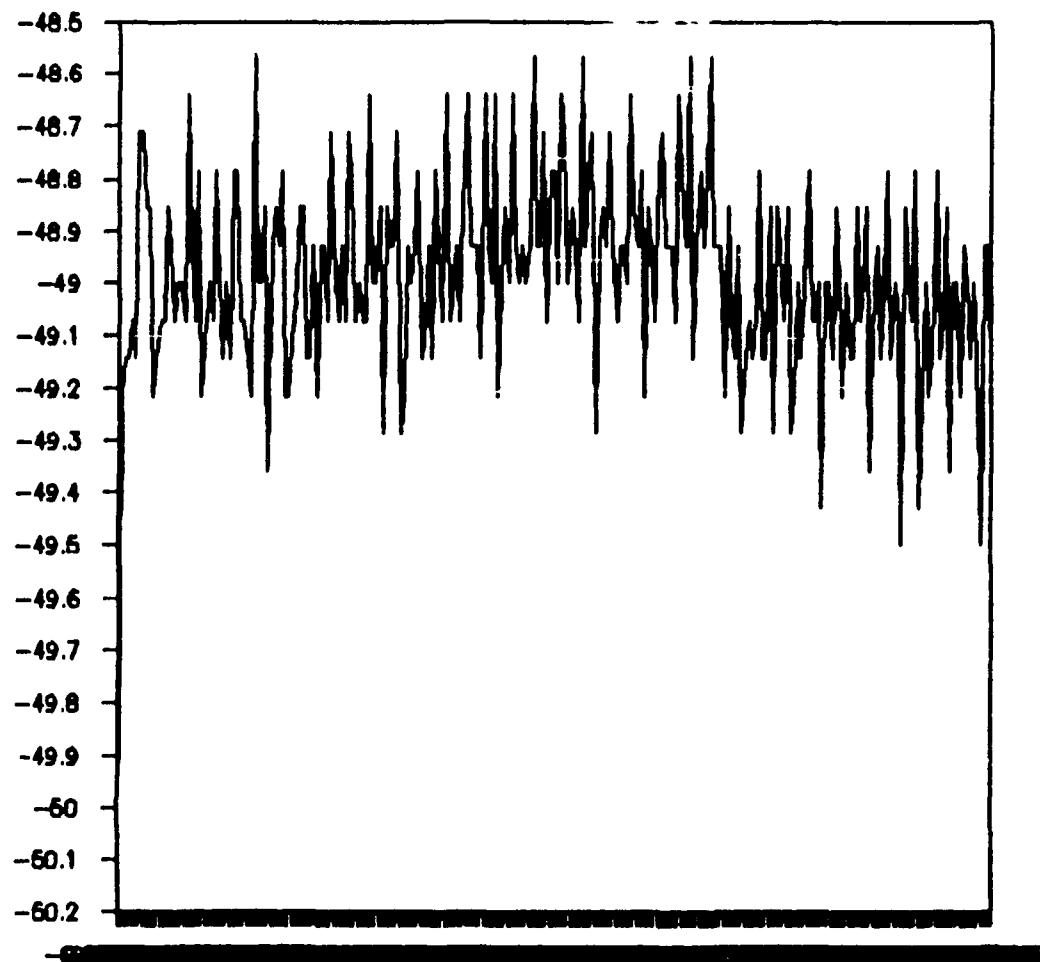


Oct 91

KTP - India ink - 100°C

small aperture

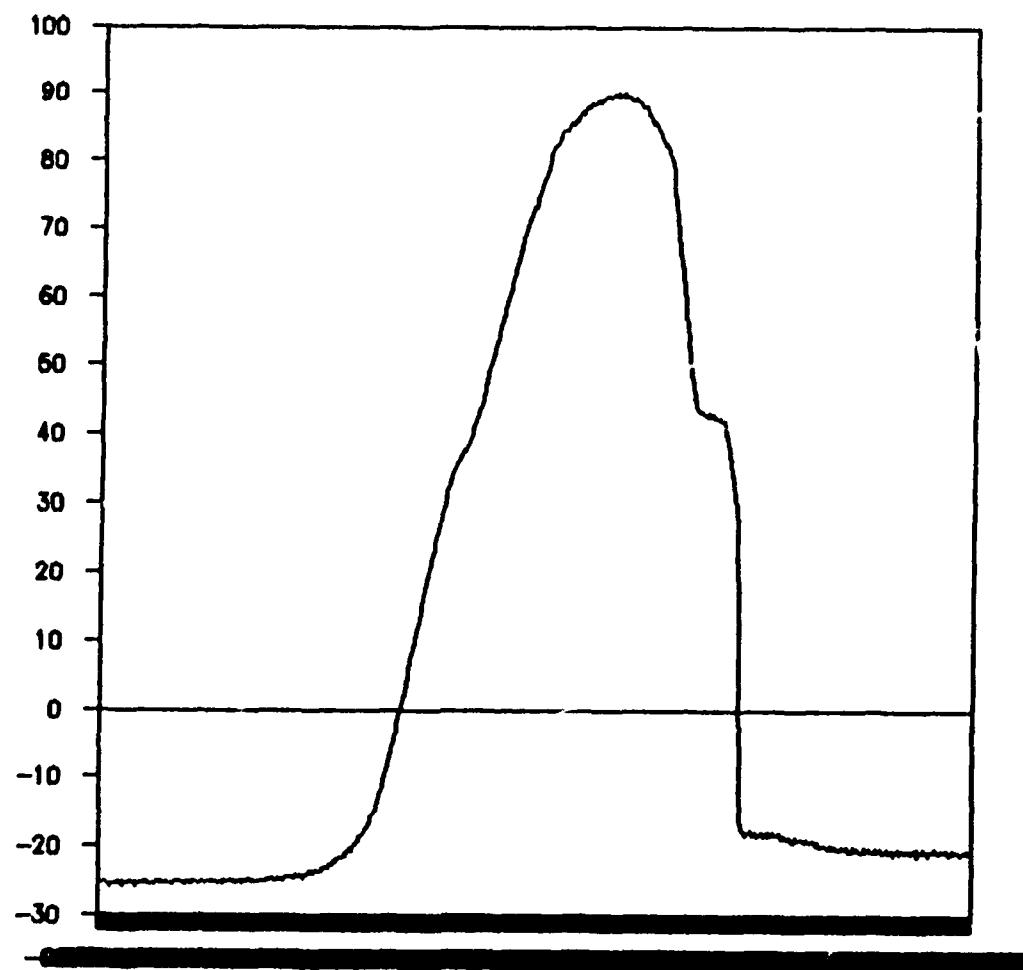
OCT9J



KTP - India un - 100°C

small aperture

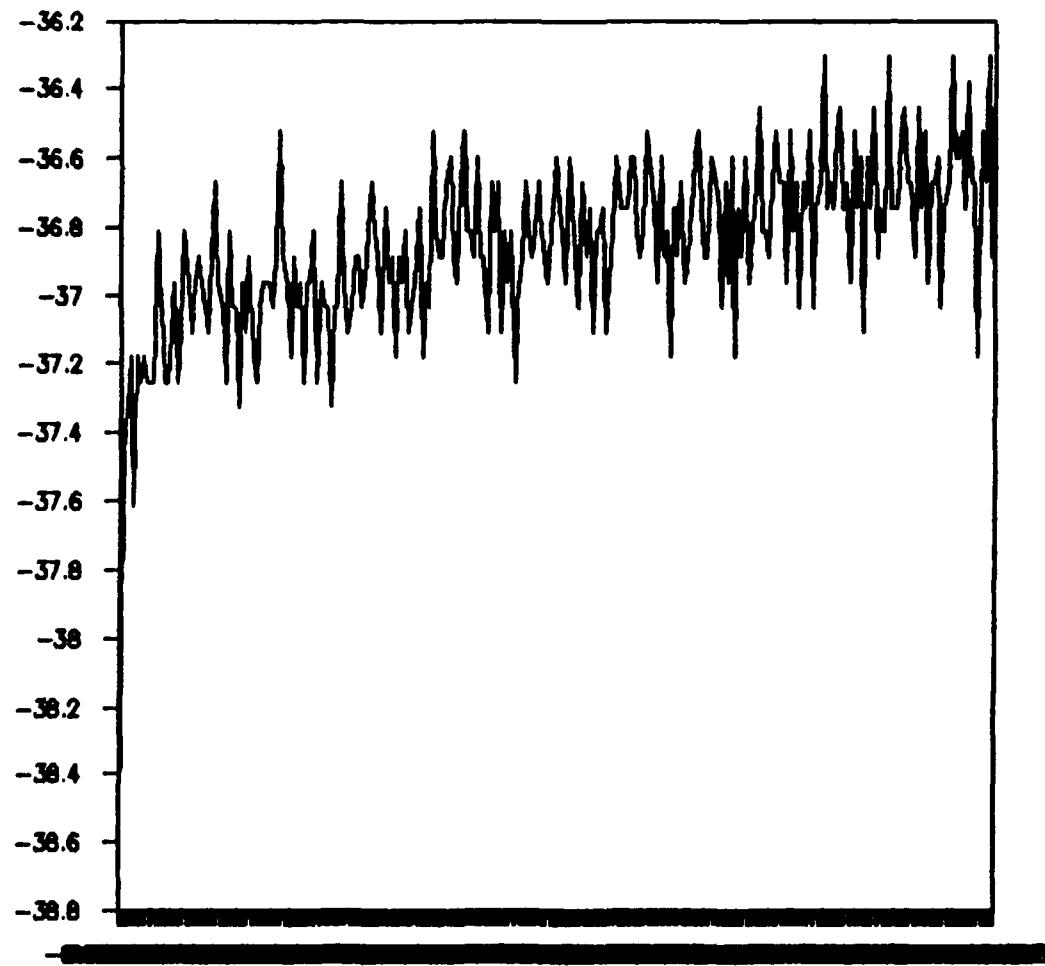
OCT9R



KTP - one step control

small aperture

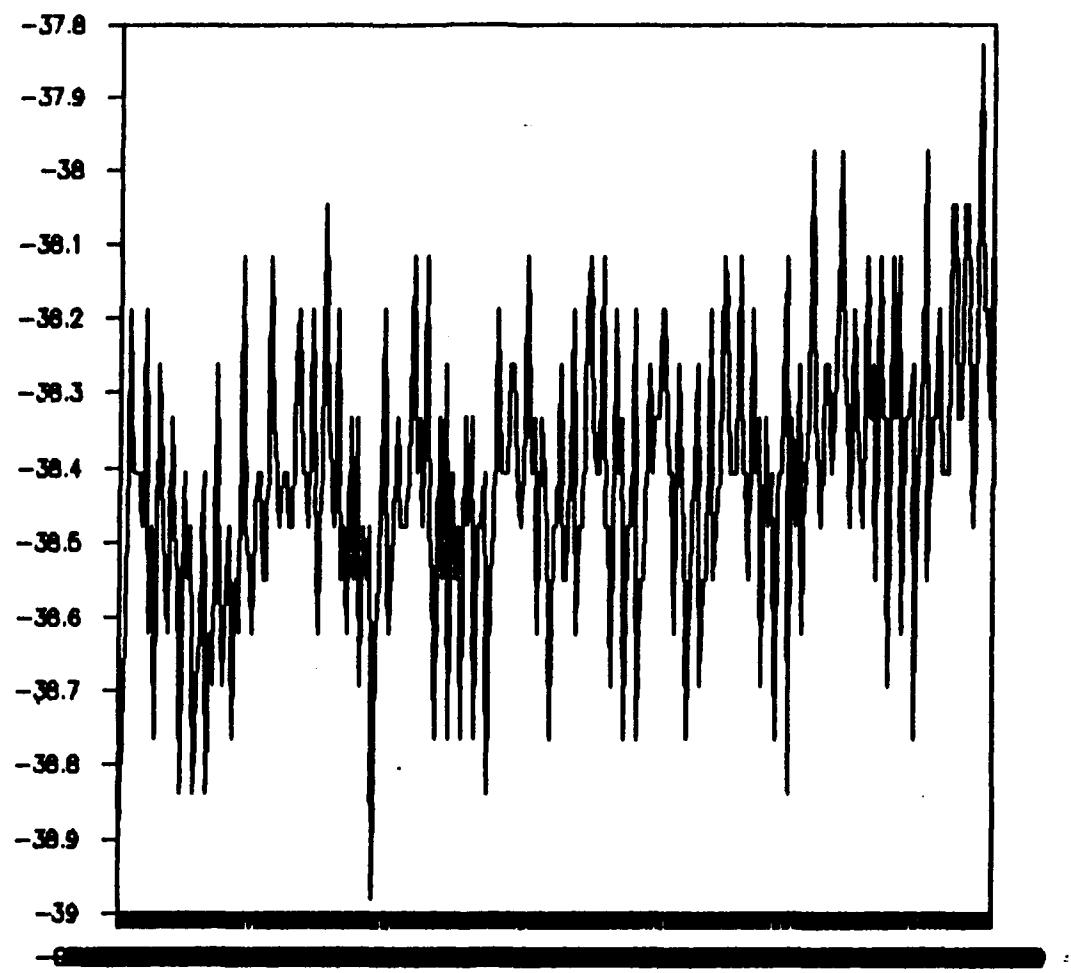
OCT9S



KTP - two strip control

small aperture

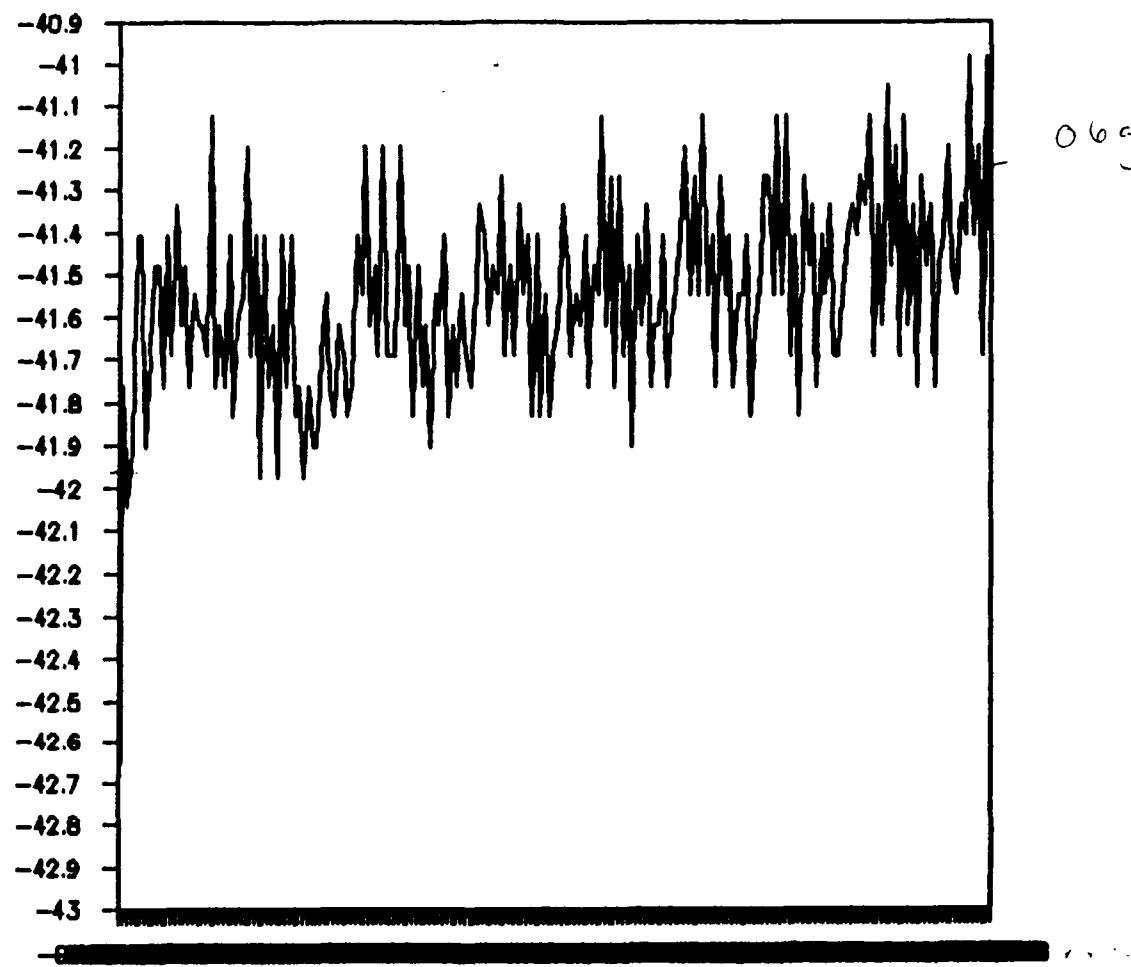
OCT9T



KTP - two strip unwelded blood
control

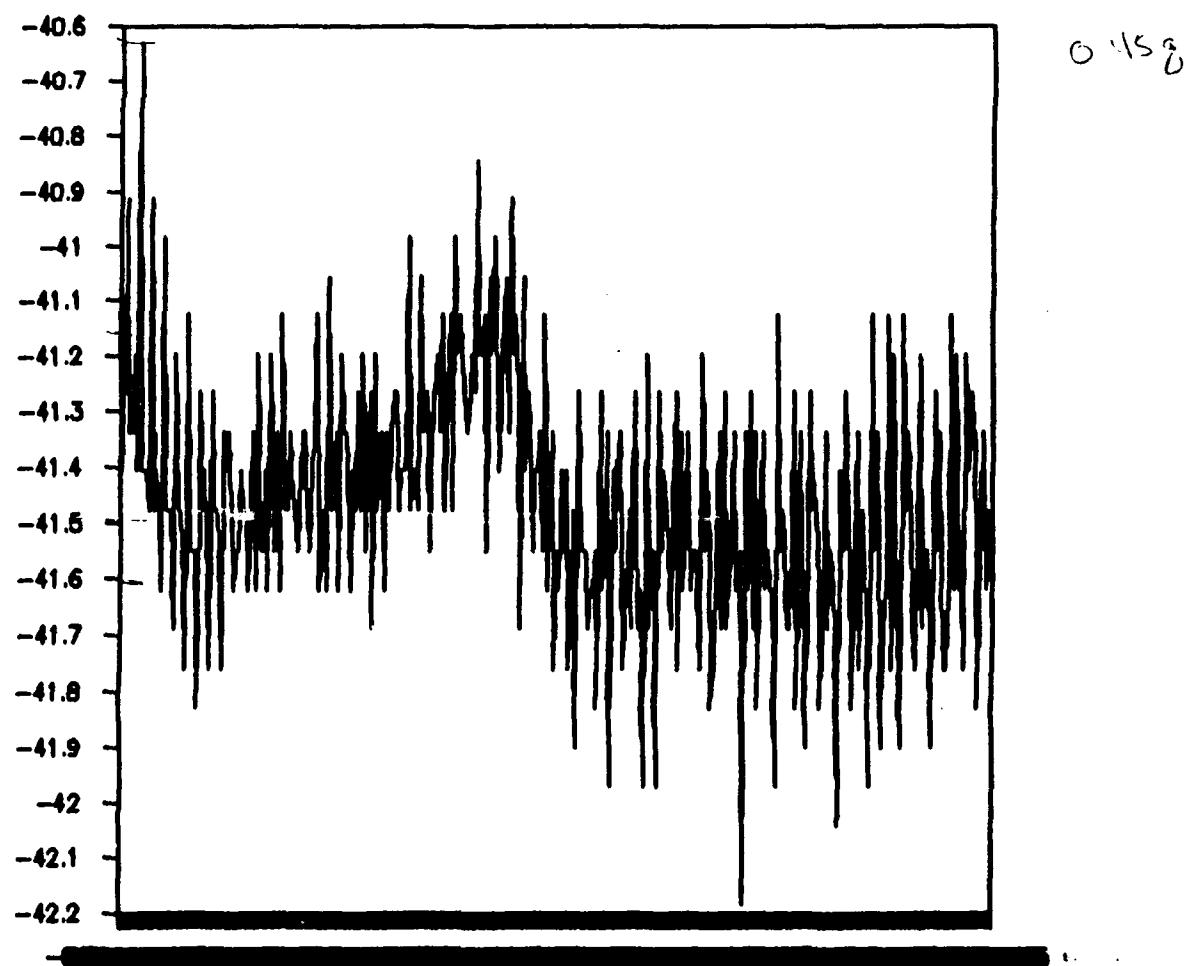
small aperture

OCT9U



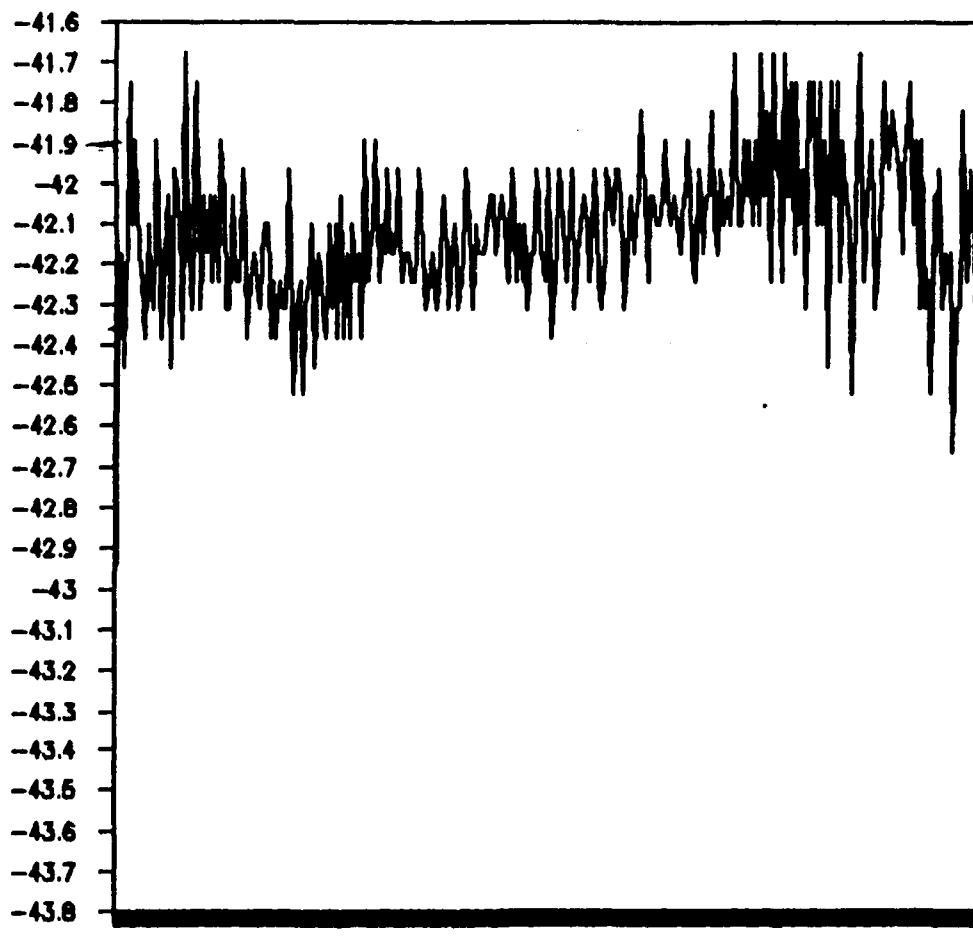
KTP - Blood - 30°C
small aperture

OCT9V1



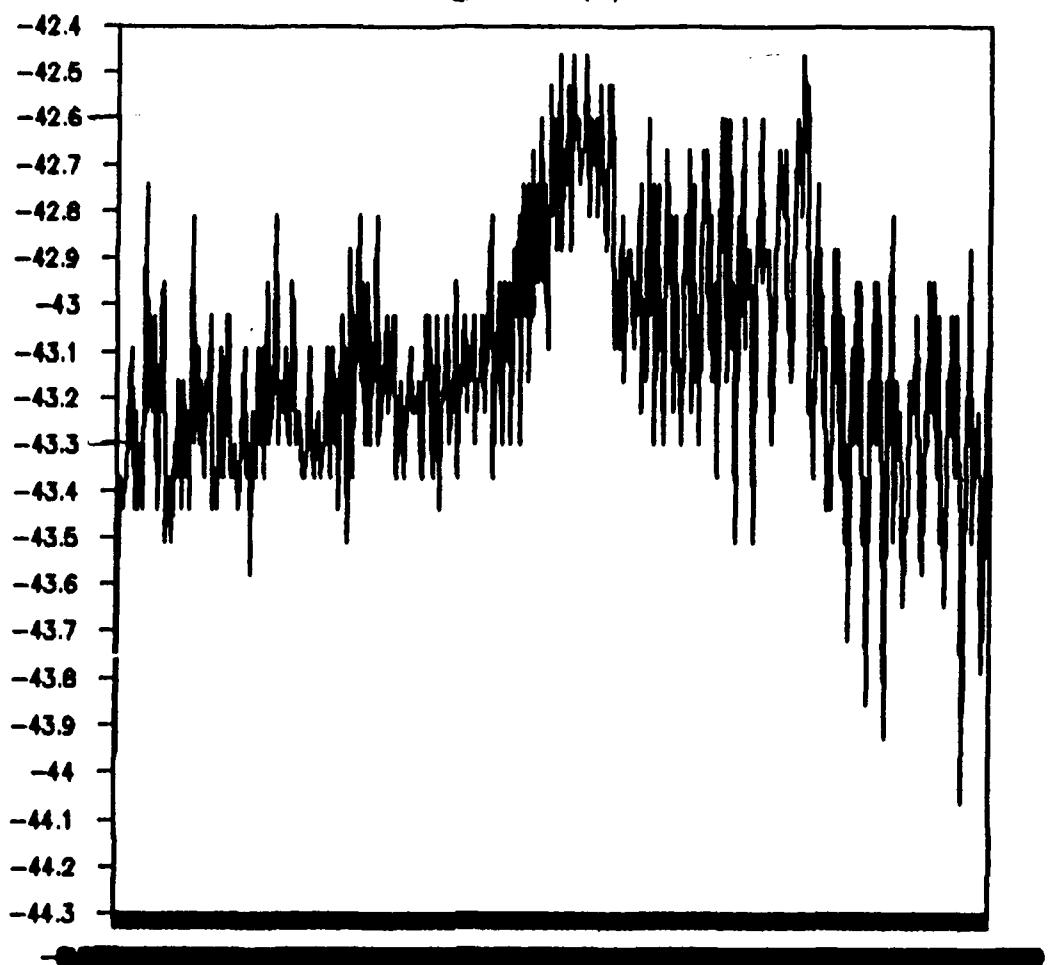
KTP - Blood - 60°C
small aperture

OCT9W



KTP - Blood - 70°C
small aperture

OCT 9 X



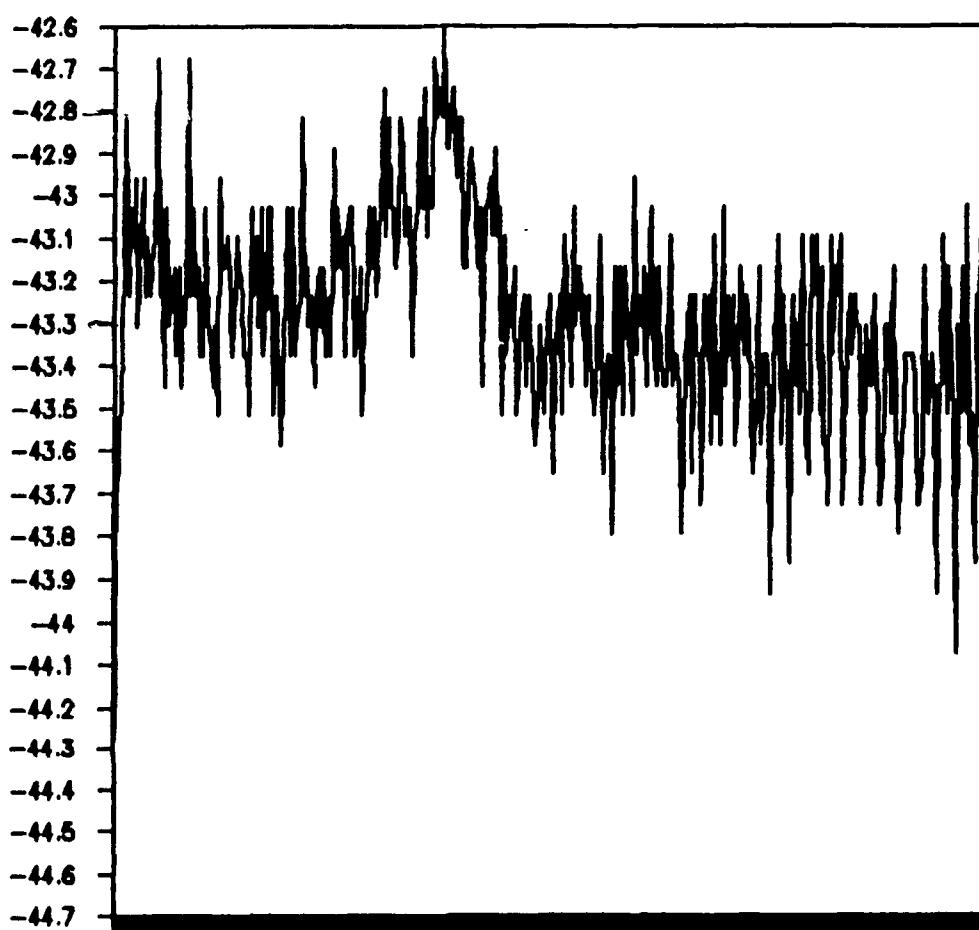
078

KTP - Blood - 80°C

small aperture

OCT9Y

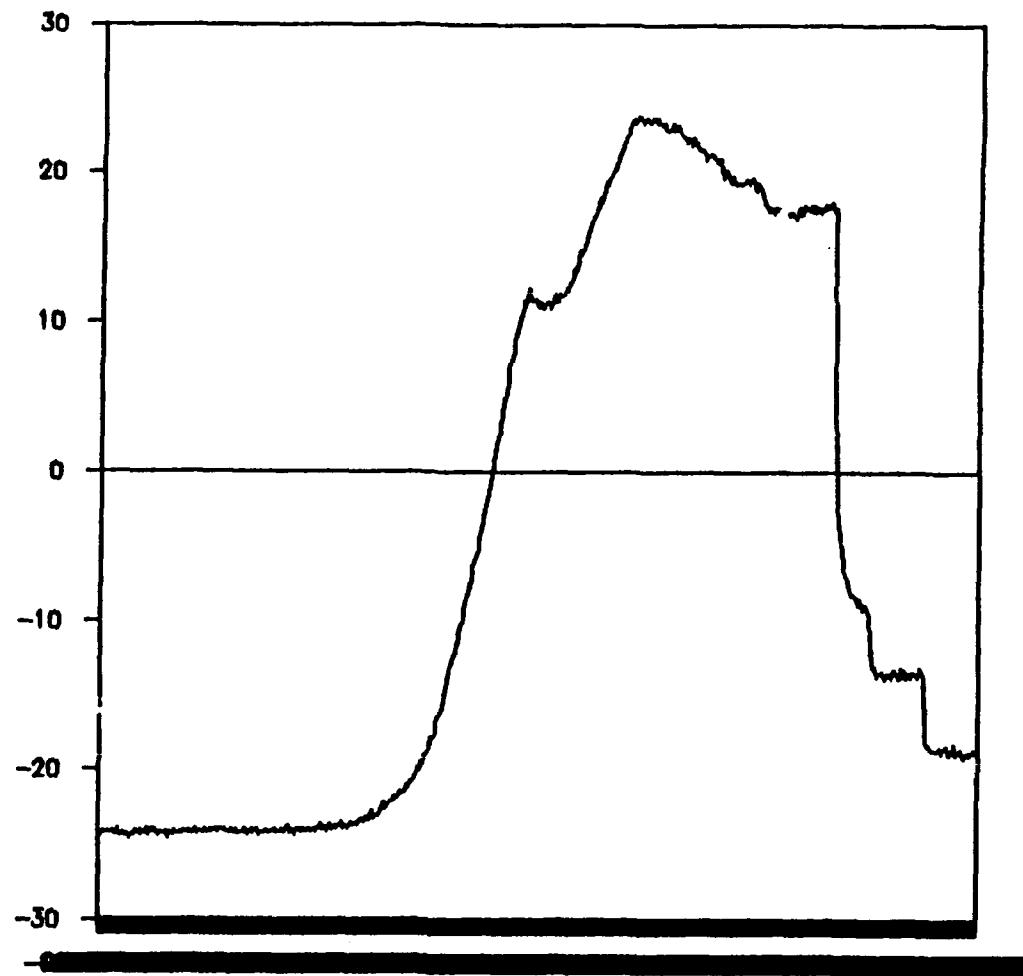
0.58



KTP - ~~EOF~~ Blood - 100°C

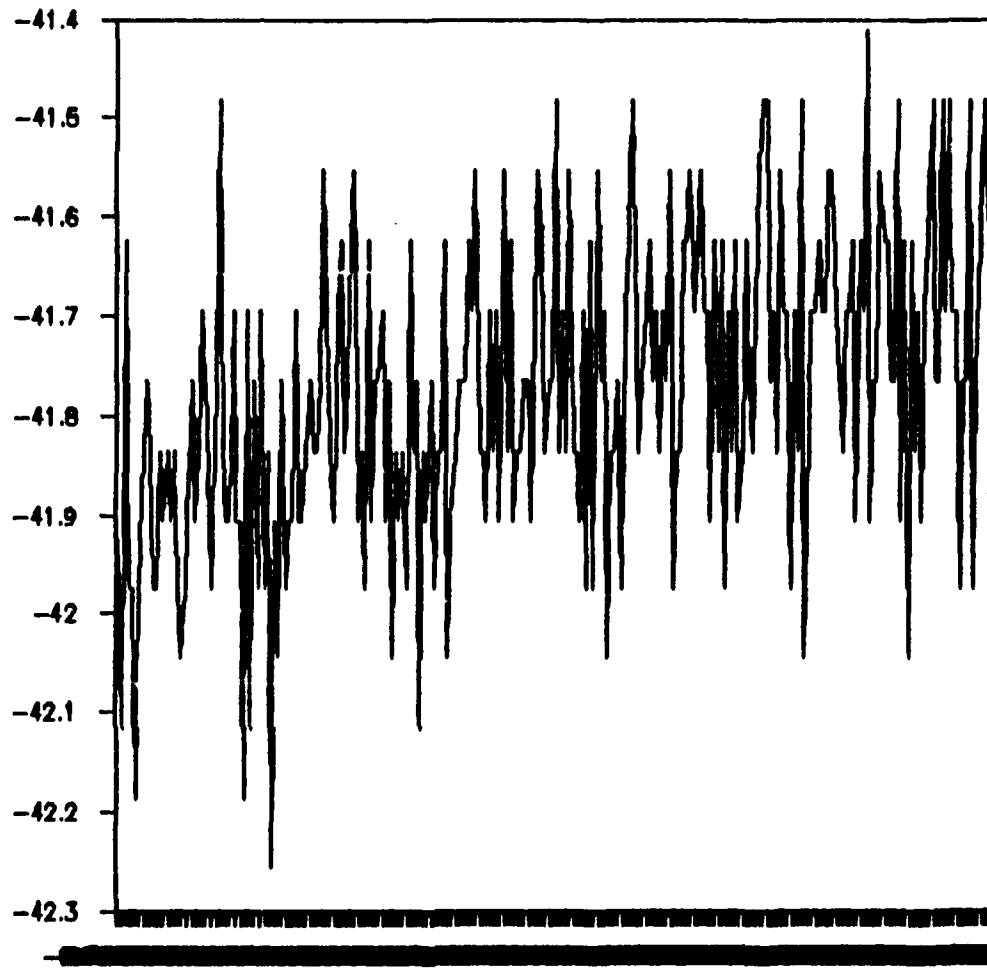
small aperture

OCT10A



KTP — one strip unwelded control
small aperture

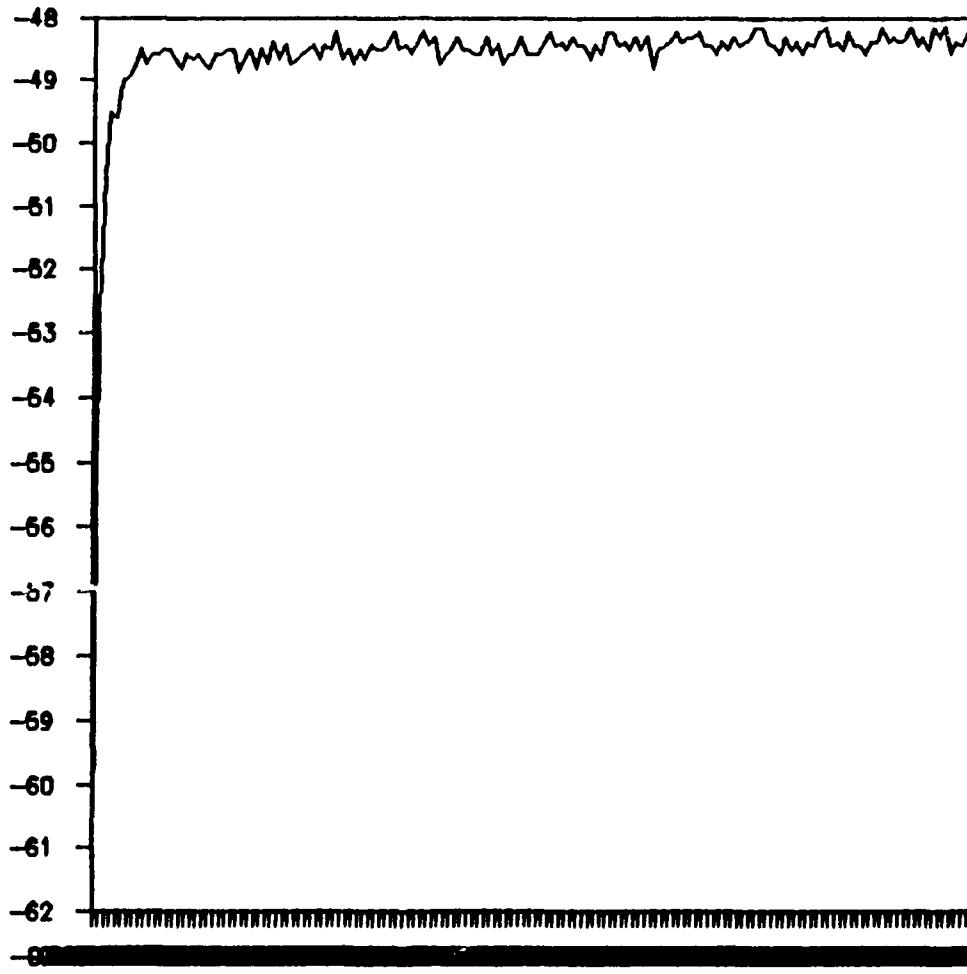
OCT10B



KTP - two step unwelded control

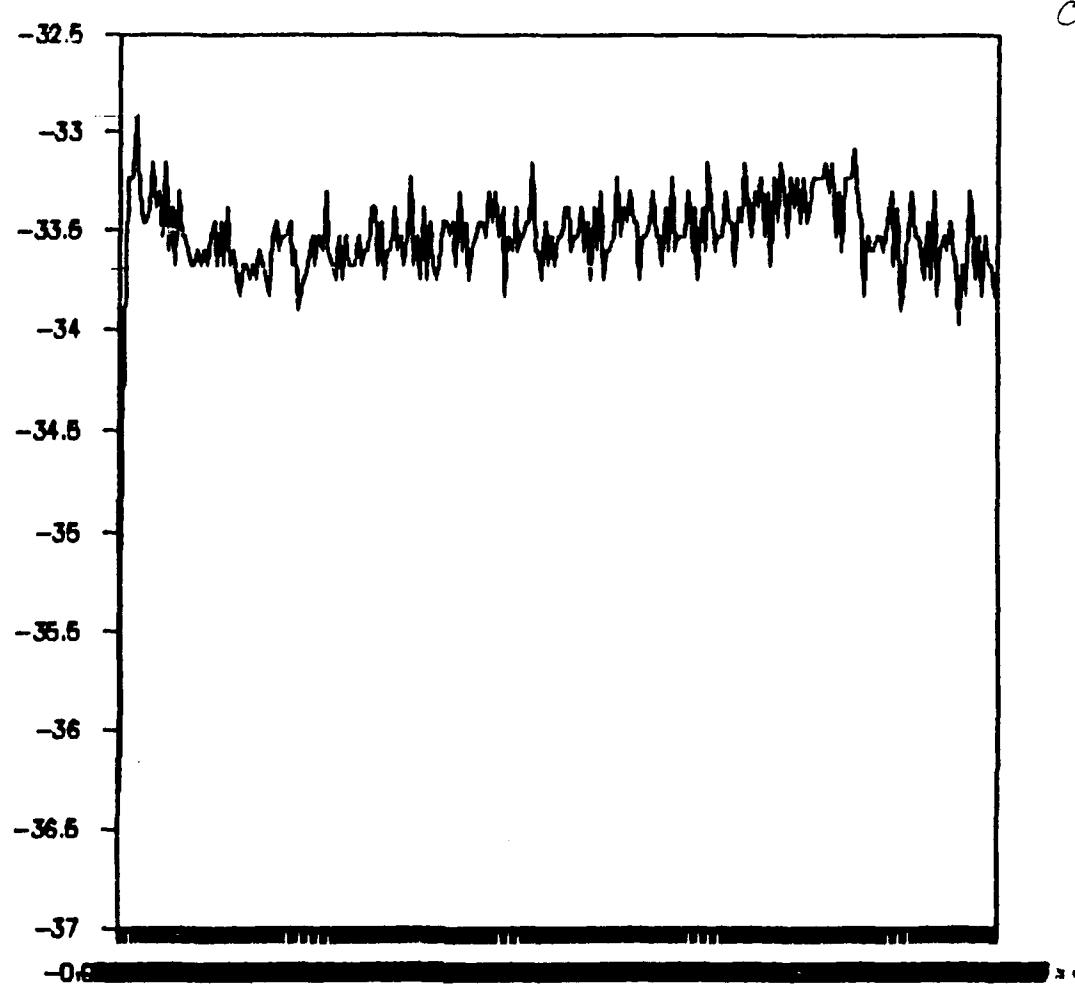
small aperture

OCT10C



KTP - 2 v ICG unwelded control
small aperture

OCT10D



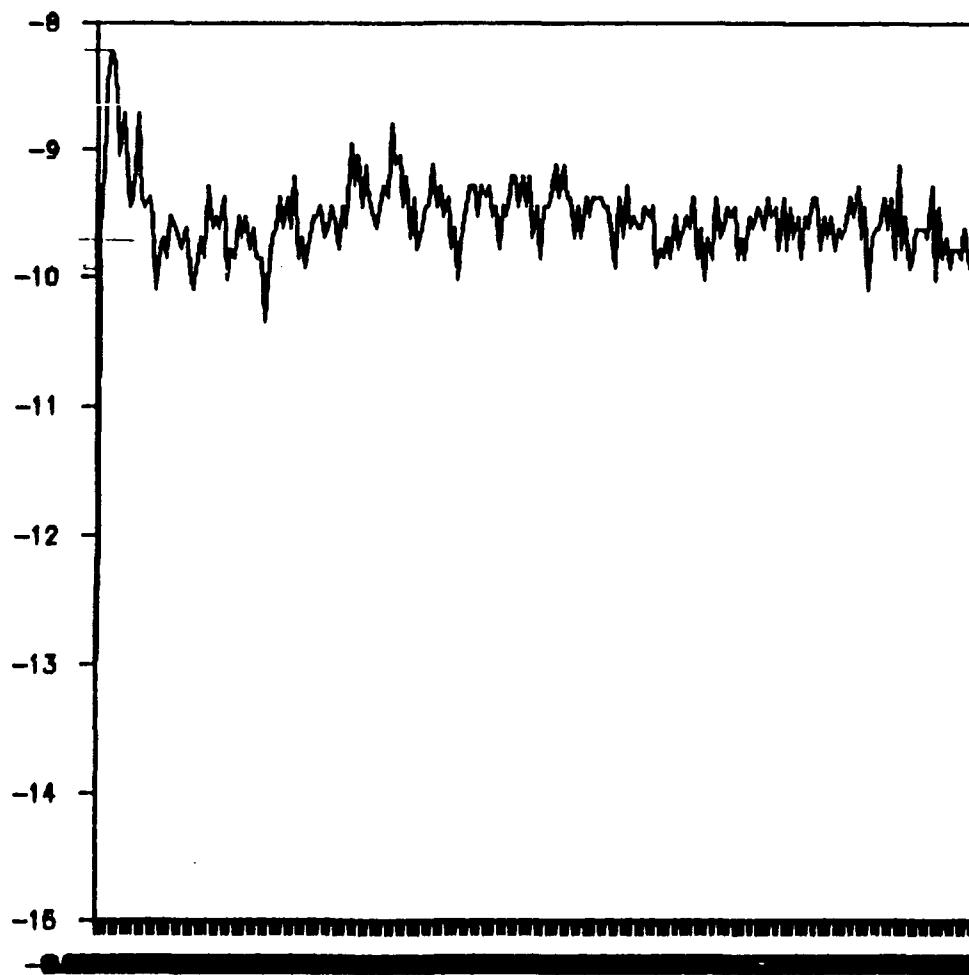
0.5g

KTP - 2x 1C6 - 50°C

small aperture

OCT10E

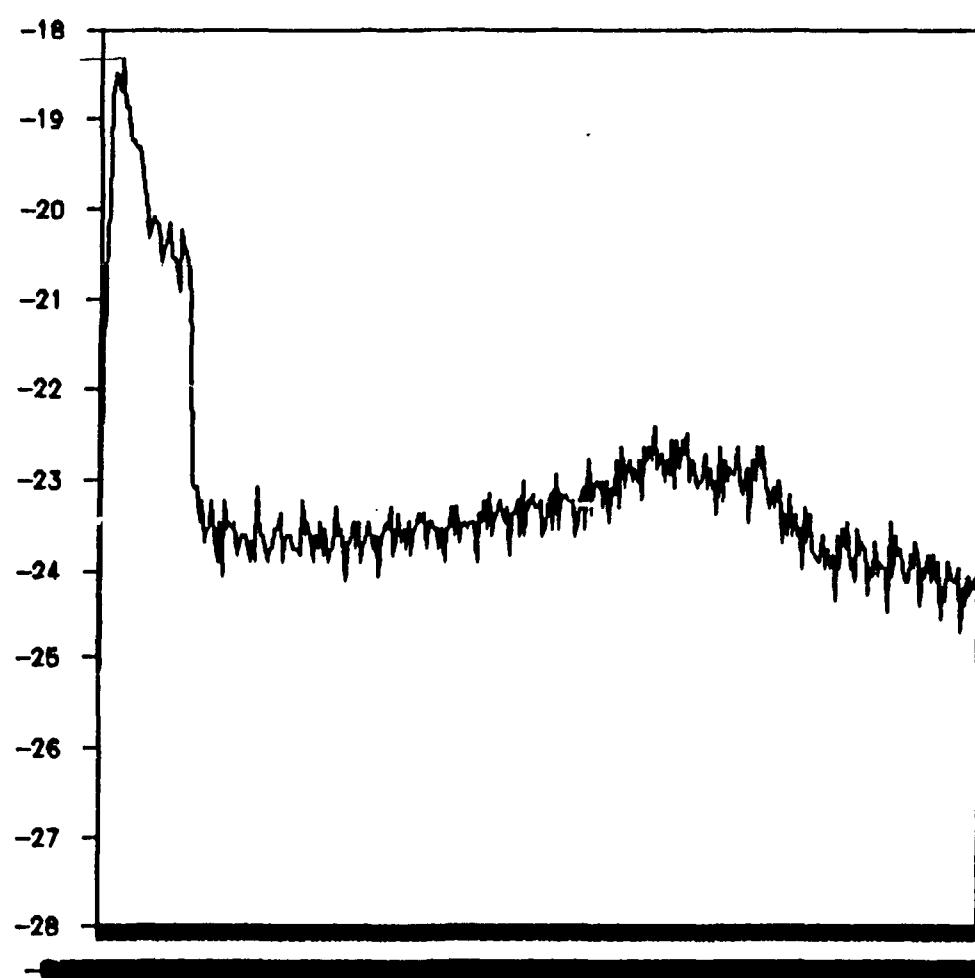
6.28



KTP - 2X 1CG - 60°C

small aperture

OCT10F1

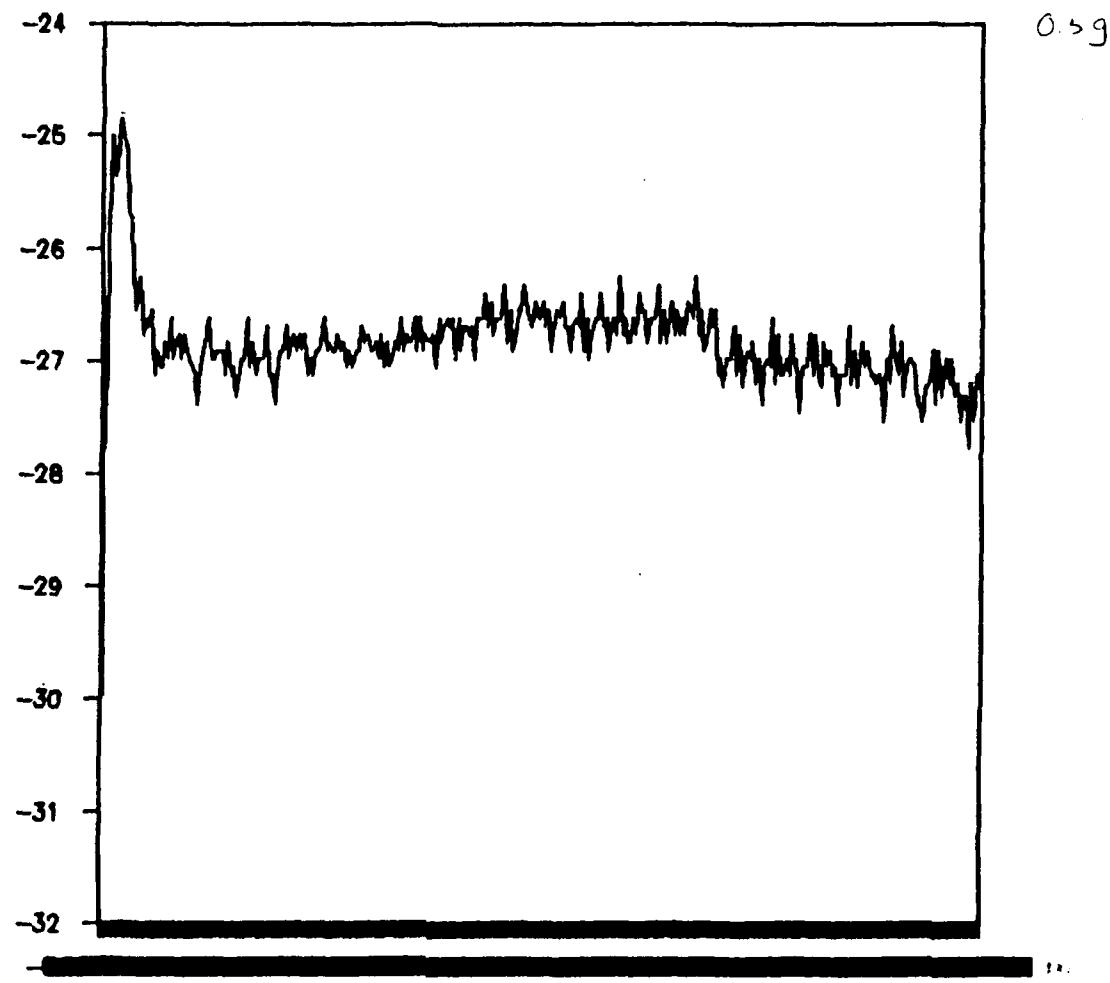


19

KTP - 2x ICG - 70°C

small aperture

OCT10G

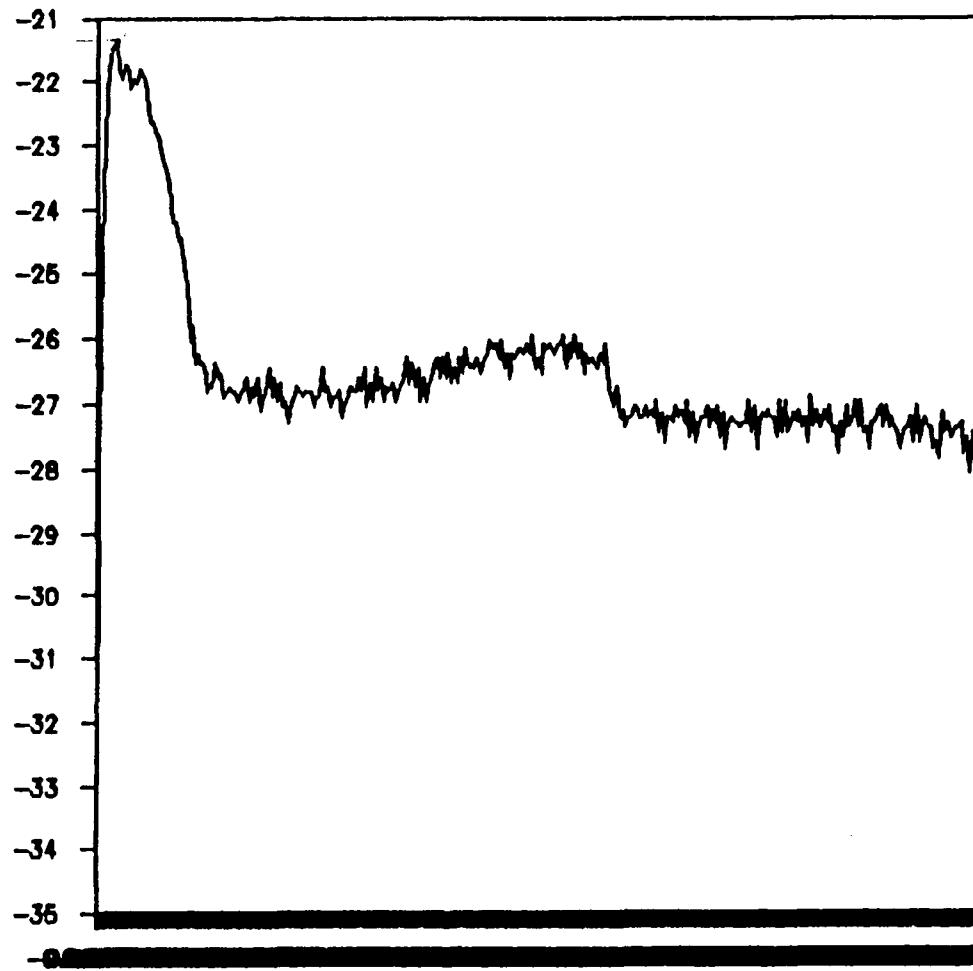


KTP - 2X 1CG - 80°C

small aperture

OCT10H

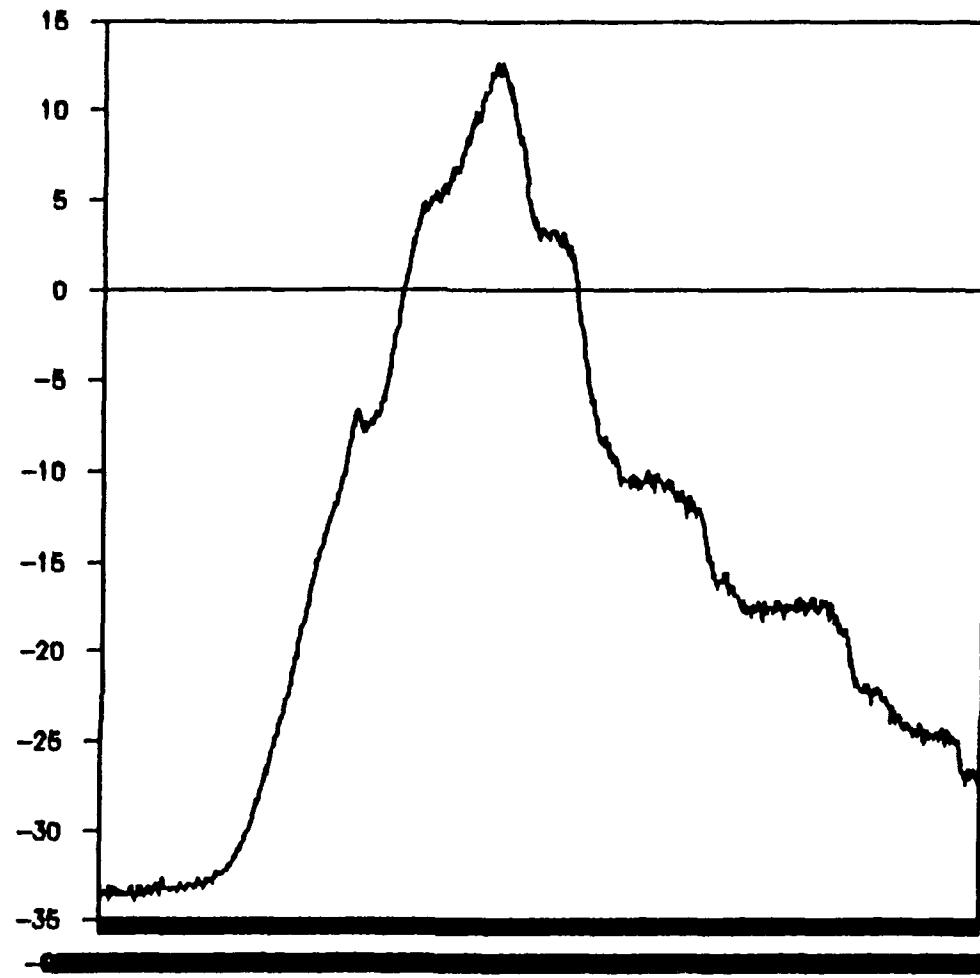
075g



KTP - 2x ICG - 100°C

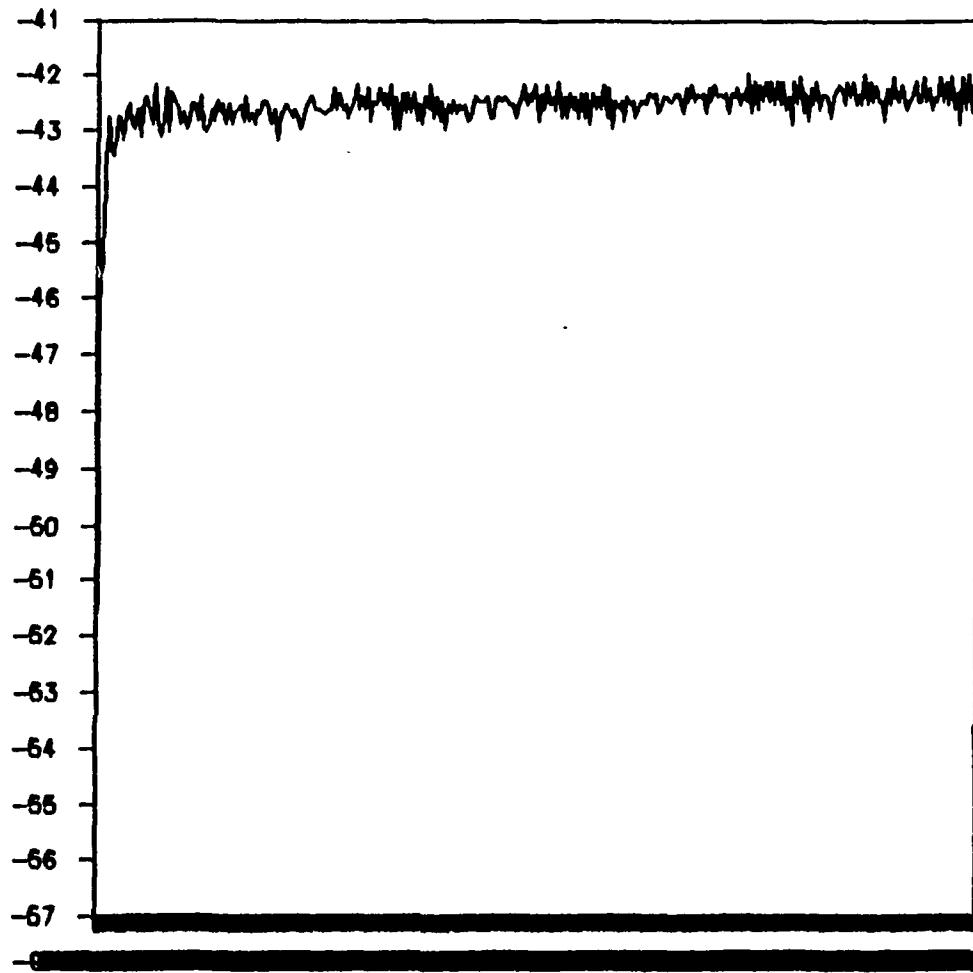
small aperture

OCT10I



KTP - one strip unwelded control
small aperture

OCT10J

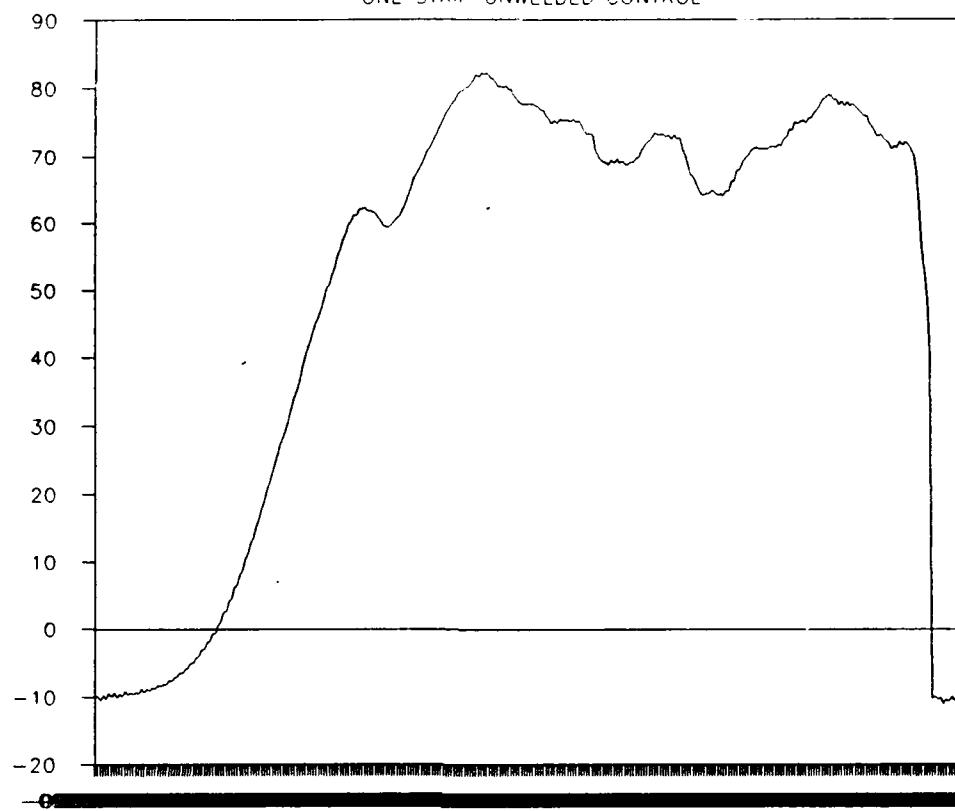


KTP - two strip unwelded control

small aperture

OCT15A

ONE STRIP UNWELDED CONTROL



All trials for OCT 15, 1990

Ar⁺⁺ Laser

Settings: Current control

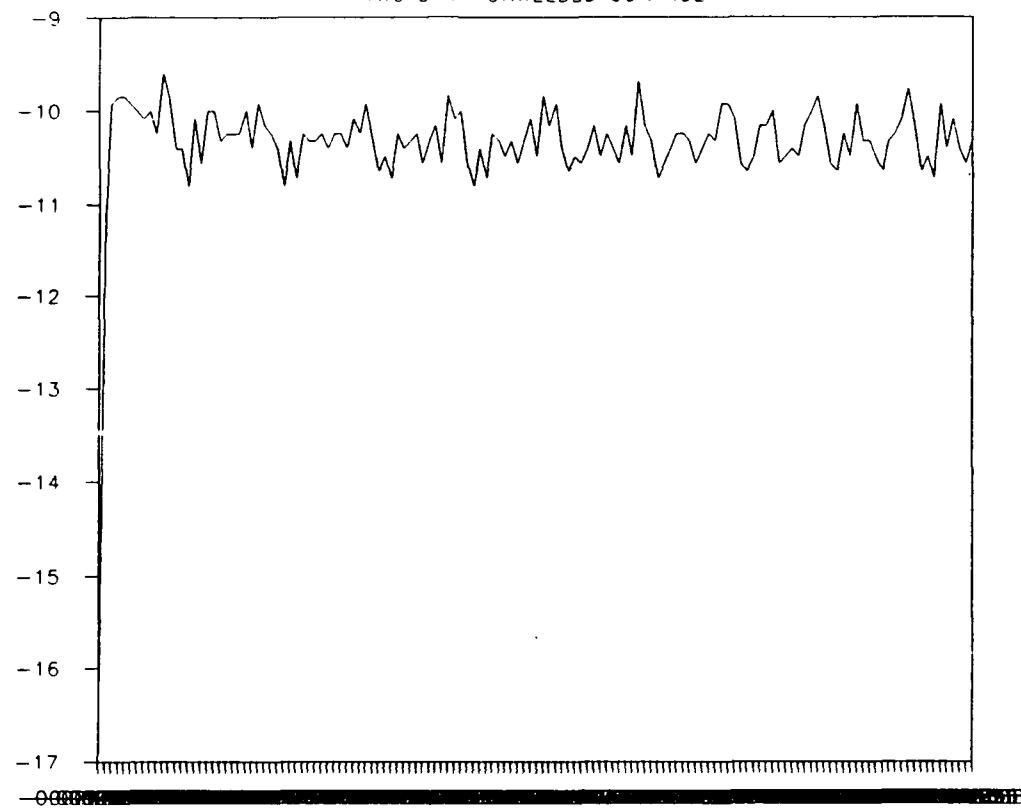
38A, 5.8W

0.7W delivered

Small Aperture

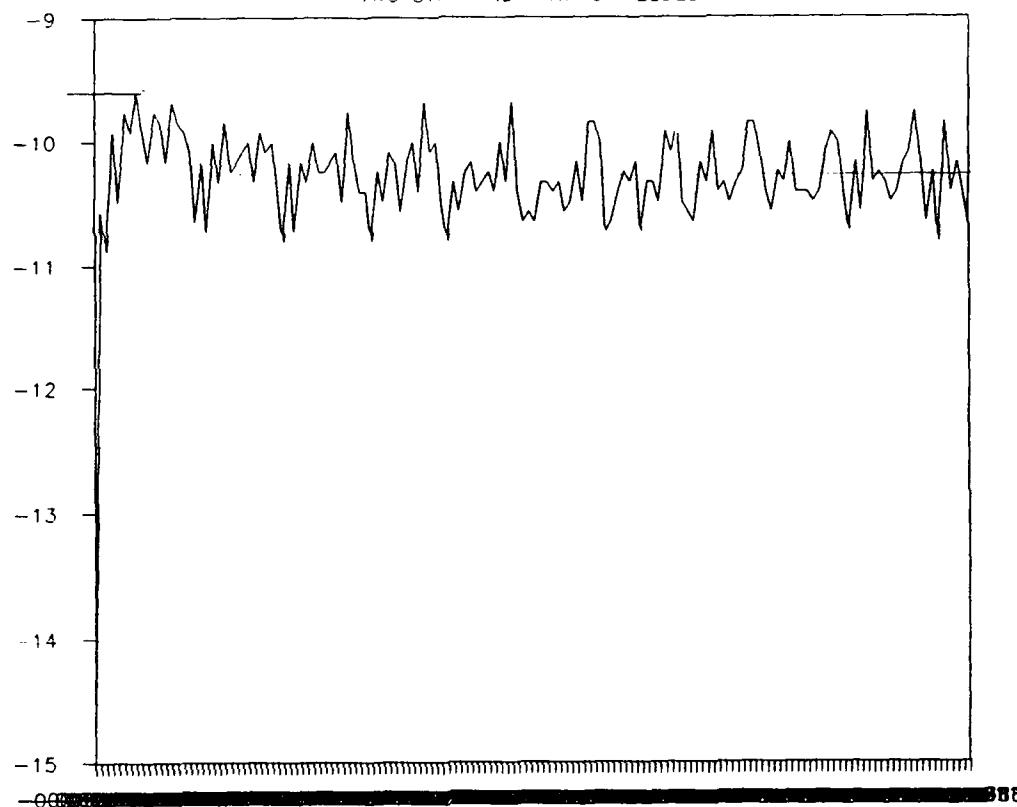
OCT15B

TWO STRIP UNWELDED CONTROL



OCT15C

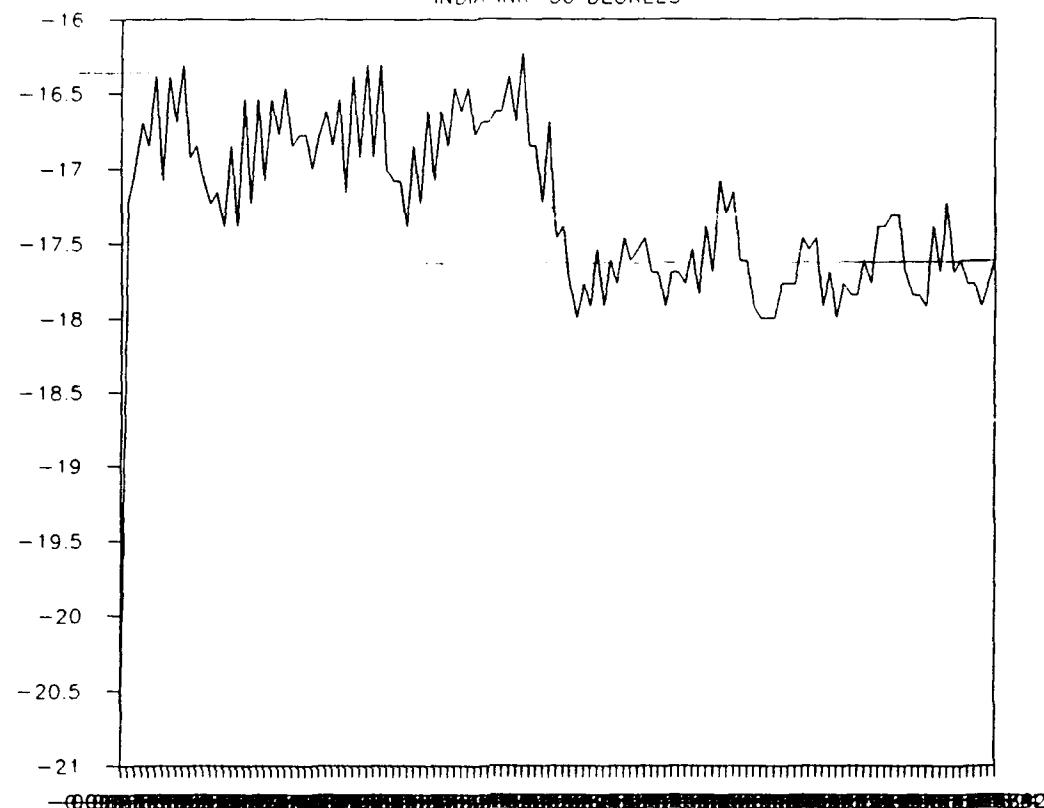
TWO STRIP INDIA INK UNWELDED



OCT15D

INDIA INK - 50 DEGREES

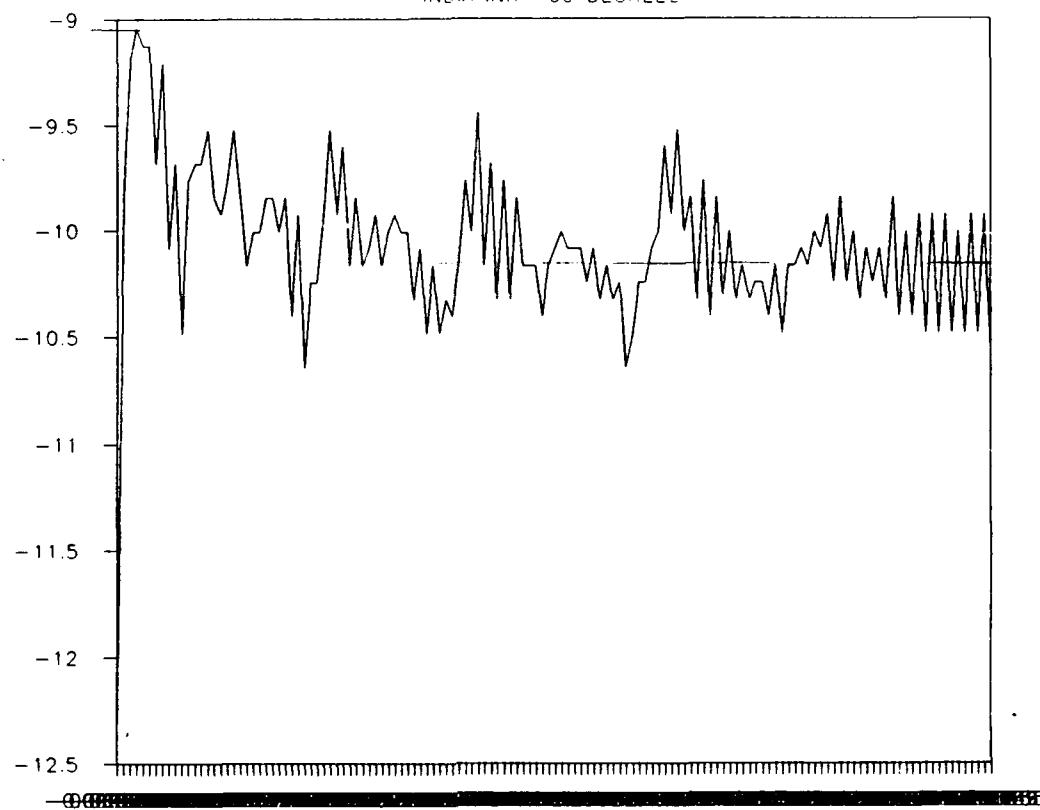
20°c



61

OCT15E

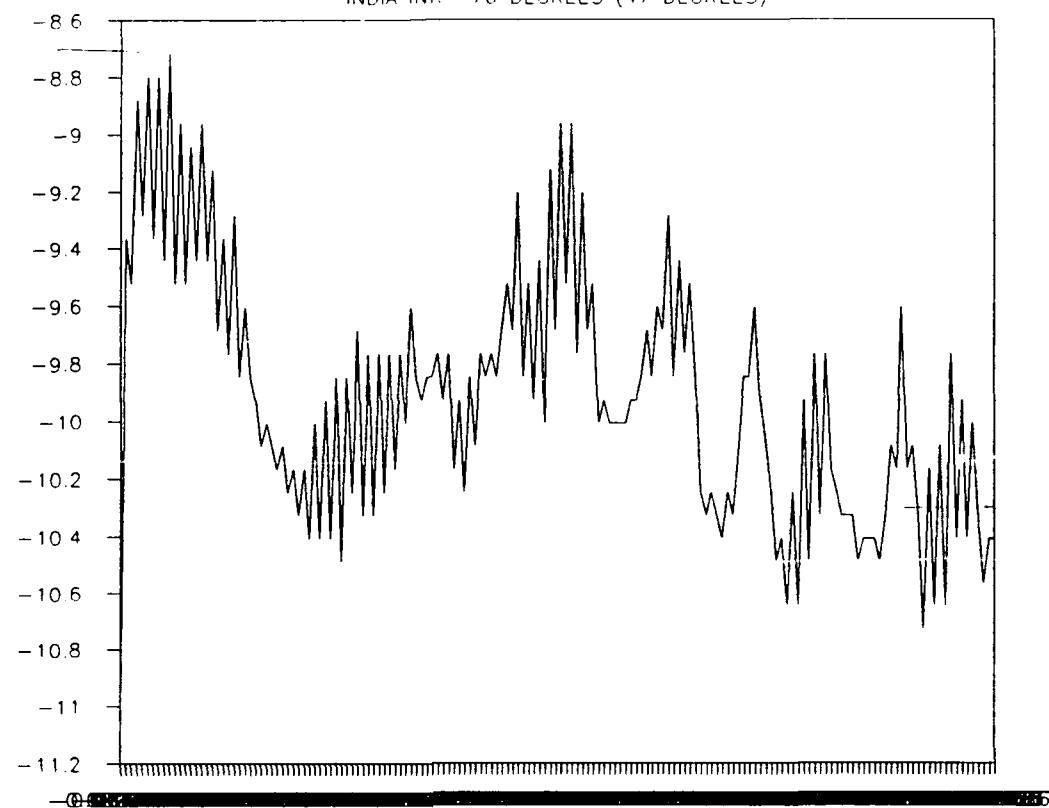
INDIA INK - 60 DEGREES



OCT15F1

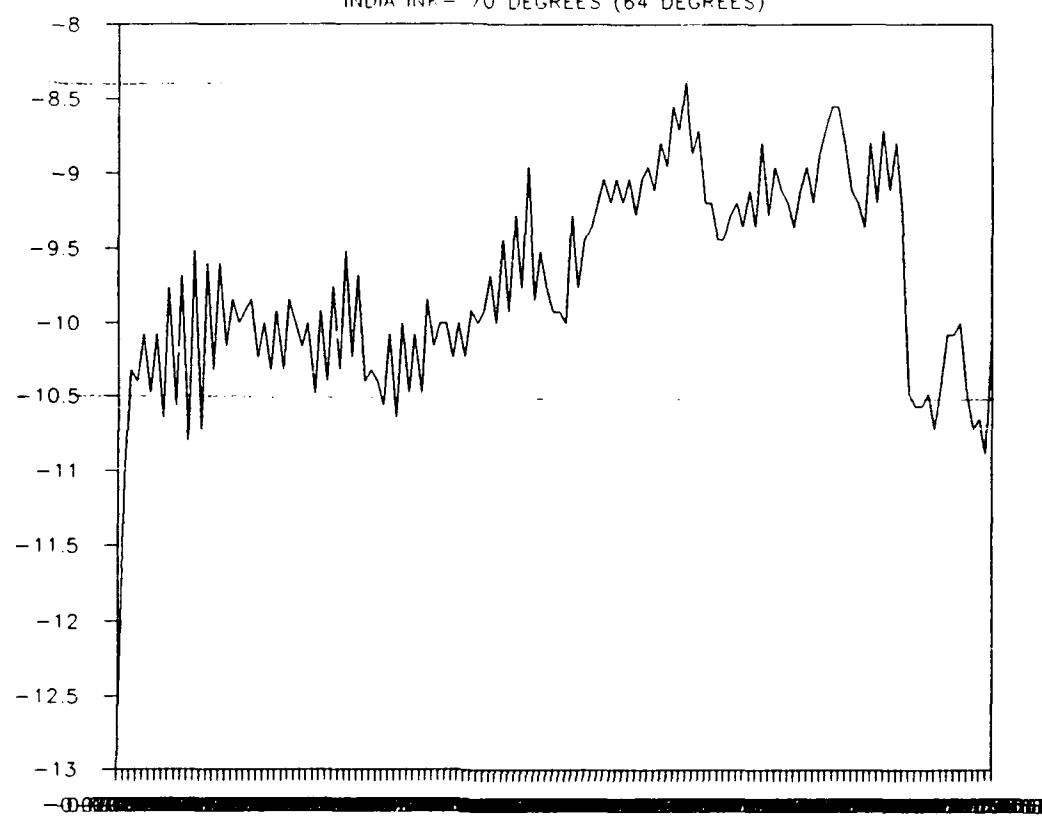
15

INDIA INK - 70 DEGREES (47 DEGREES)



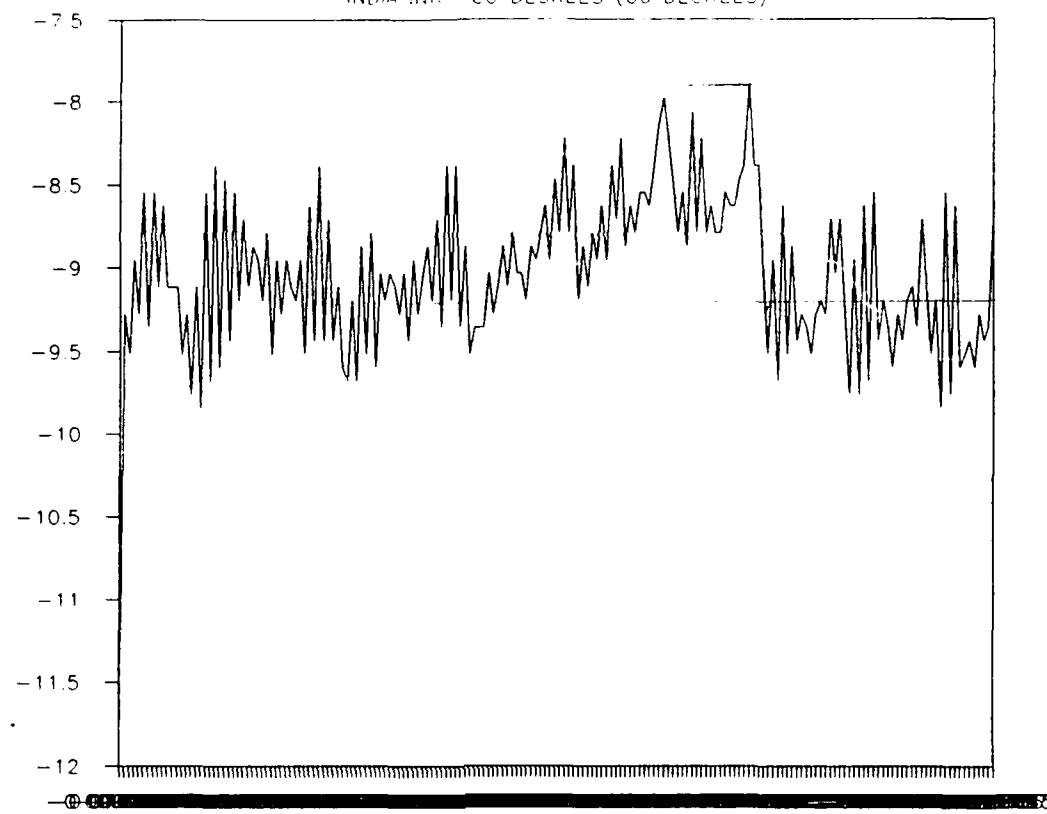
OCT15G1

INDIA INK - 70 DEGREES (64 DEGREES)



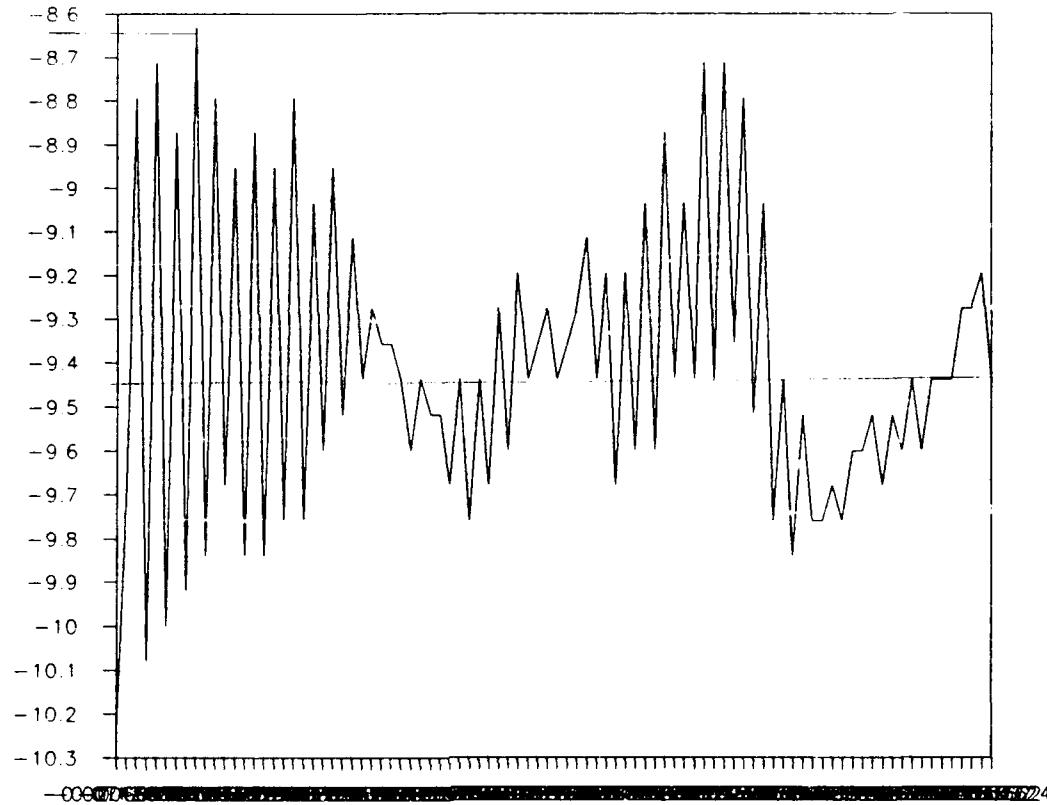
OCT15H1

INDIA INK - 80 DEGREES (66 DEGREES)



OCT15I

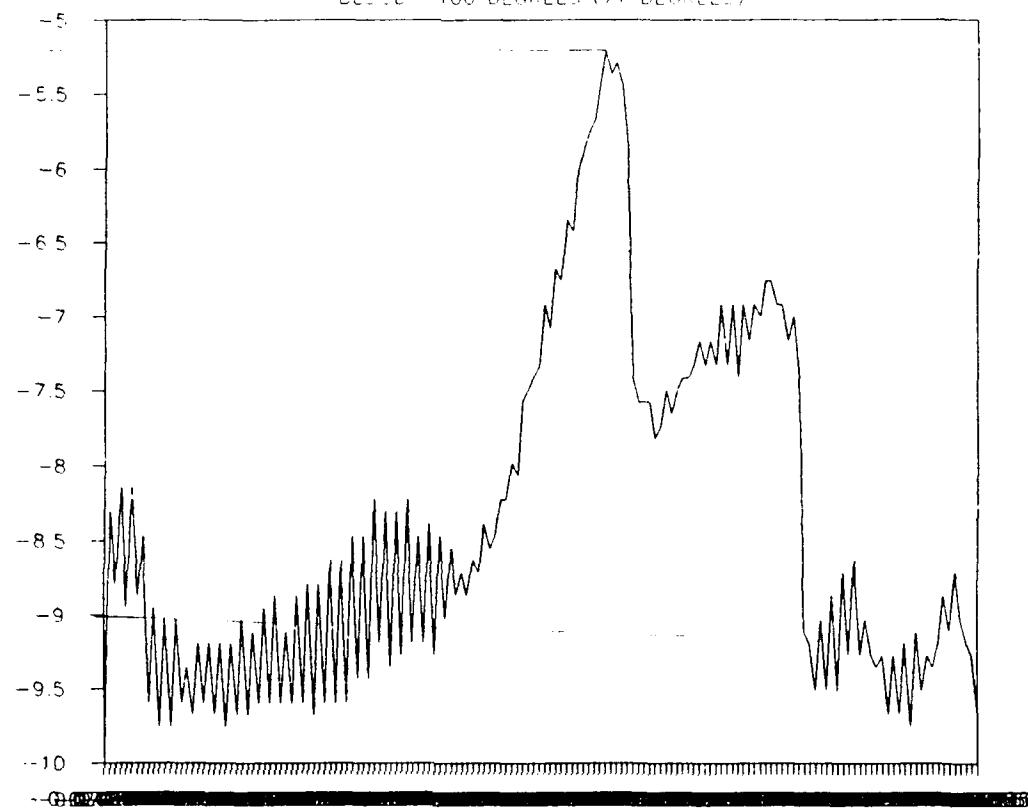
TWO STRIP UNWELDED BLOOD CONTROL



409

OCT15J1

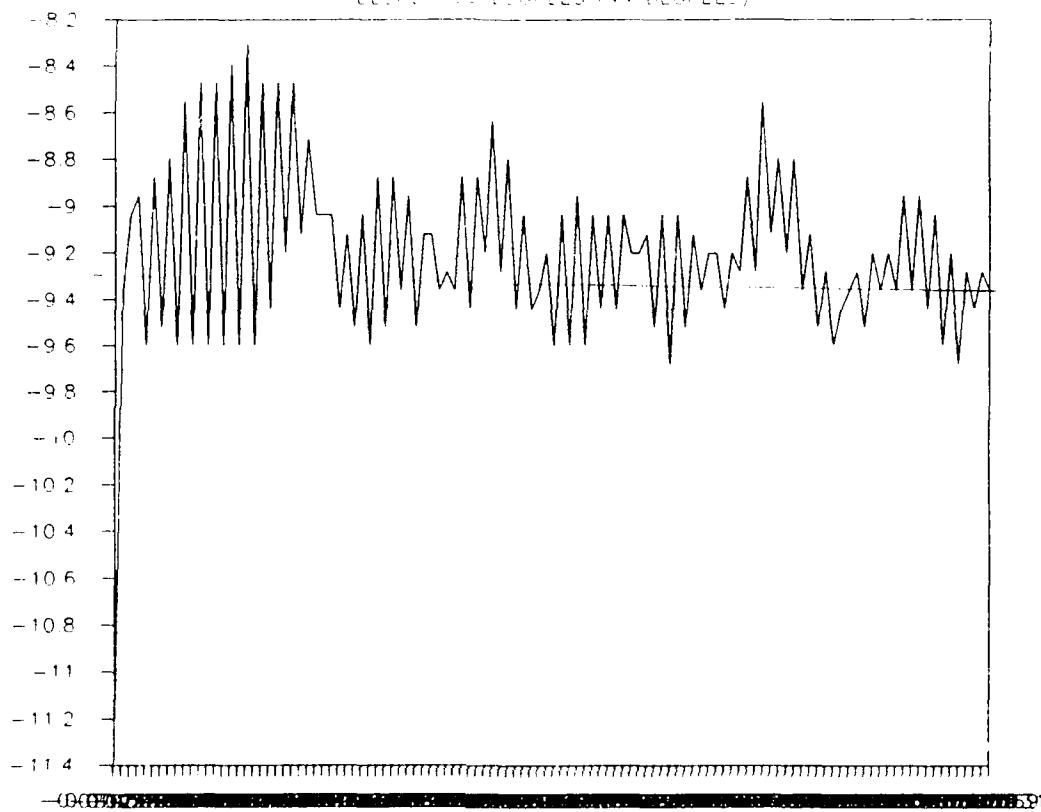
BLOOD - 100 DEGREES (77 DEGREES)



OCT 15 K 1

5.2 C

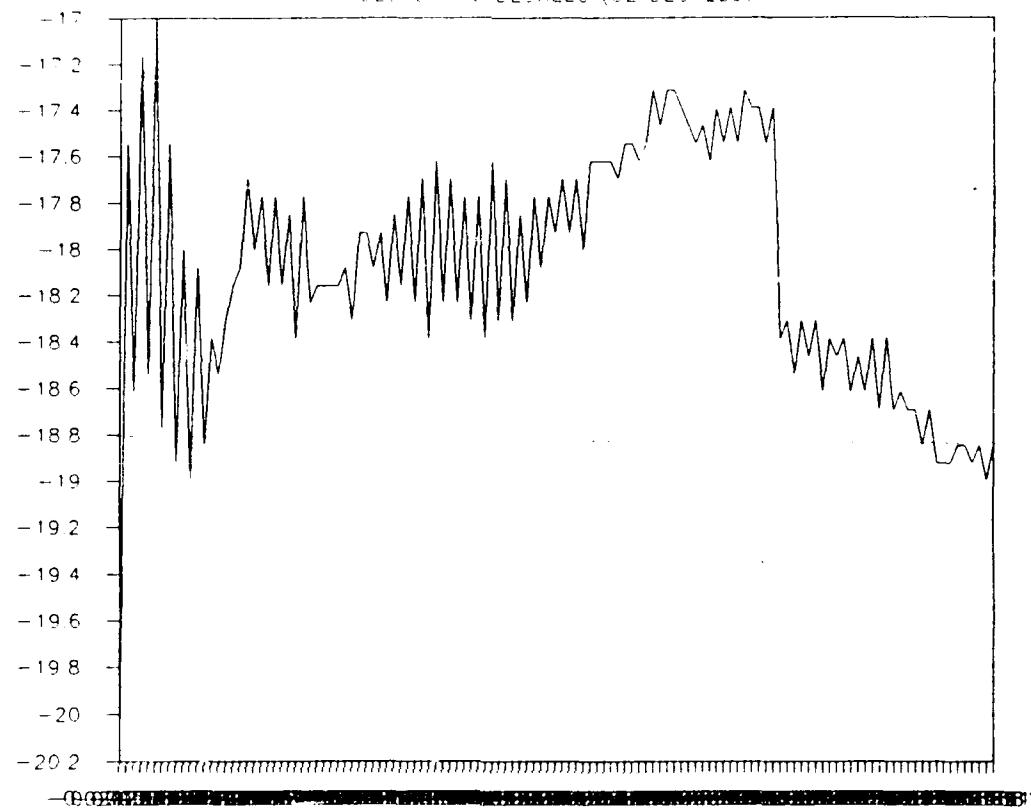
BLOOD - 50 DEGREES (44 DEGREES)



OCT15M1

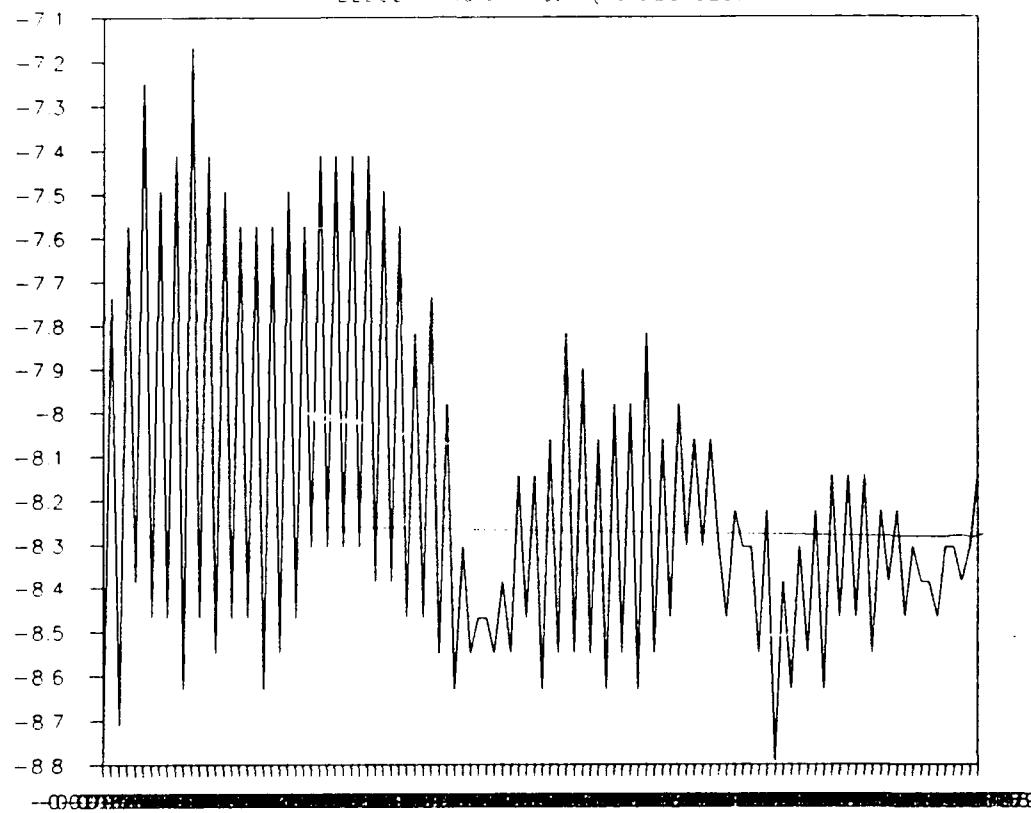
105

BLOOD - 70 DEGREES (62 DEGREES)



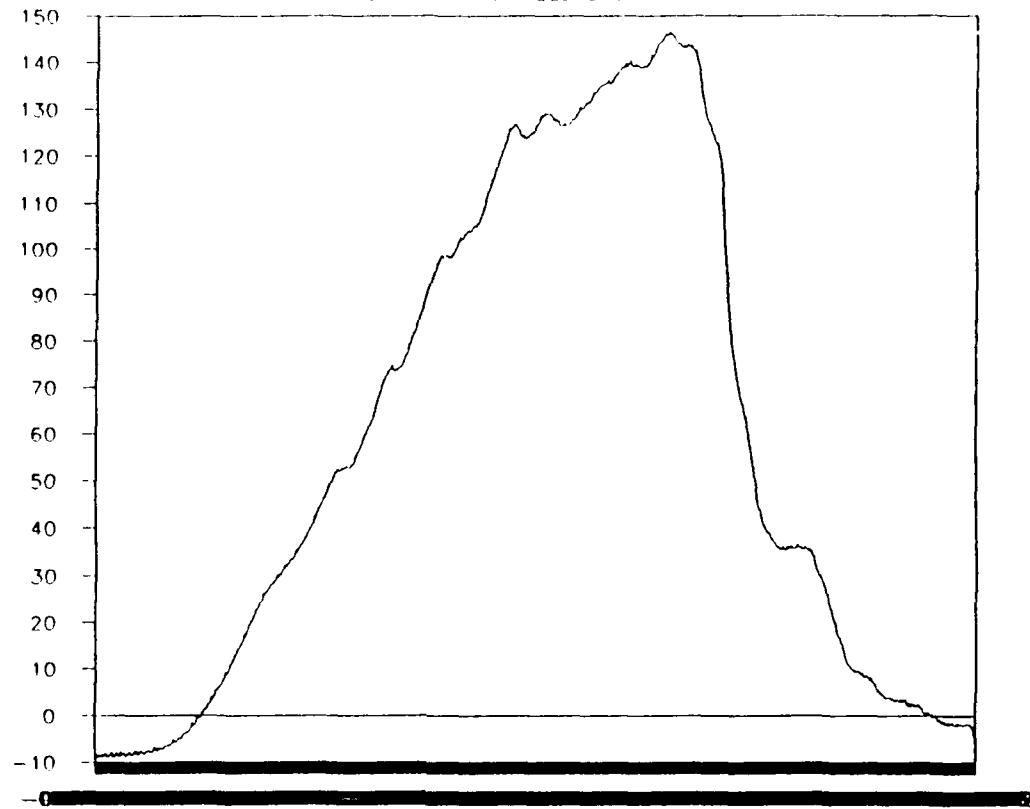
OCT15P1

BLOOD- 100 DEGREES (56 DEGREES)



OCT16A

ONE STRIP UNWELDED CONTROL



All trials for OCT 16, 1990

Ar⁺⁺ Laser

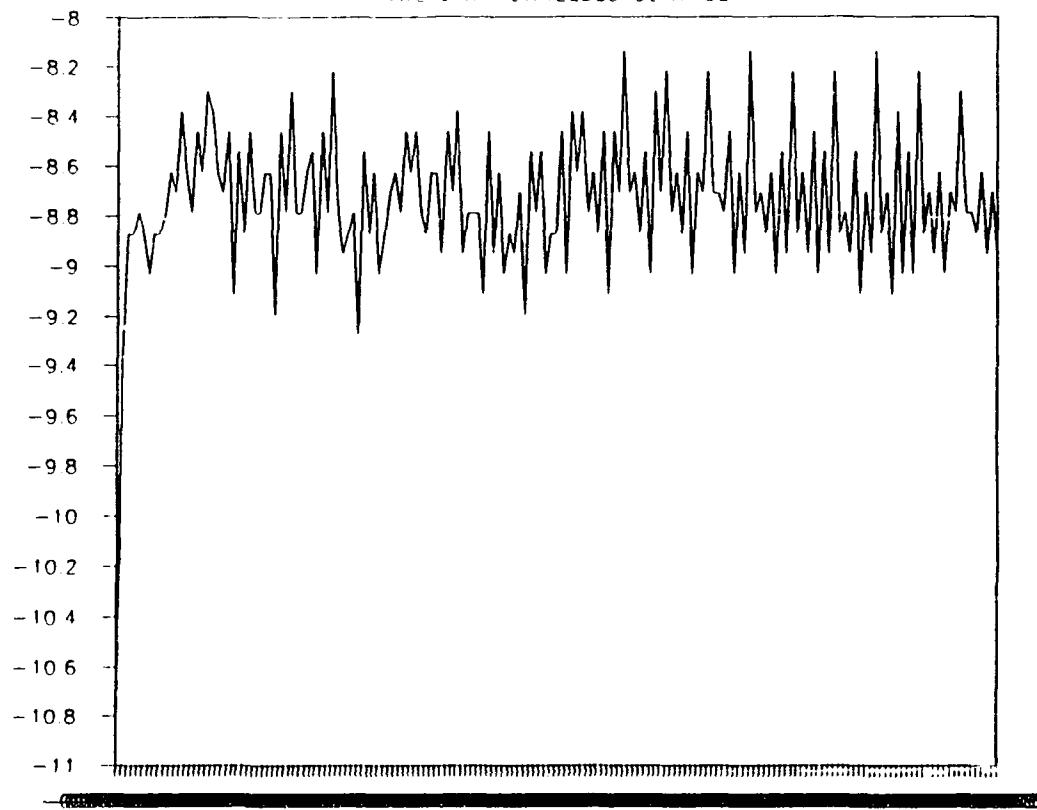
Setting: Current Control

38A, 58W

0.8W delivered
Small Aperture

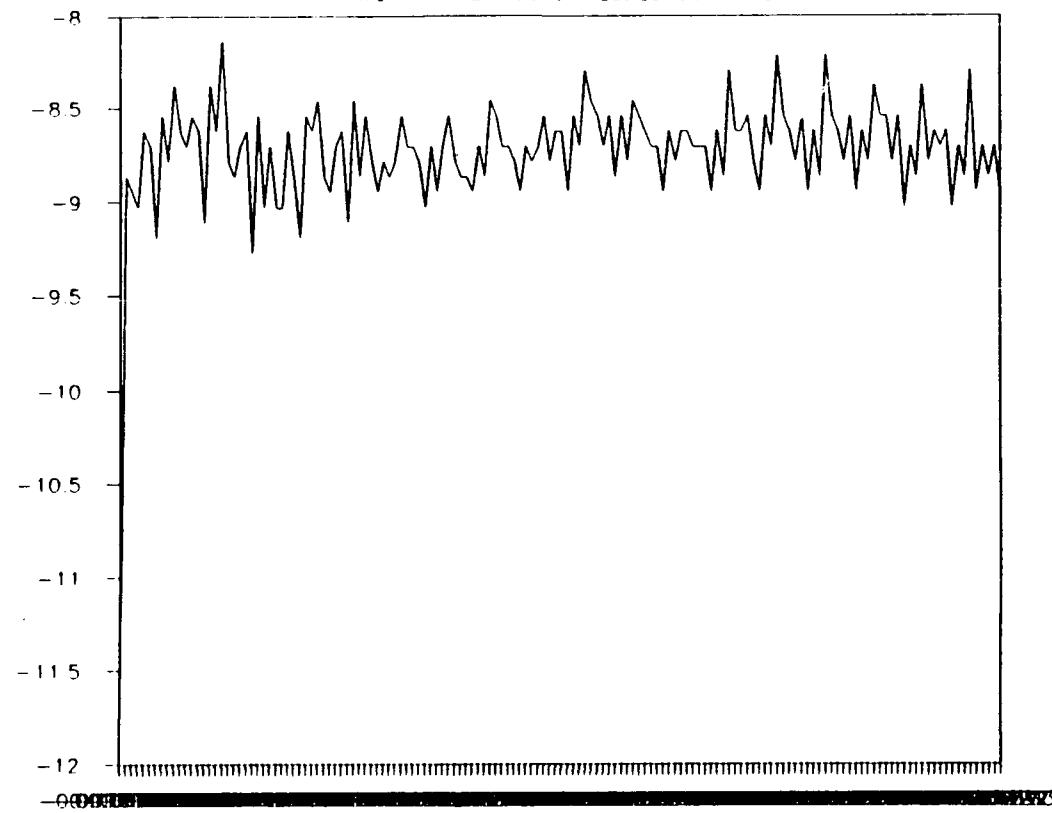
OCT16B

TWO STRIP UNWELDED CONTROL



OCT16C

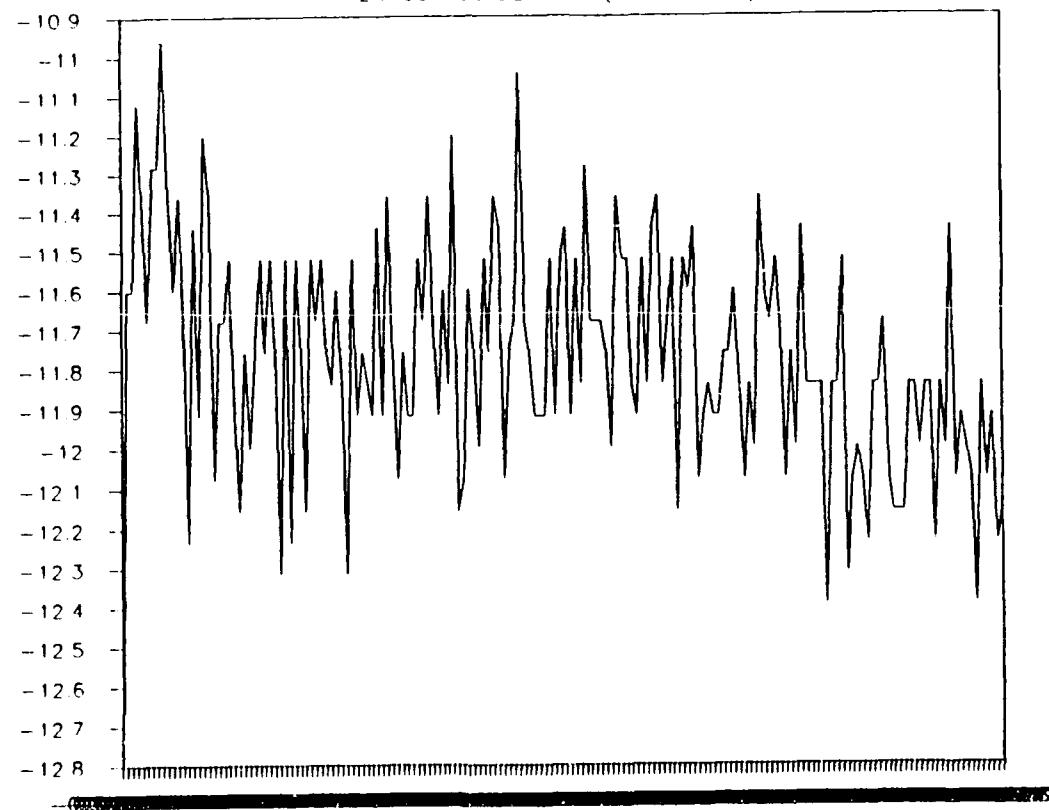
TWO STRIP 2X JCG UNWELDED CONTROL



0.3°

OCT16D1

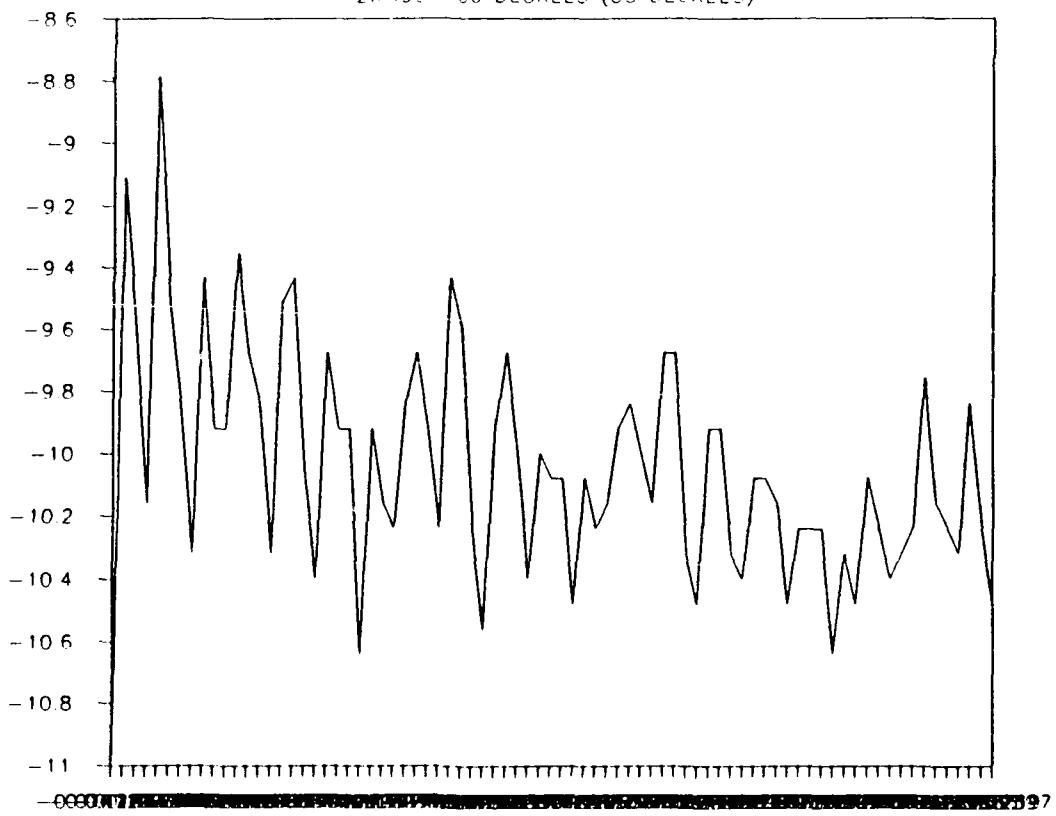
2X ICG - 50 DEGREES (44 DEGREES)

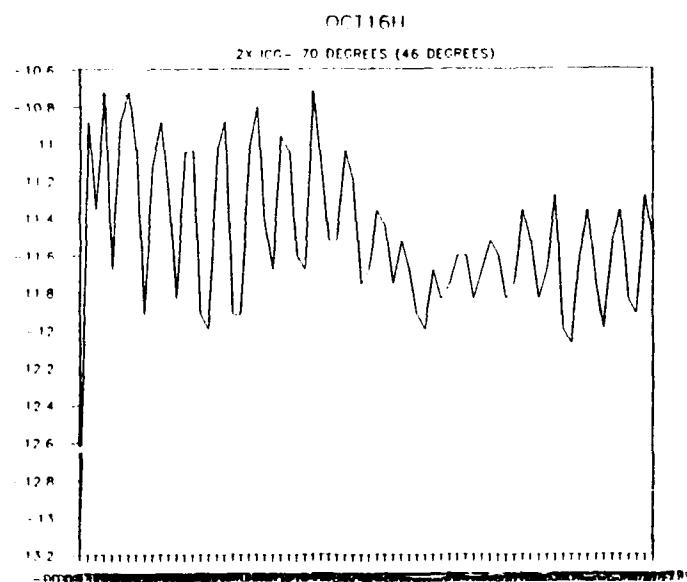


OCT16F

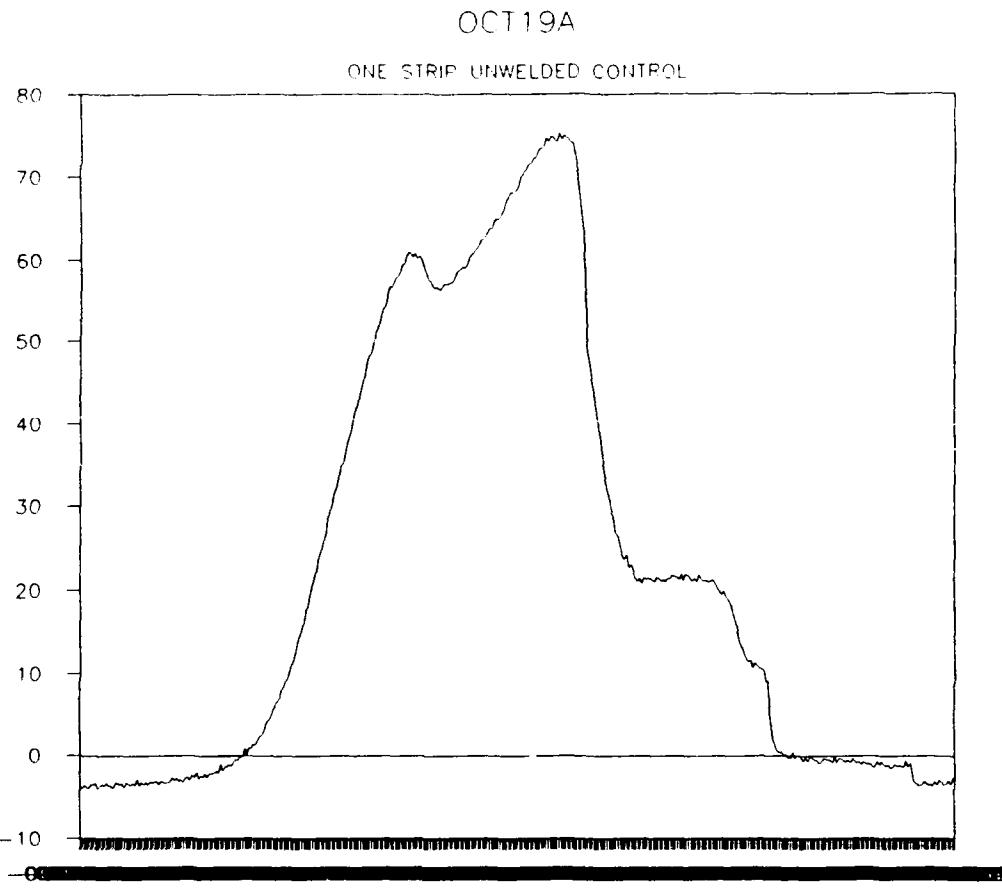
059

2X ICG - 60 DEGREES (53 DEGREES)





C 16



for all trials on OCT 19, 1990

Ar⁺⁺ Laser

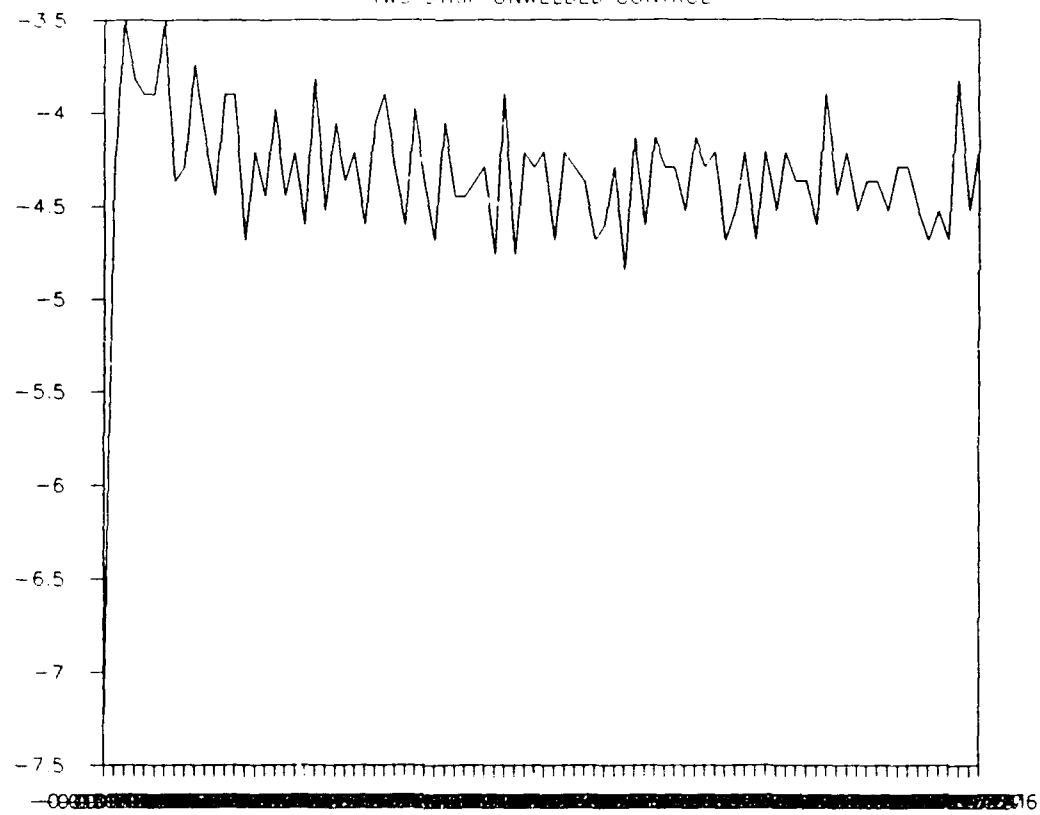
Settings : Current Control

32 A, 6.0 W

0.8 W delivered
Small Aperture

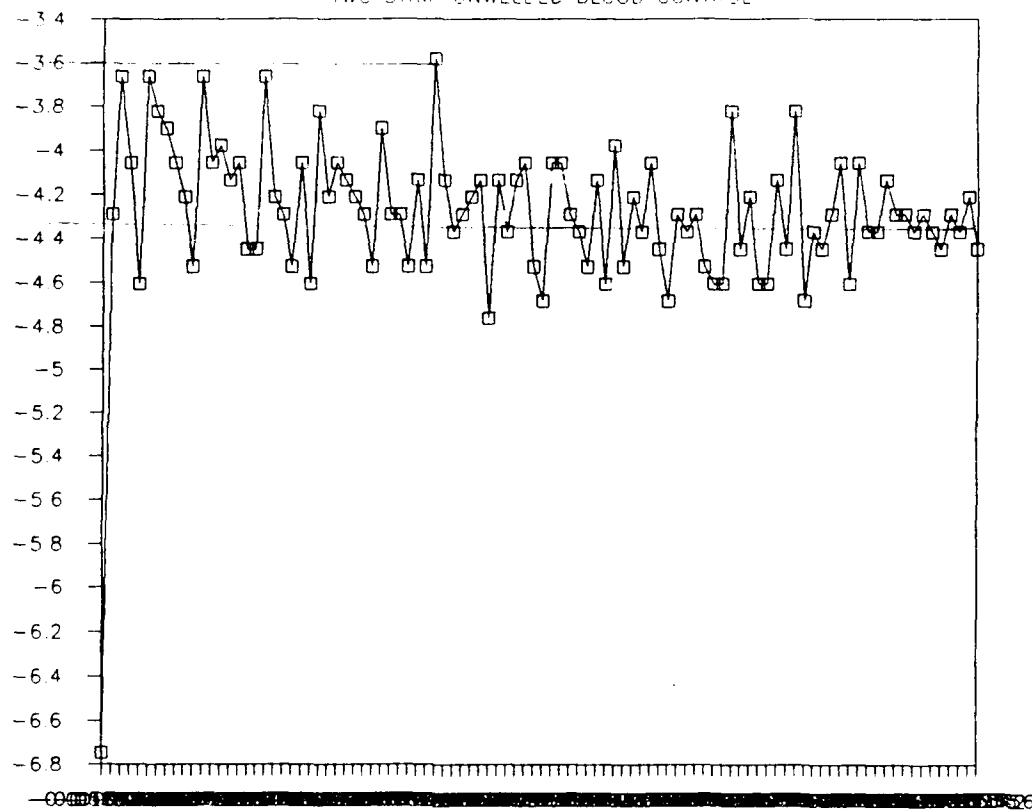
OCT19B

TWO STRIP UNWELDED CONTROL



OCT19C

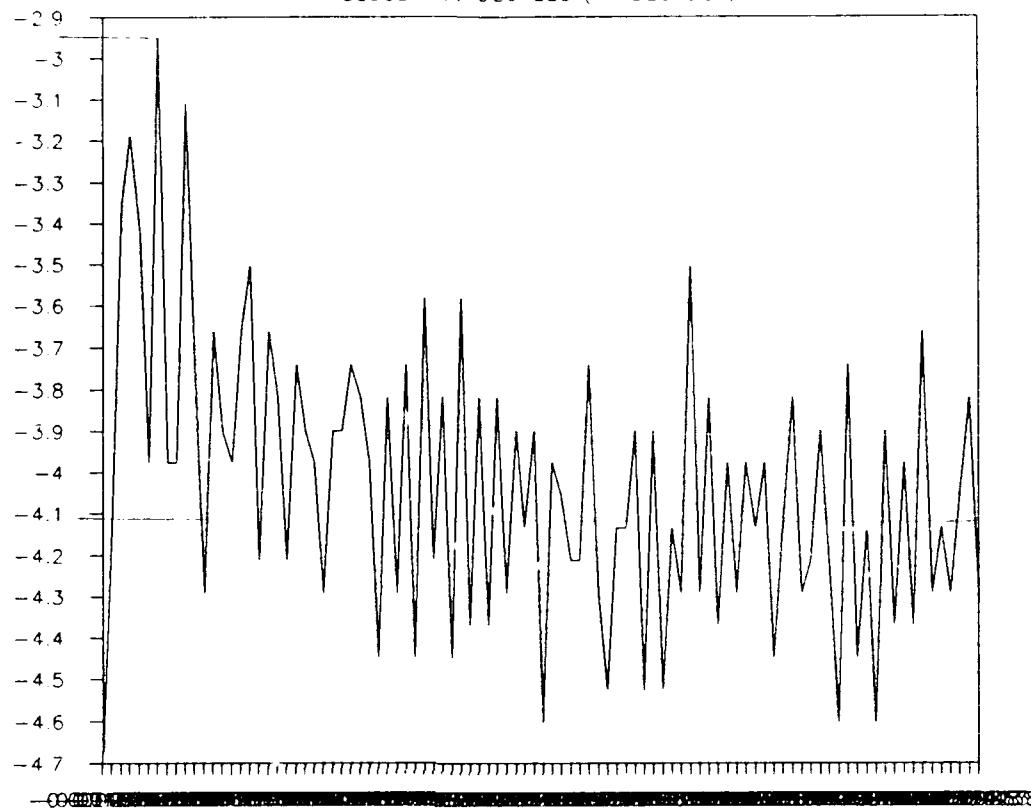
TWO STRIP UNWELDED BLOOD CONTROL

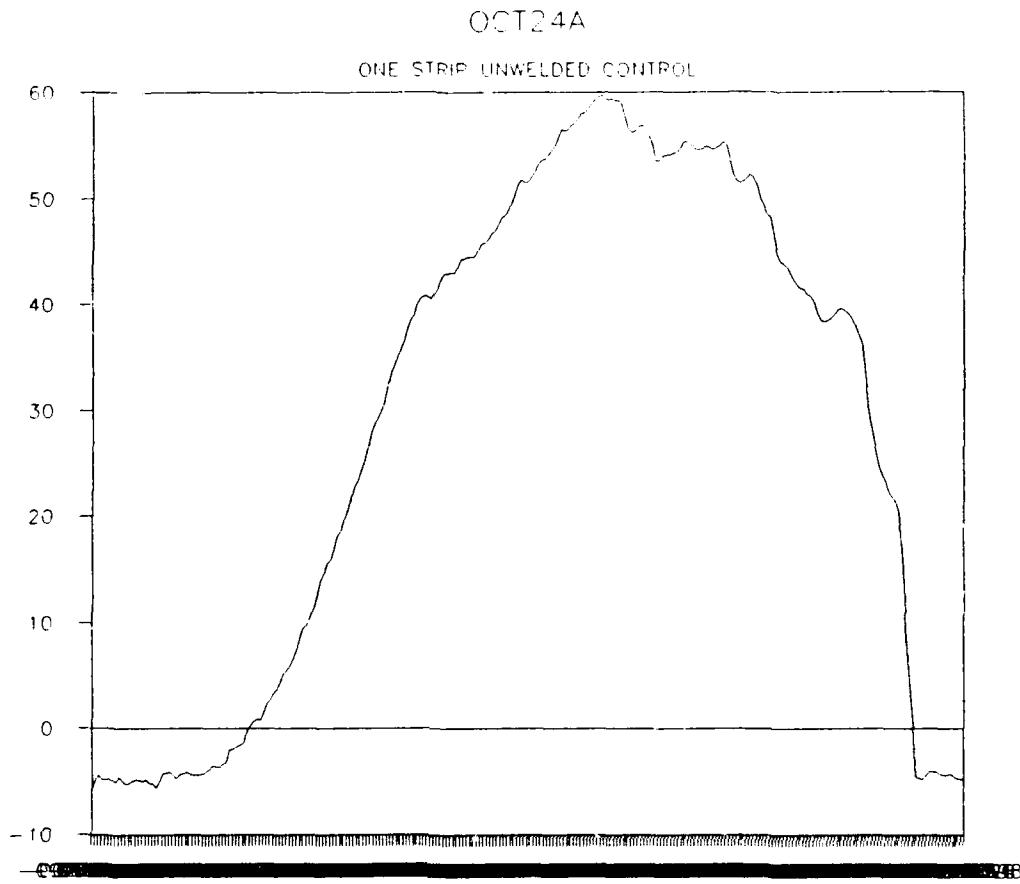


C. S.

OCT19D1

BLOOD - 50 DEGREES (44 DEGREES)





For all trials on OCT 24, 1996

Ar⁺⁺ Laser

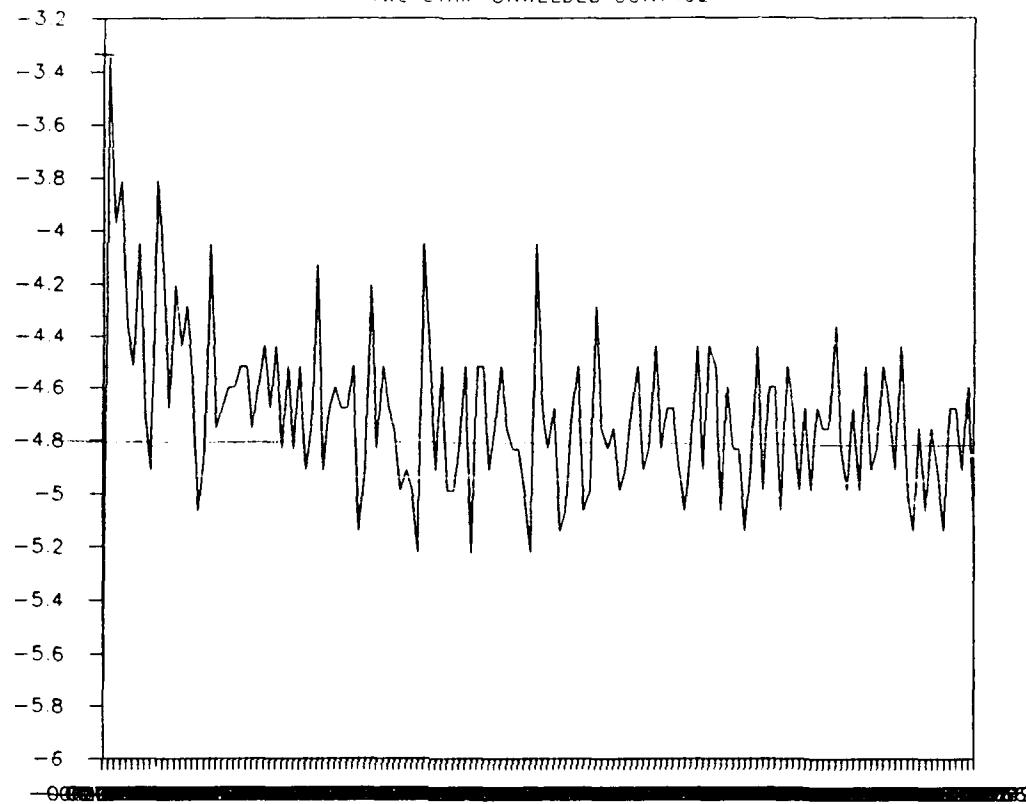
Setting: Current control

38A, 5.6W

0.7 W delivered
Small Aperture

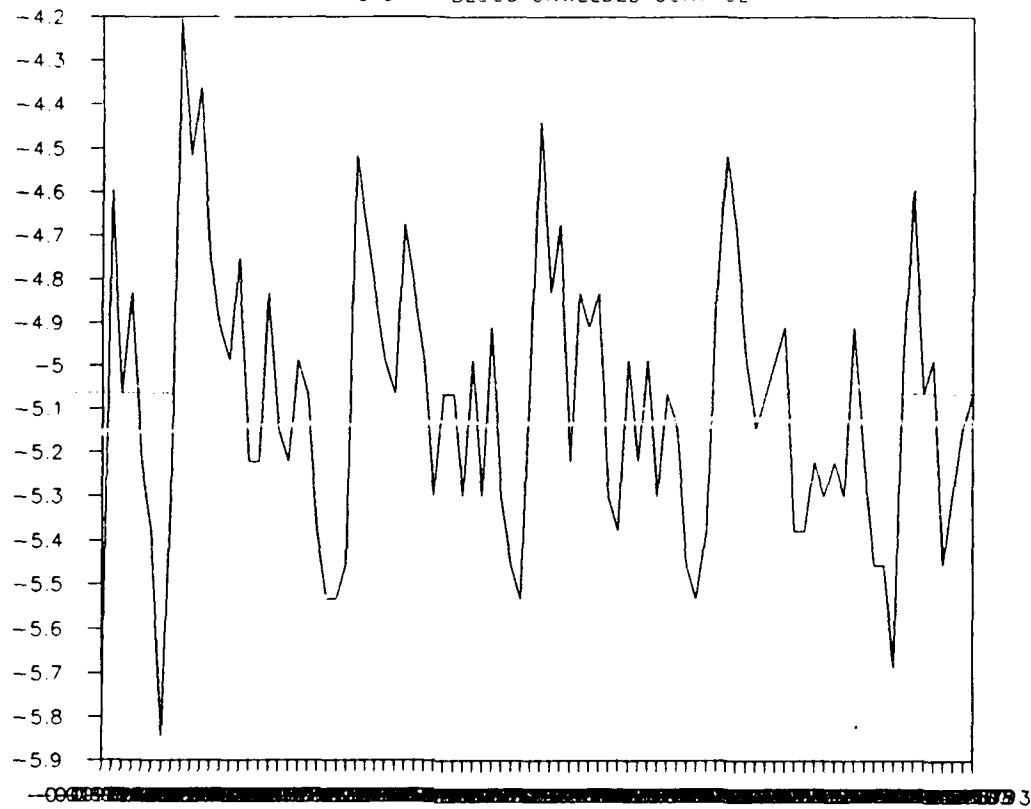
OCT24B

TWO STRIP UNWELDED CONTROL



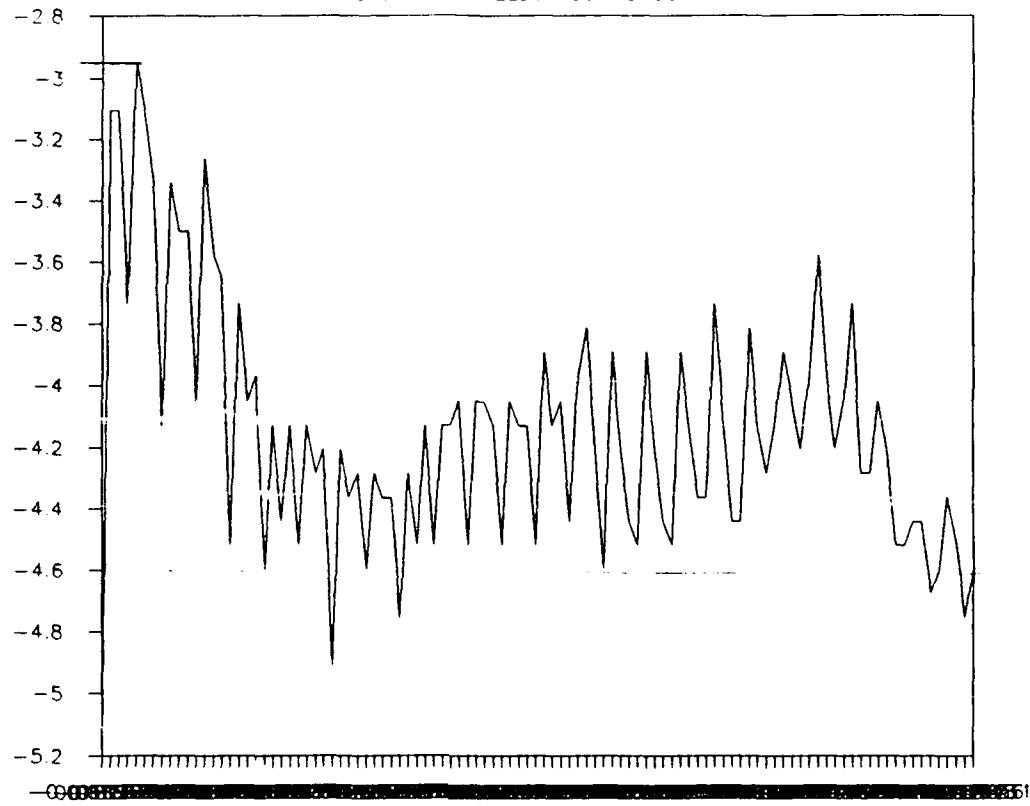
OCT24C

TWO STRIP BLOOD UNWELDED CONTROL



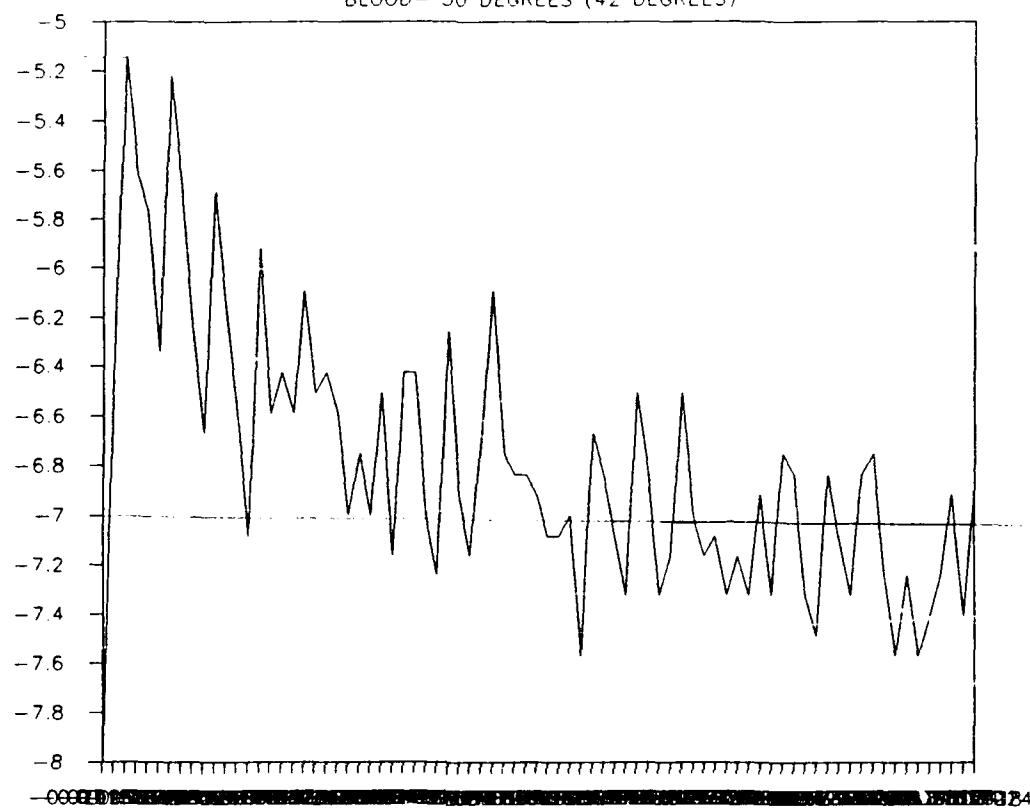
OCT24D

TWO STRIP UNWELDED BLOOD CONTROL



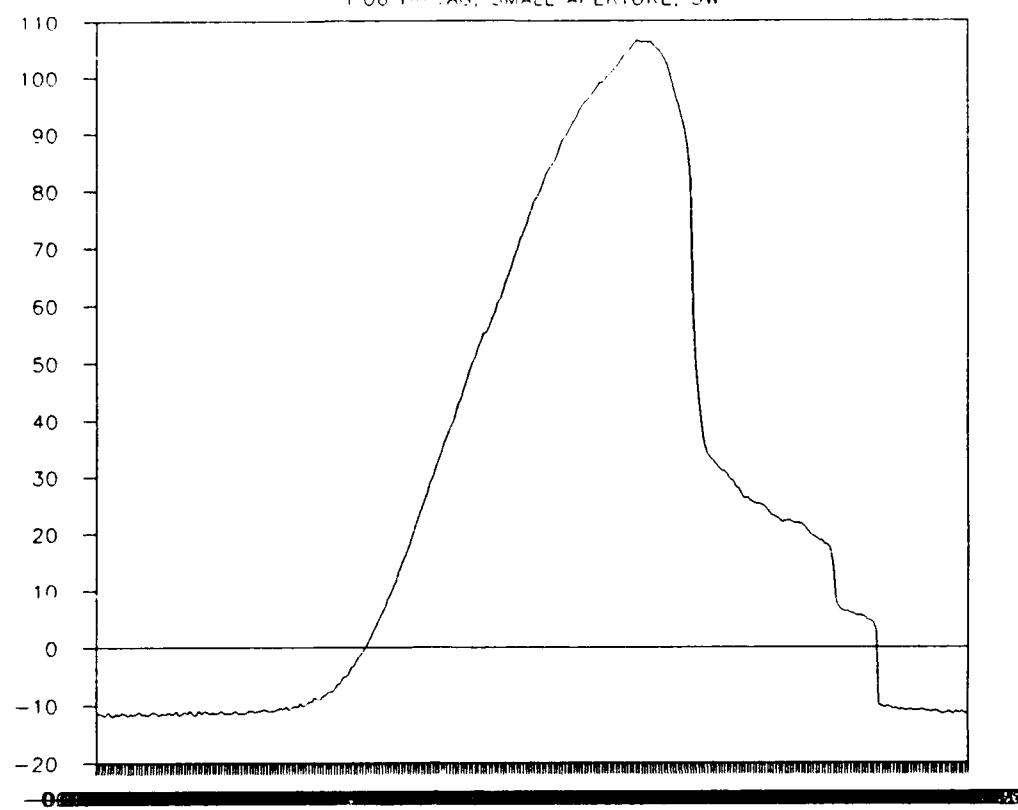
OCT24E1

BLOOD - 50 DEGREES (42 DEGREES)



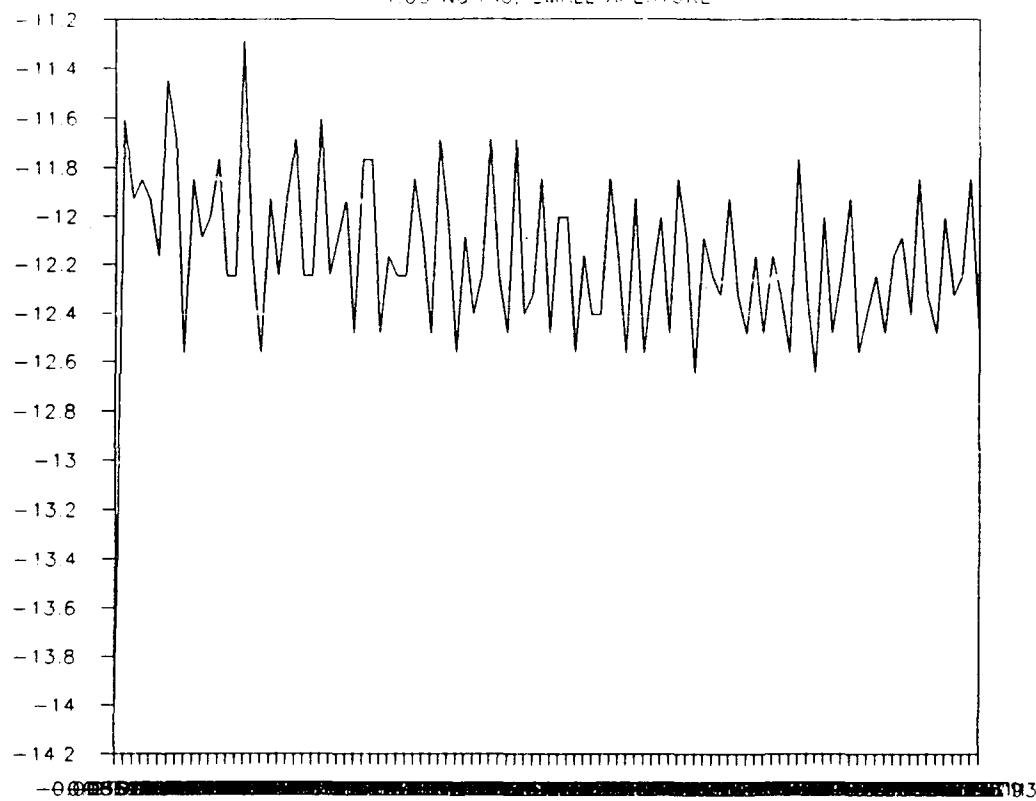
NOV8A- ONE STRIP CONTROL

1.06 NH-YAG, SMALL APERTURE, 5W



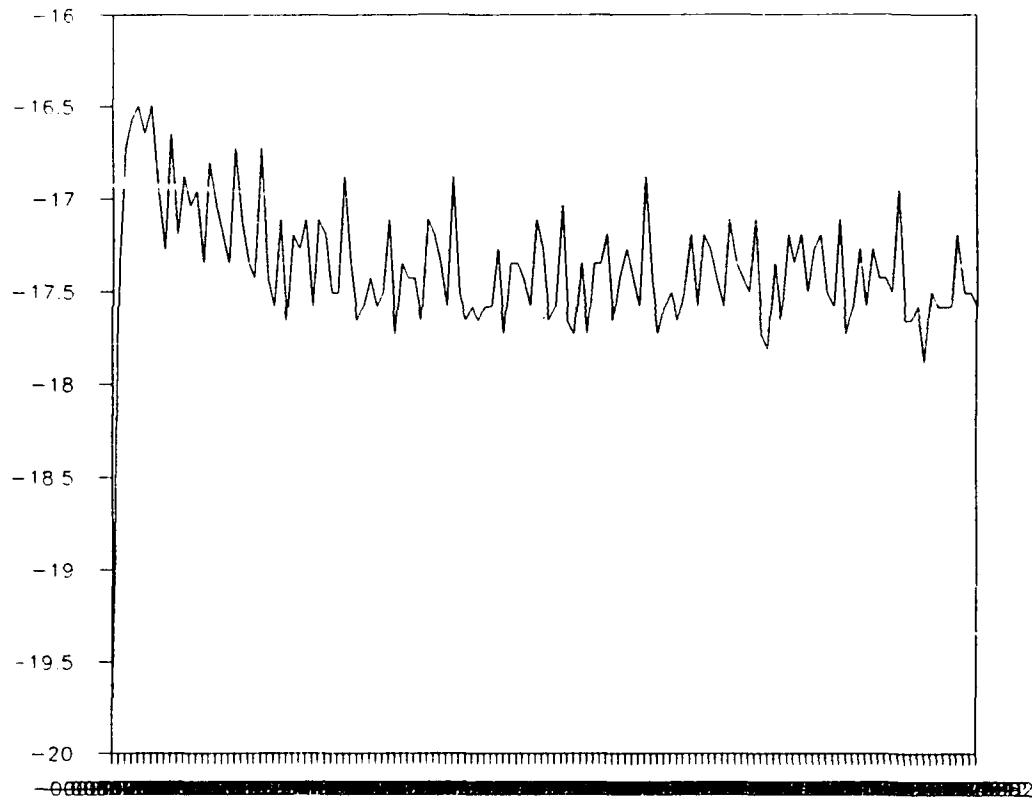
NOV8C - INDIA INK UNWELDED CONTROL

1.06 Nd YAG, SMALL APERTURE



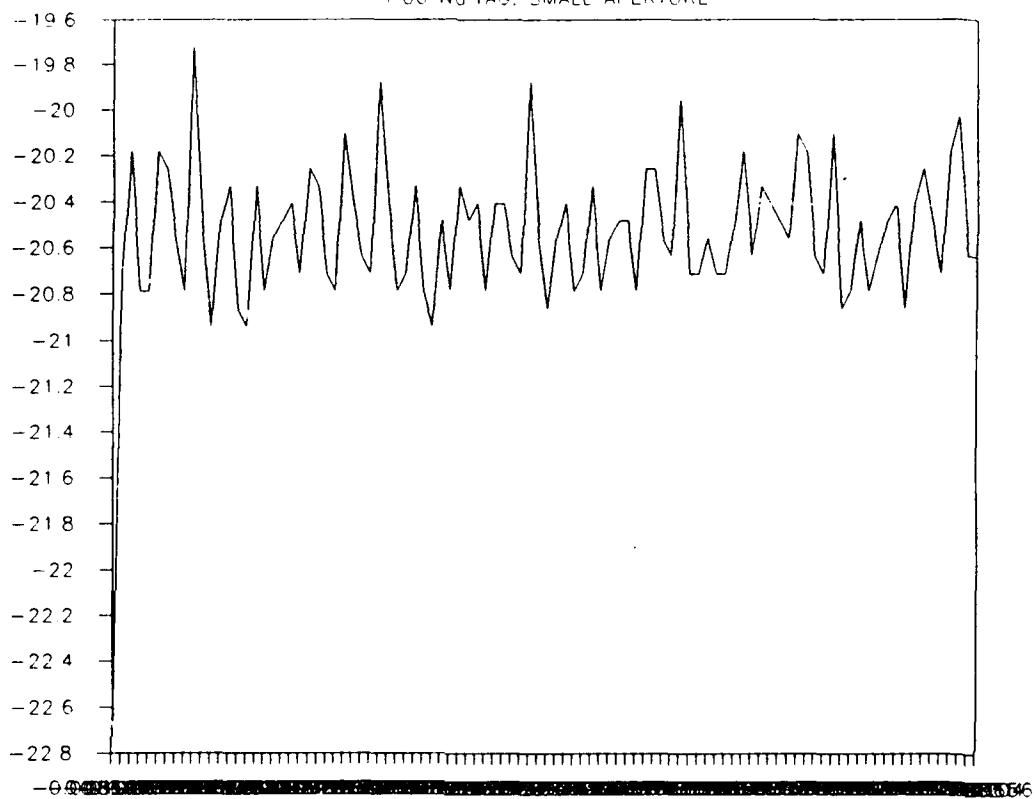
NOV8D- INDIA INK CONTROLLED AT 50

106 Nd:AG SMALL APERTURE



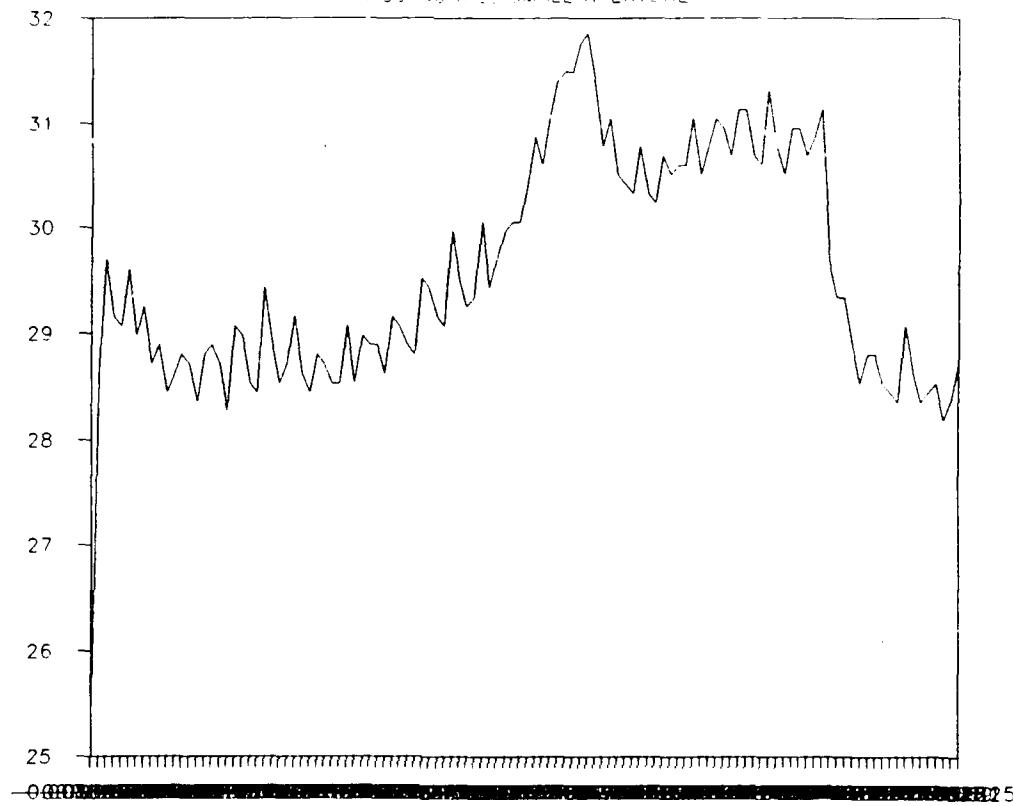
NOV3E - INDIA INK CONTROLLED AT 60

1.06 Nd YAG, SMALL APERTURE



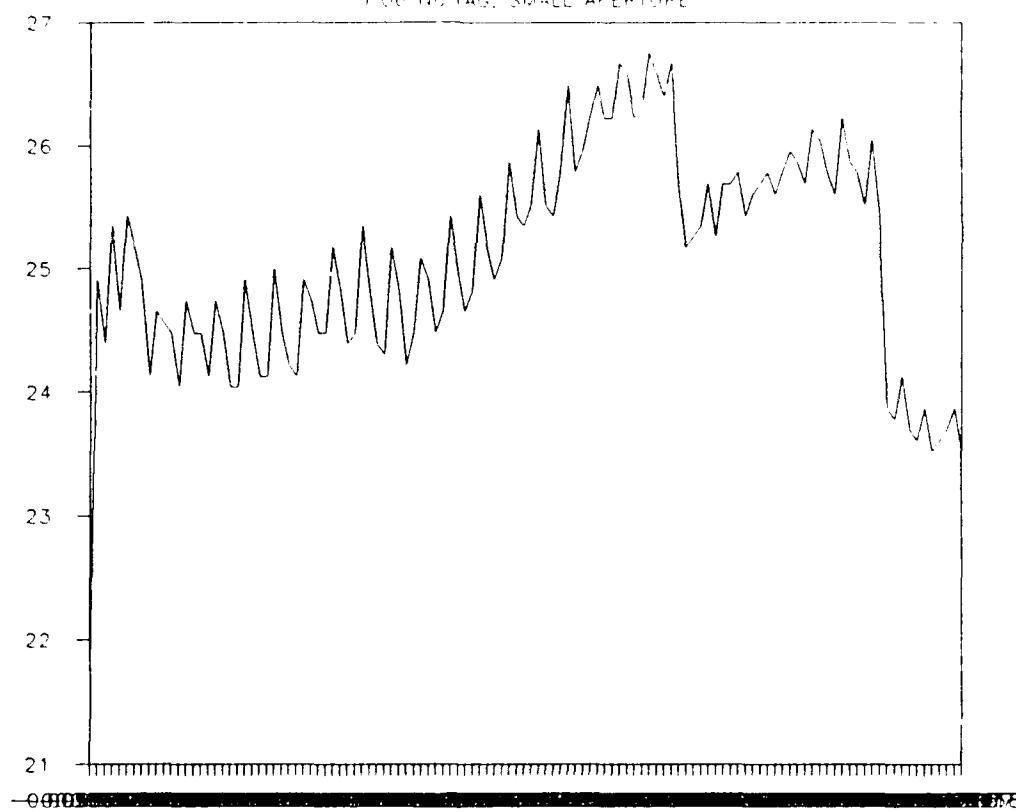
NOV8F-INDIA INK CONTROLLED AT 70

106 Nd:YAG, SMALL APERTURE



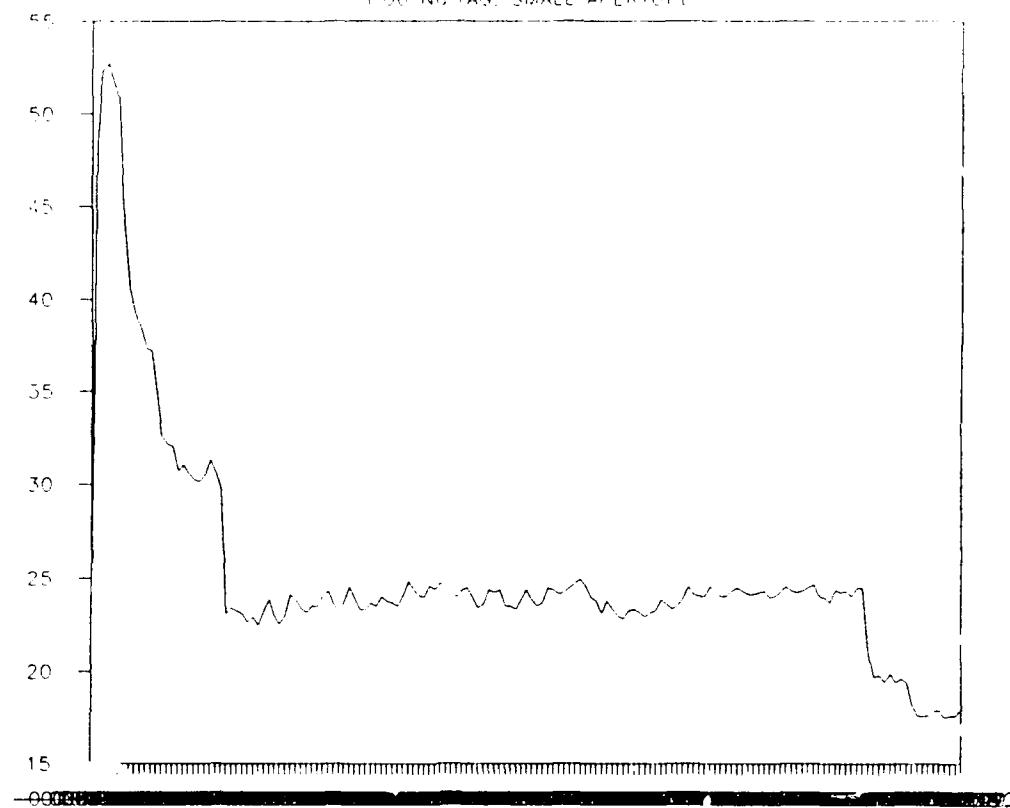
NOV3G-INDIA INK CONTROLLED AT 80

1.0E-11 FAS, SMALL APERTURE



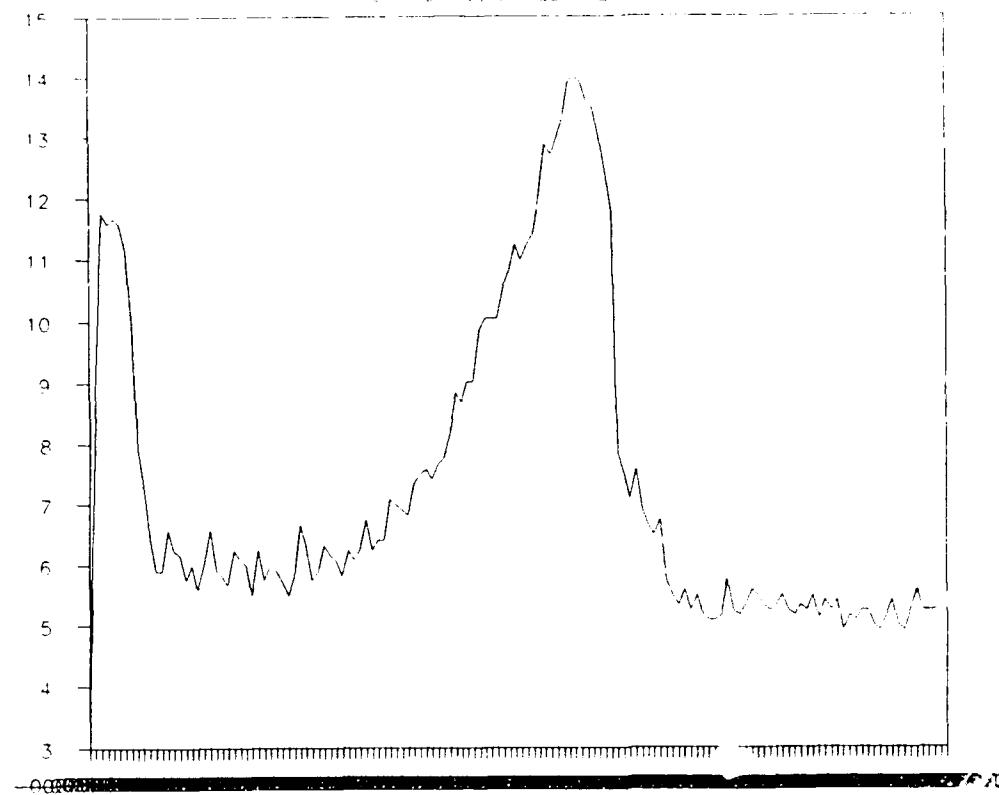
NOV8H-INDIV. INK CONTROLLED AT 100

1.06 ND YAG, SMALL APERTURE



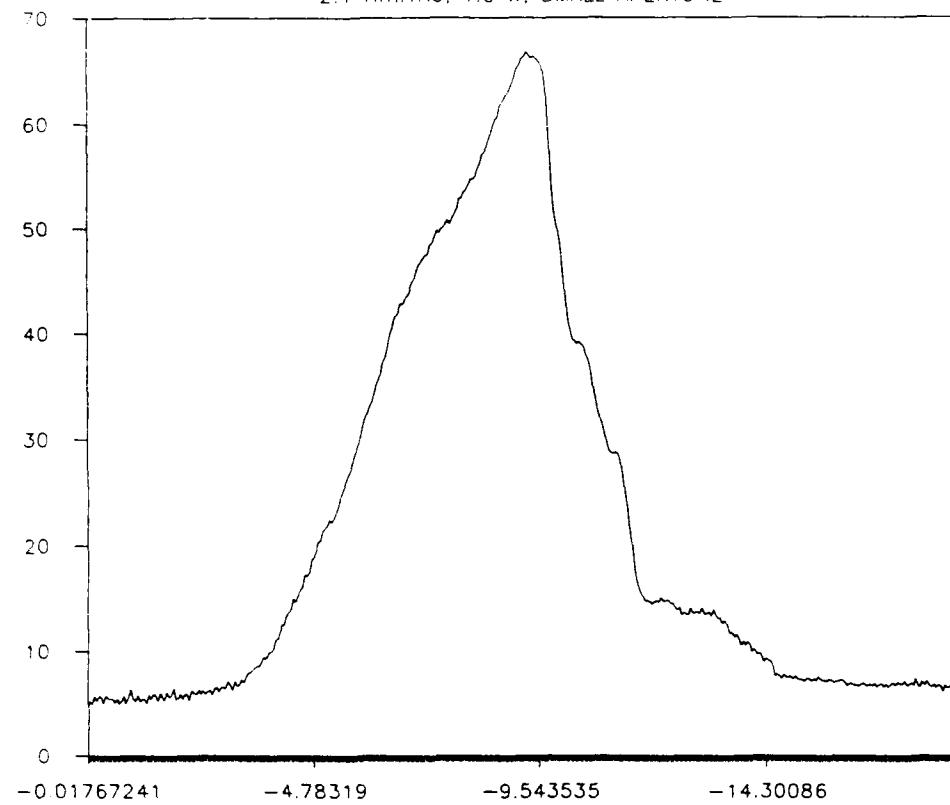
NOV81-INDIA INK CONTROLLED AT 100

1.06 MICR. SMALL APERTURE



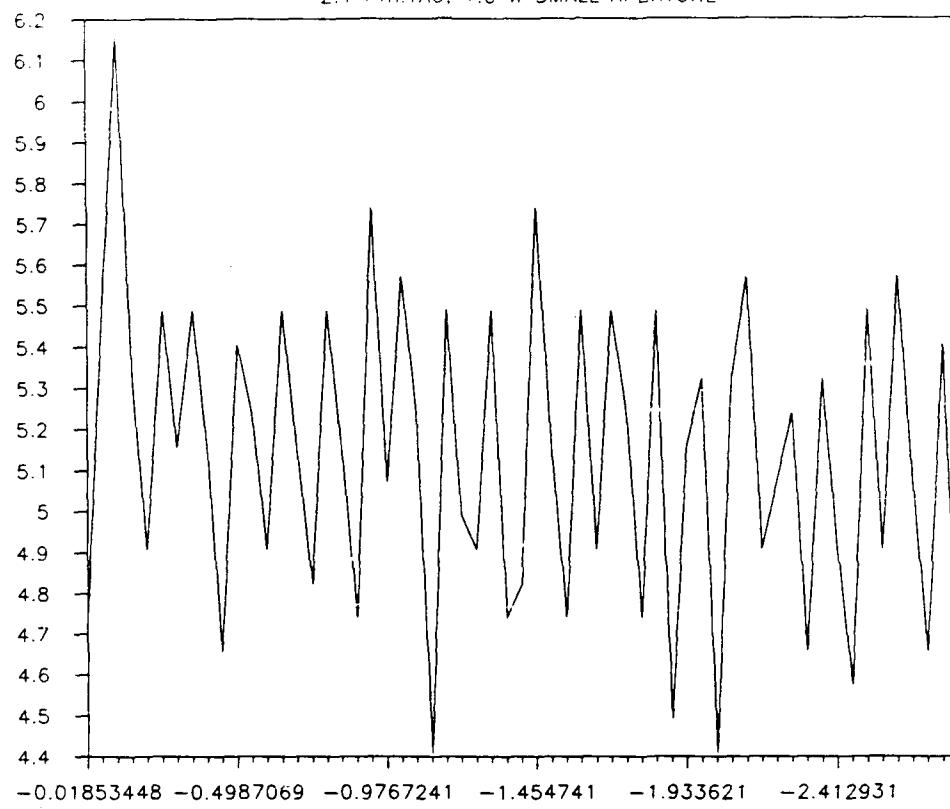
DEC17A—ONE STRIP UNWELDED CONTROL

2.1 Hm:YAG, 1.0 W, SMALL APERTURE



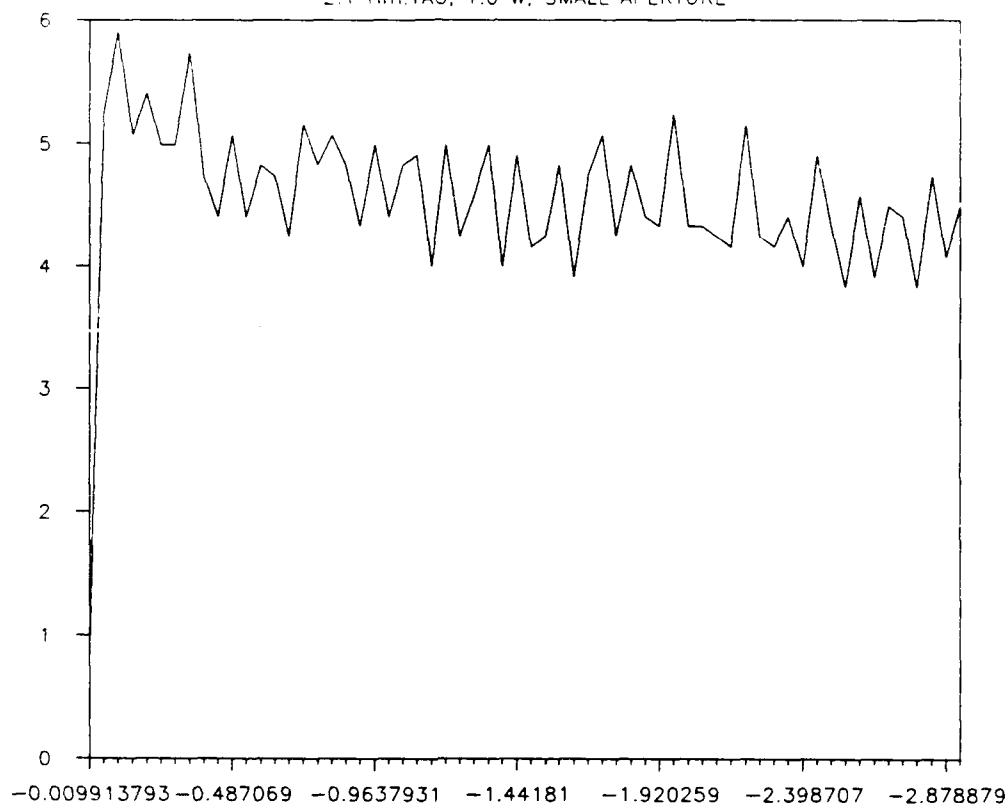
DEC17B-TWO STRIP UNWELDED CONTROL

2.1 Hz YAG, 1.0 W SMALL APERTURE



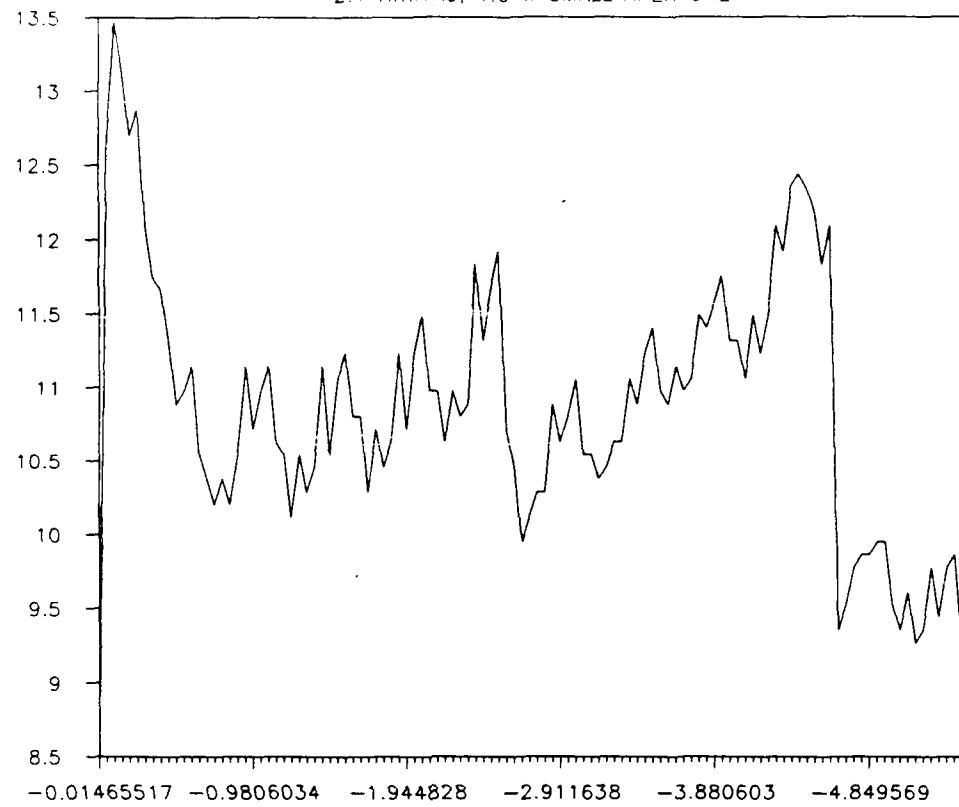
DEC17C-TWO STRIP INDIA INK UNWELDED

2.1 Hm:YAG, 1.0 W, SMALL APERTURE



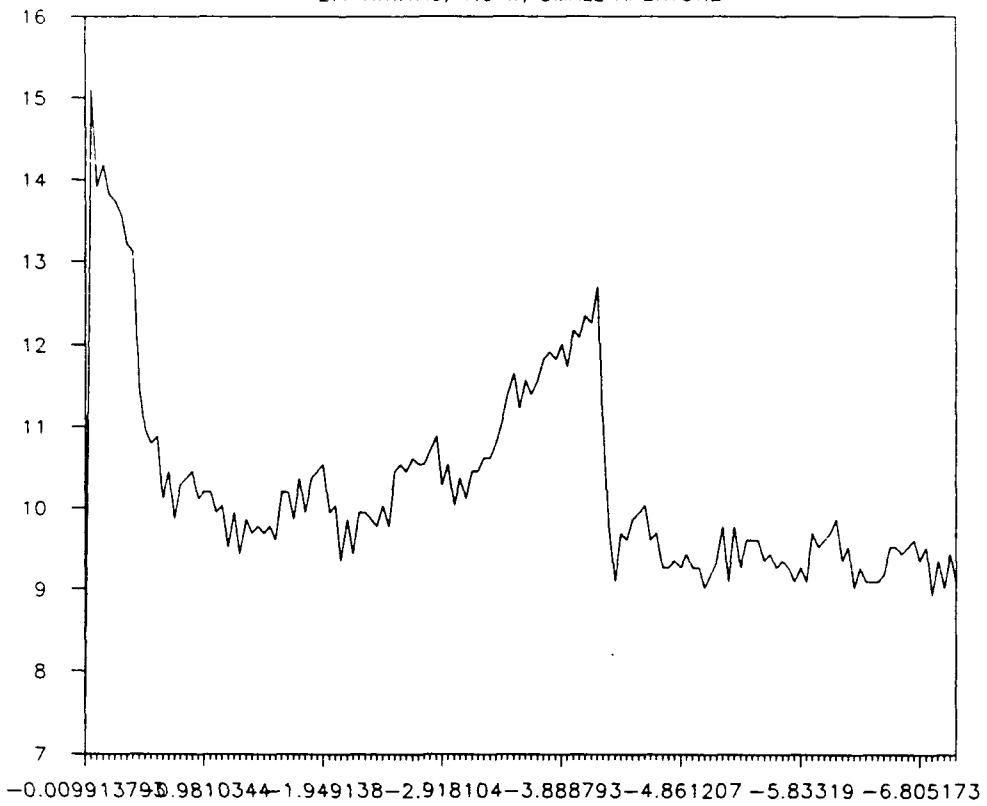
DEC17E-INDIA INK CONTROLLED AT 100

2.1 Hm:YAG, 1.0 W SMALL APERTURE



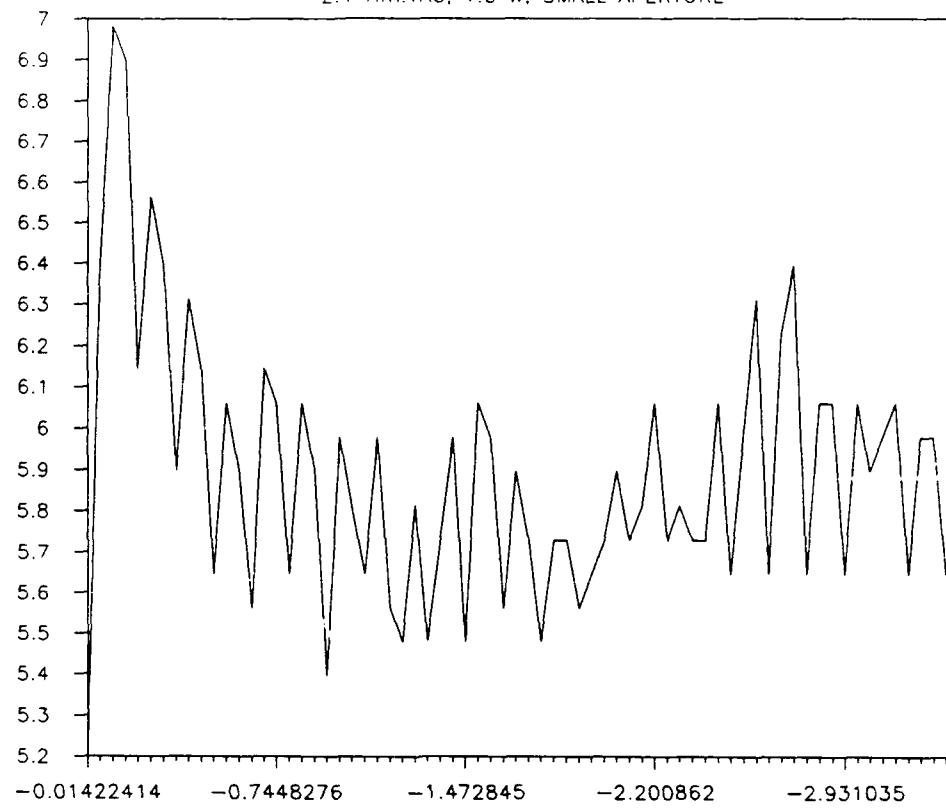
DEC17F - NO CHROMO SET=100, TC MAX=70

2.1 Hm:YAG, 1.0 W, SMALL APERTURE



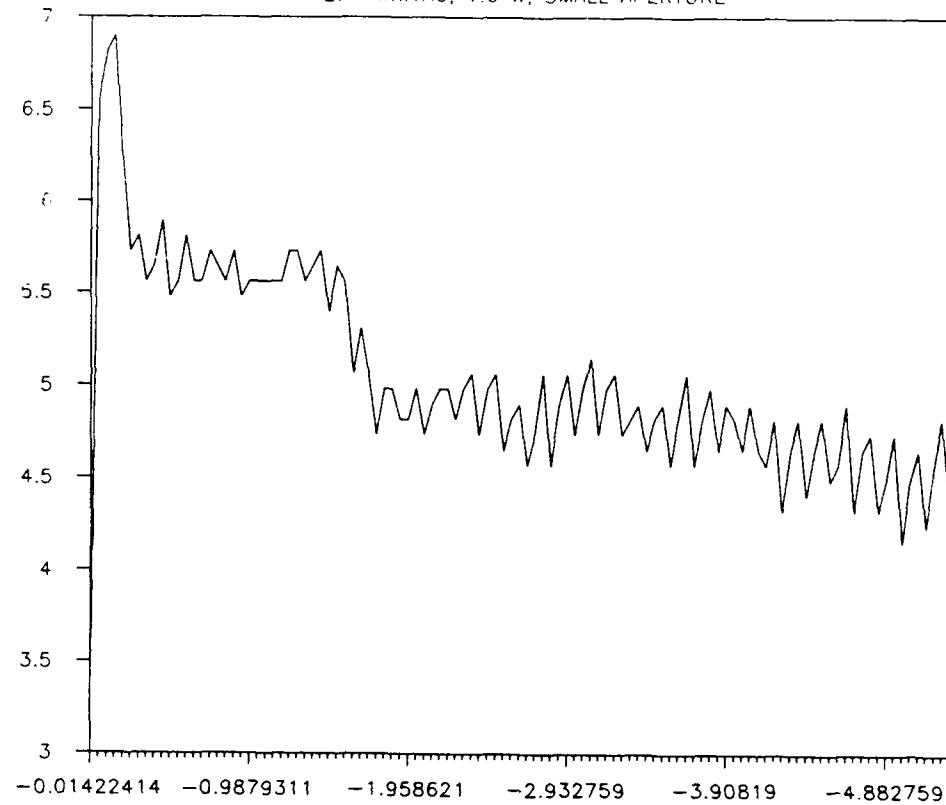
DEC17G-INDIA INK CONTROLLED AT 50, 10s

2.1 Hz:YAG, 1.0 W, SMALL APERTURE



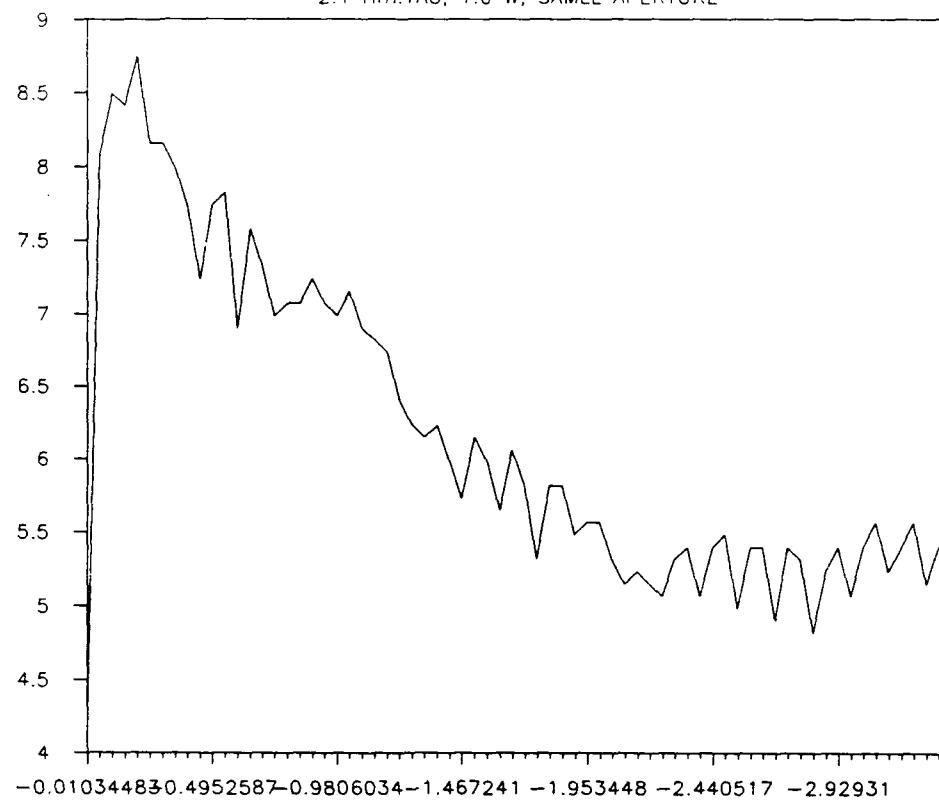
DEC17H-INDIA INK CONTROLLED AT 60, 10 s

2.1 Hm:YAG, 1.0 W, SMALL APERTURE



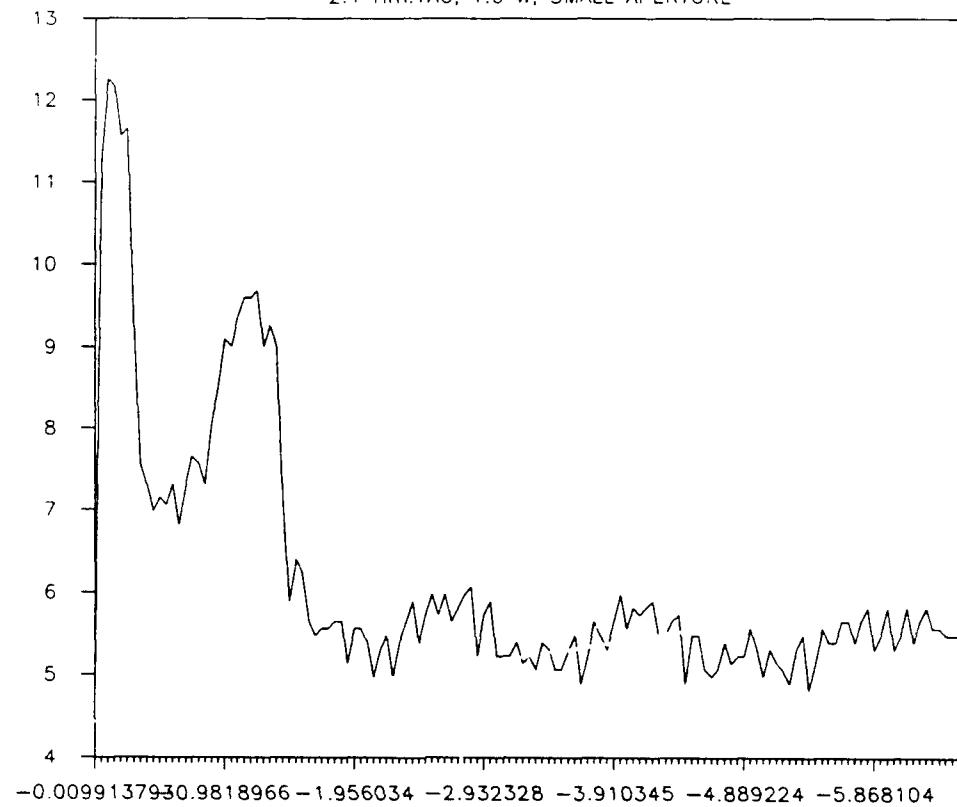
DEC17I-INDIA INK CONTROLLED AT 70, 10 s

2.1 Hm:YAG, 1.0 W, SAMLL APERTURE



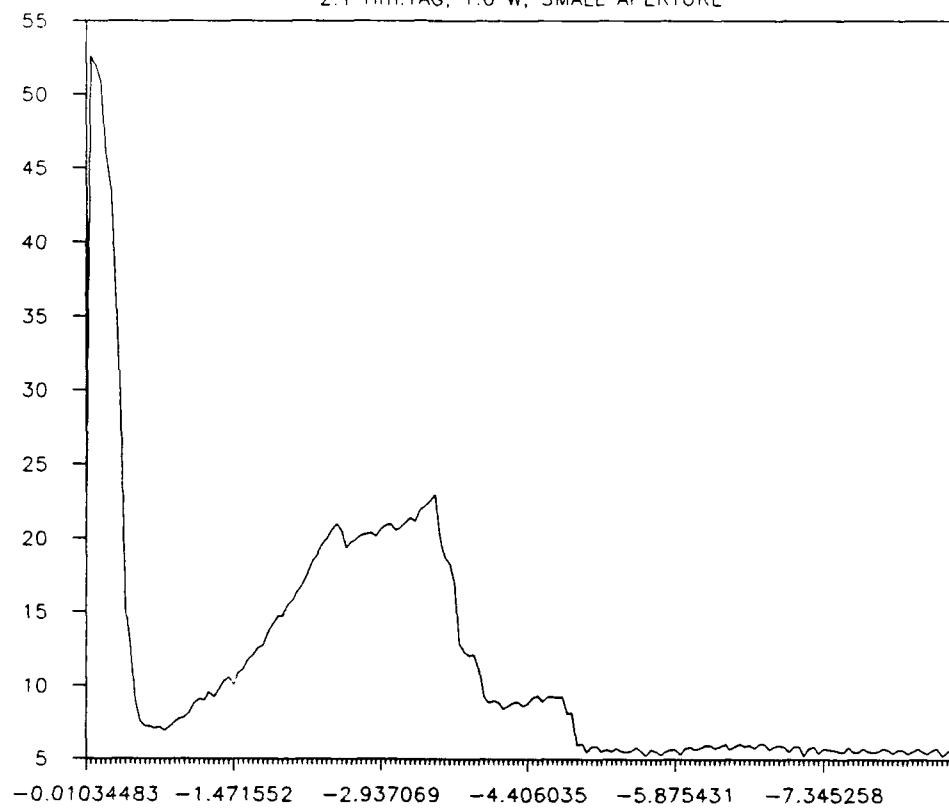
DEC17J-INDIA INK CONTROLLED AT 80, 10 s

2.1 Hm:YAG, 1.0 W, SMALL APERTURE



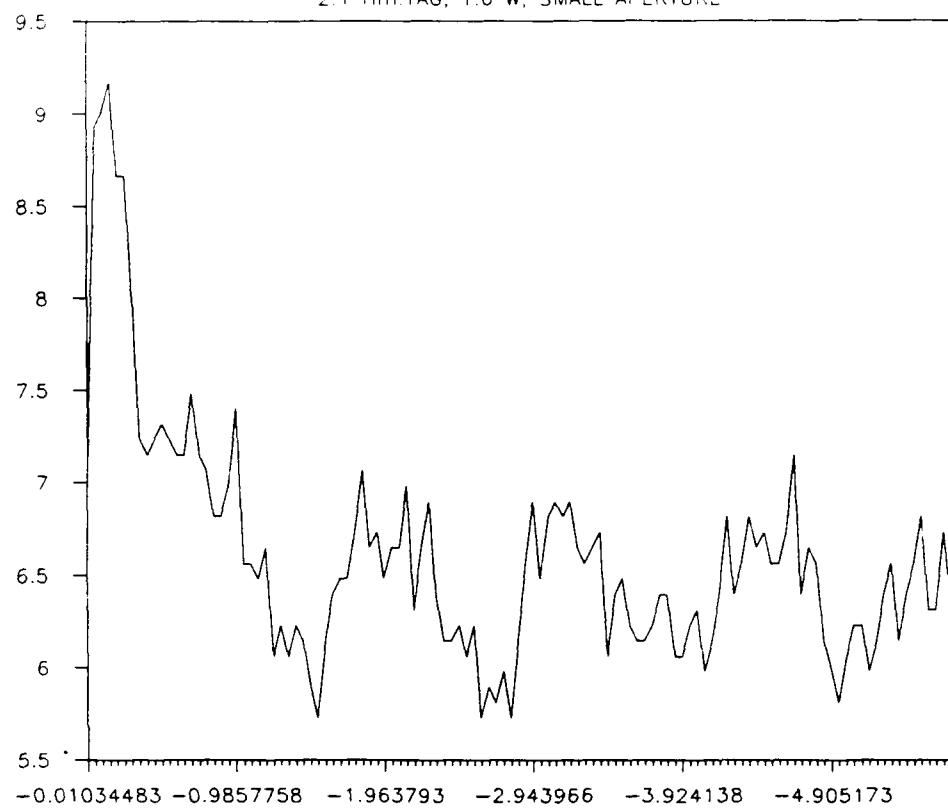
DEC17K-INDIA INK CONTROLLED AT 100, 10s

2.1 Hm:YAG, 1.0 W, SMALL APERTURE



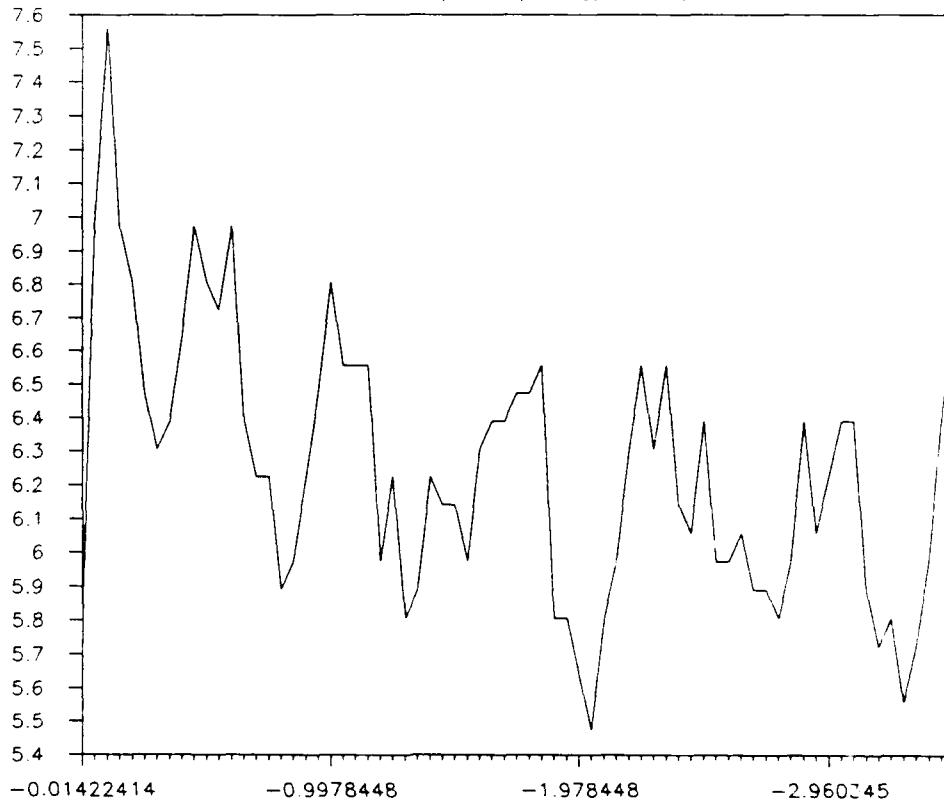
DEC17L-INDIA INK CONTROLLED AT 100, 10s

2.1 Hm:YAG, 1.0 W, SMALL APERTURE



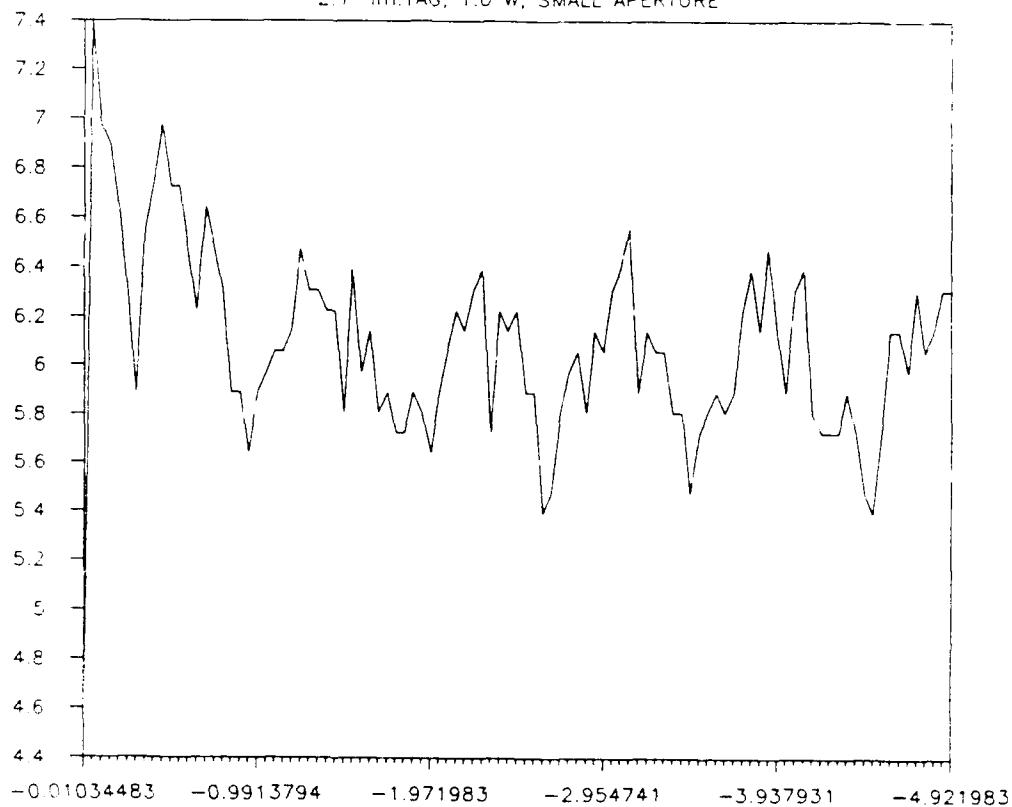
DEC17M-NO CHROMO, CONTROLLED AT 50, 10s

2.1 Hm:YAG, 1.0 W, SMALL APERTURE



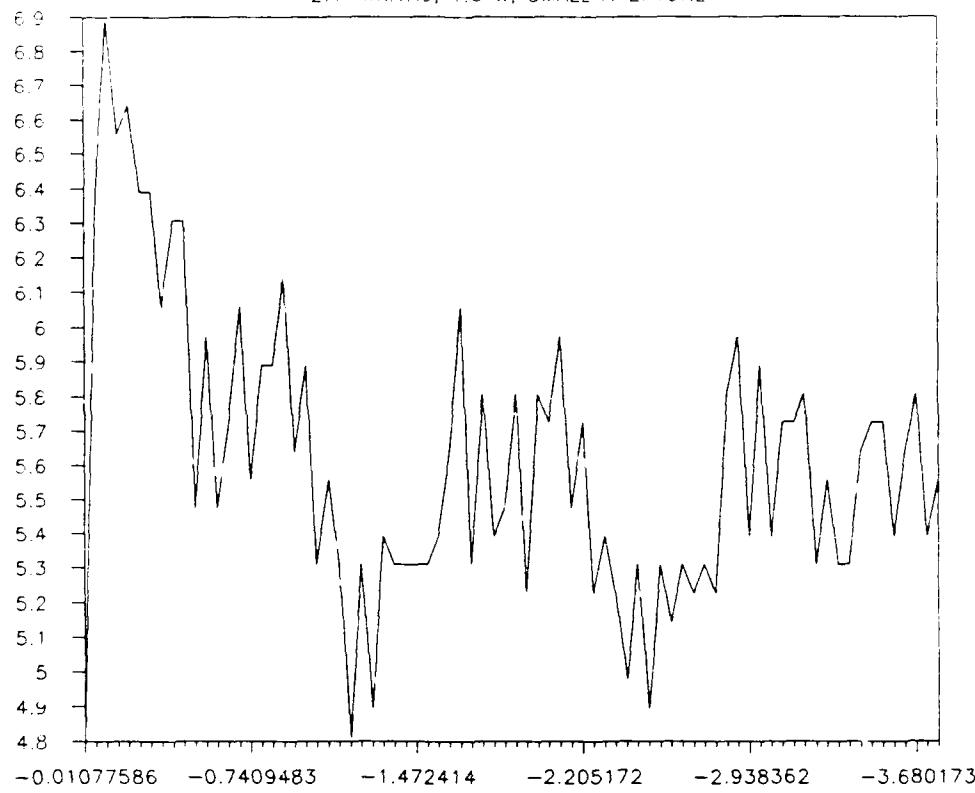
DEC17N-NO CHROMO CONTROLLED AT 60, 10s

2.1 nm:YAG, 1.0 W, SMALL APERTURE



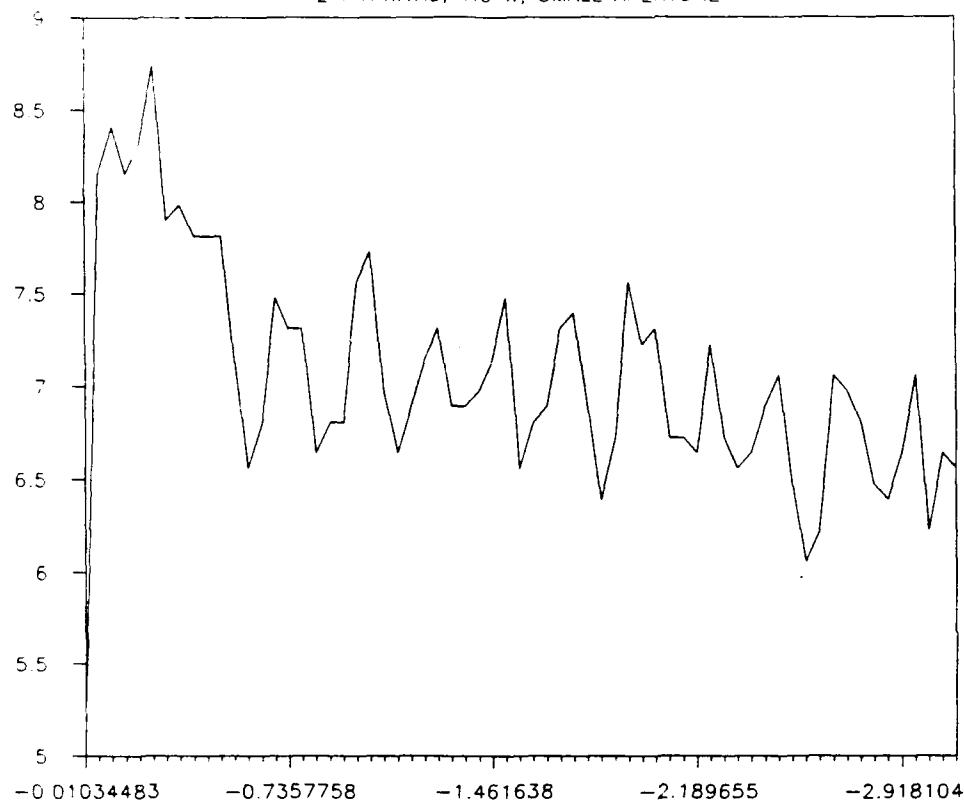
DEC170-NO CHROMO CONTROLLED AT 70, 10s

2.1 Hz:YAG, 1.0 W, SMALL APERTURE



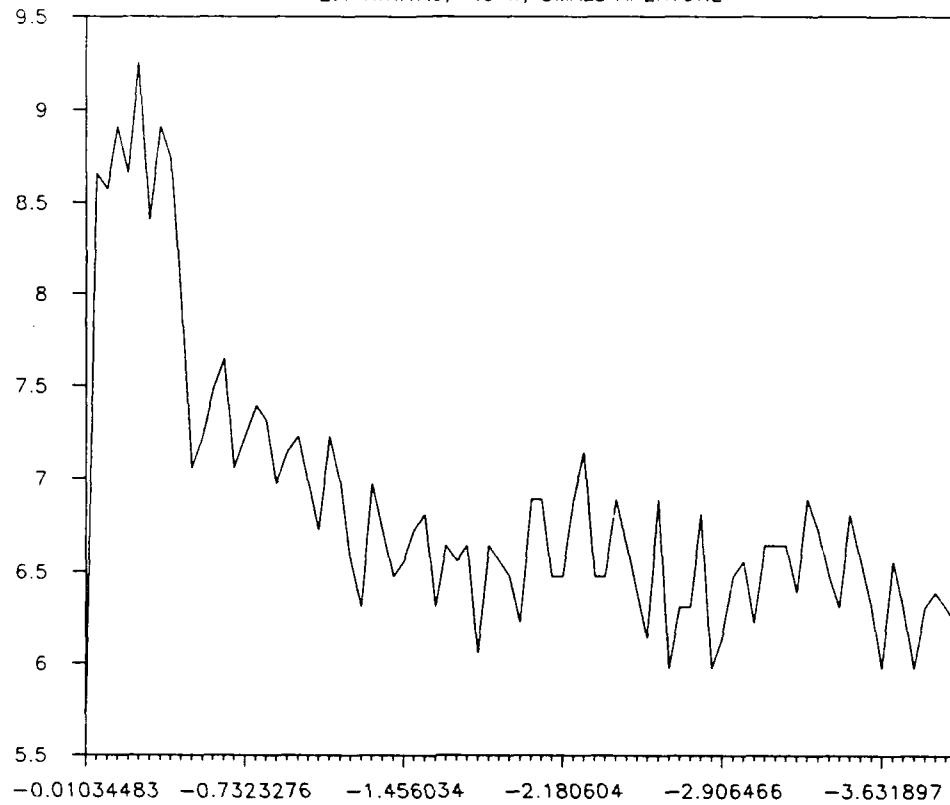
DEC17P-NO CHROMO CONTROLLED AT 80, 10 s

21 Hz:YAG, 1.0 W, SMALL APERTURE



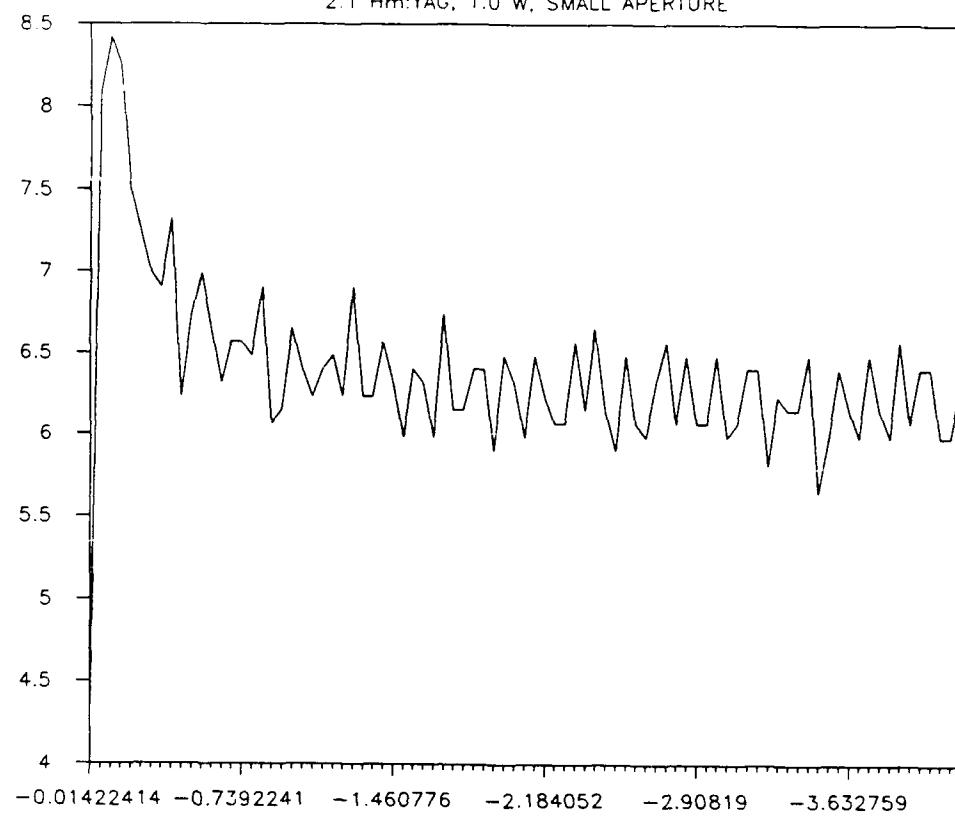
DEC17Q-NO CHROMO CONTROLLED AT 100, 10s

2.1 Hm:YAG, 1.0 W, SMALL APERTURE



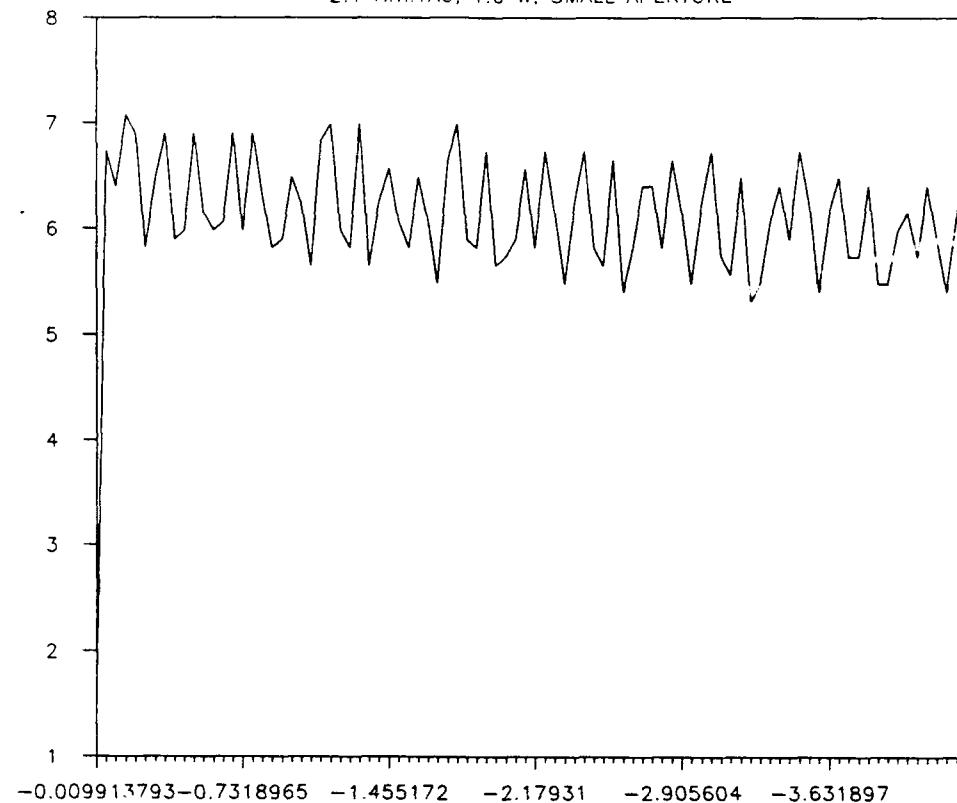
DEC17R-2X ICG CONTROLLED AT 50, 10s

2.1 Hm:YAG, 1.0 W, SMALL APERTURE



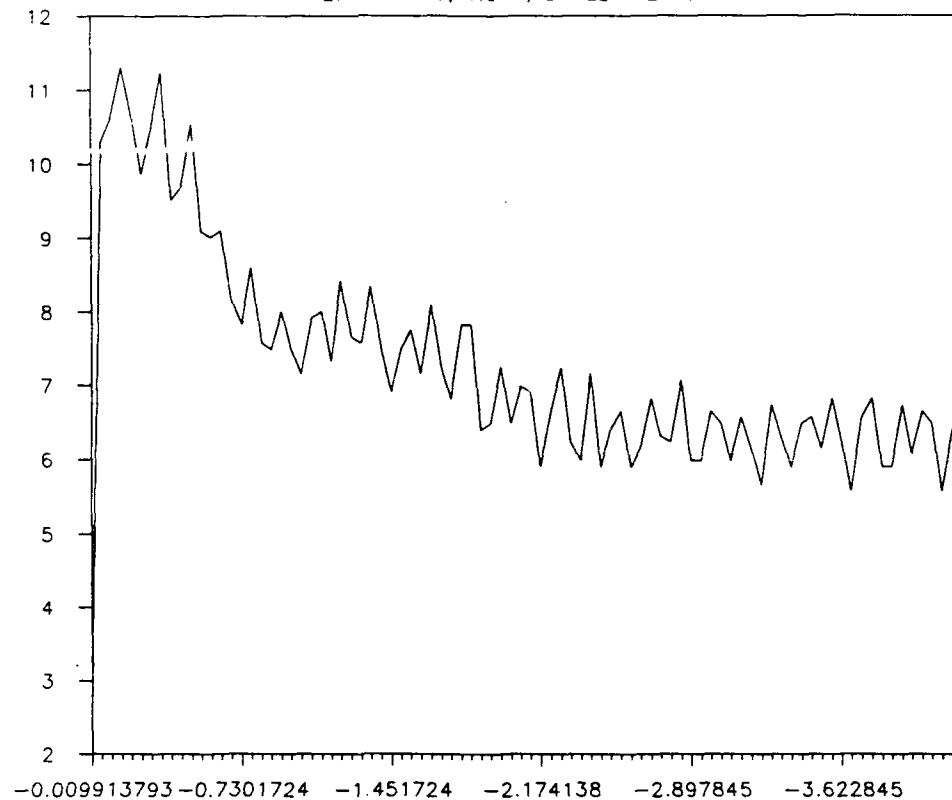
DEC17S- 2X ICG CONTROLLED AT 60, 10s

2.1 Hm:YAG, 1.0 W, SMALL APERTURE



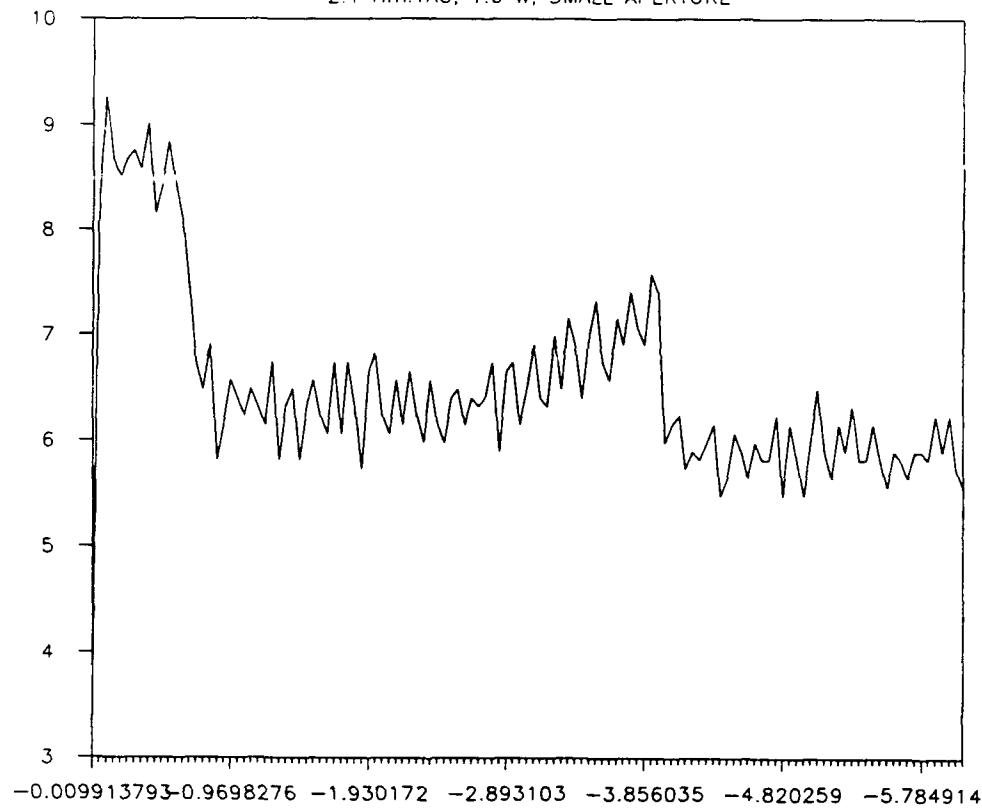
DEC17T-2X ICG CONTROLLED AT 70, 10s

2.1 Hm:YAG, 1.0 W, SMALL APERTURE



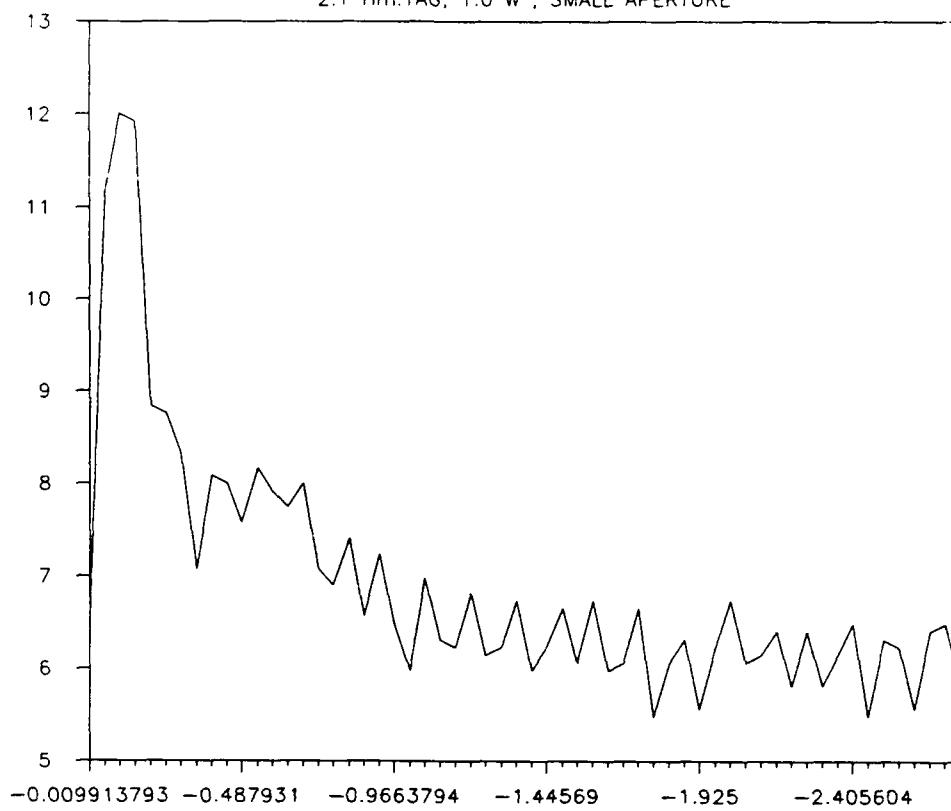
DEC17U-2X ICG CONTROLLED AT 80, 10s

2.1 Hm:YAG, 1.0 W, SMALL APERTURE



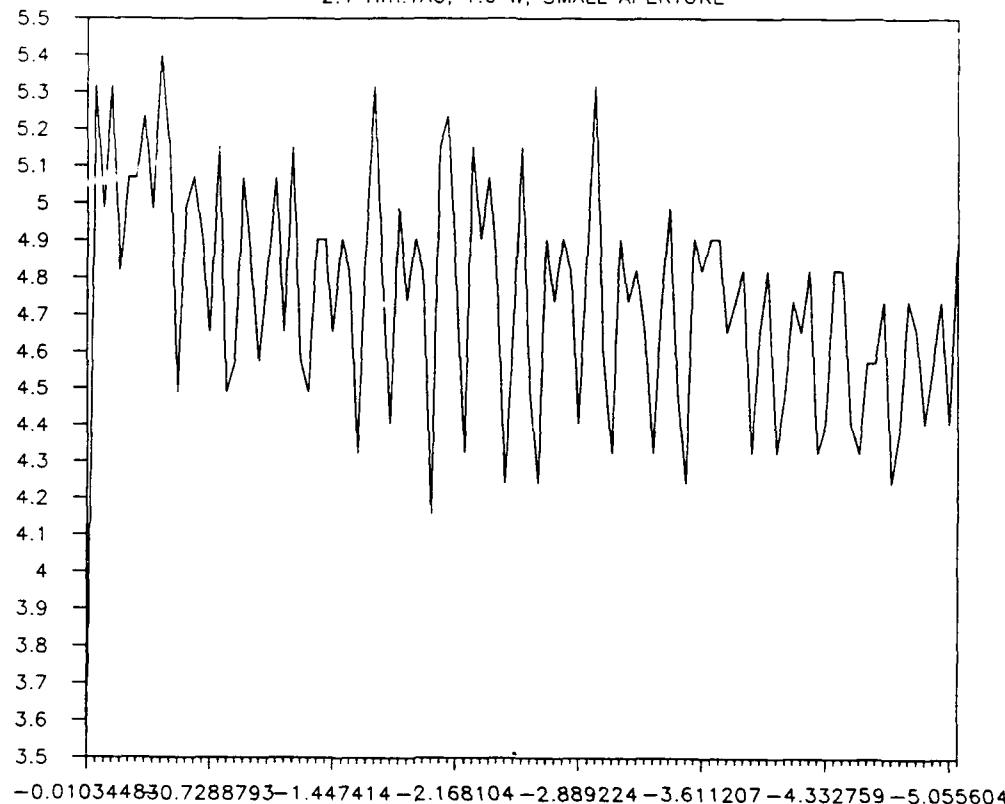
DEC17V-2X ICG SET AT 100, TC MAX=90,

2.1 Hm:YAG, 1.0 W , SMALL APERTURE



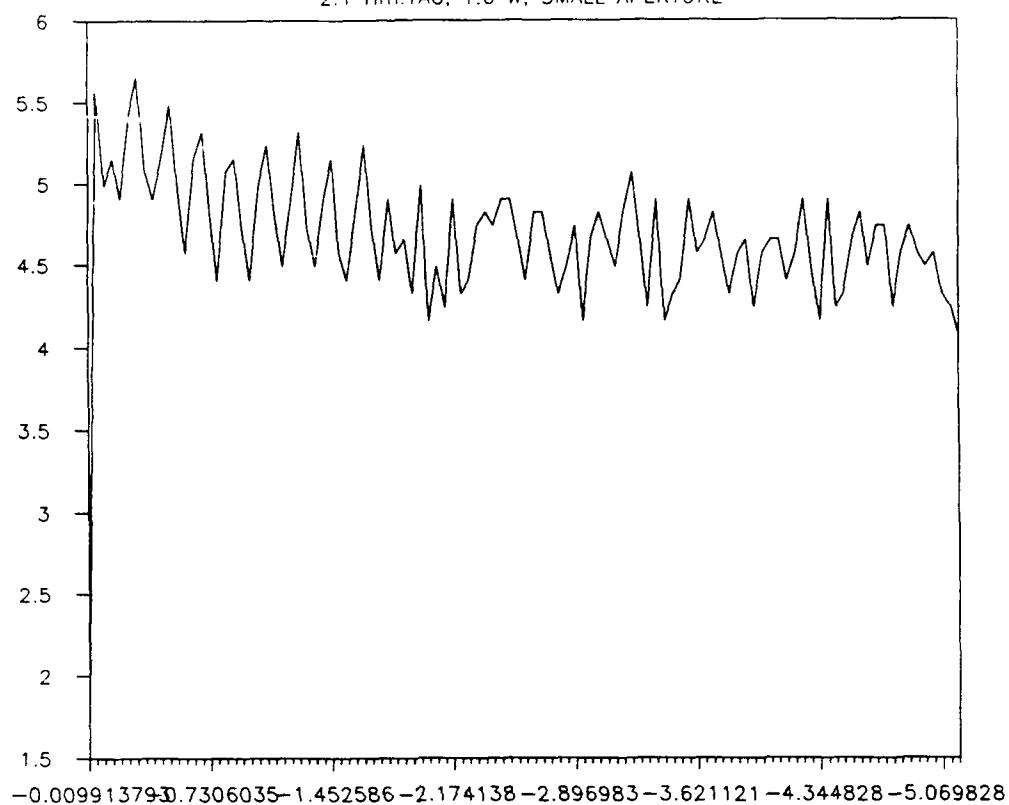
DEC17W- 2X ICG UNWELDED CONTROL

2.1 Hm:YAG, 1.0 W, SMALL APERTURE



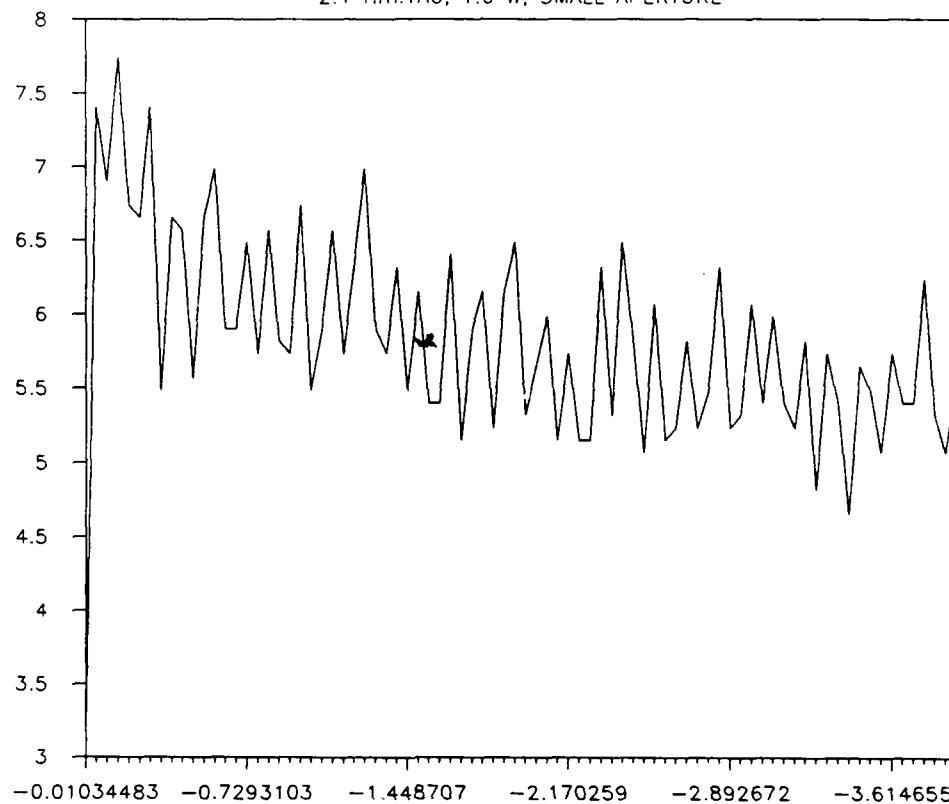
DEC17X-BLOOD UNWELDED CONTROL

2.1 Hm:YAG, 1.0 W, SMALL APERTURE



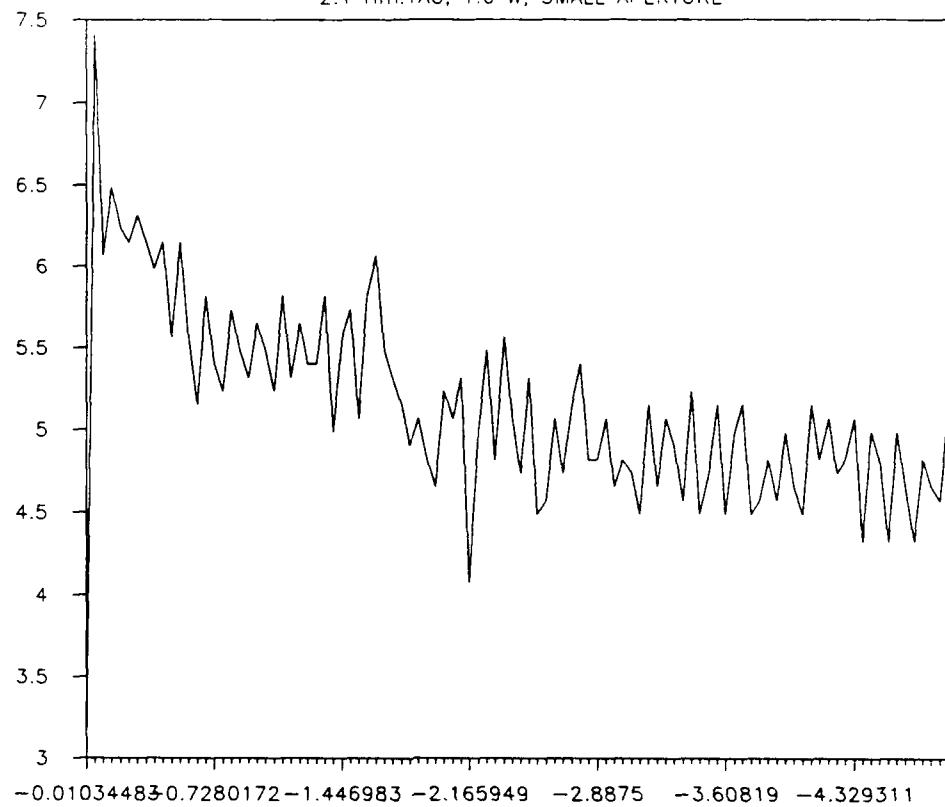
DEC17Y- BLOOD CONTROLLED AT 50, 10 s

2.1 Hm:YAG, 1.0 W, SMALL APERTURE



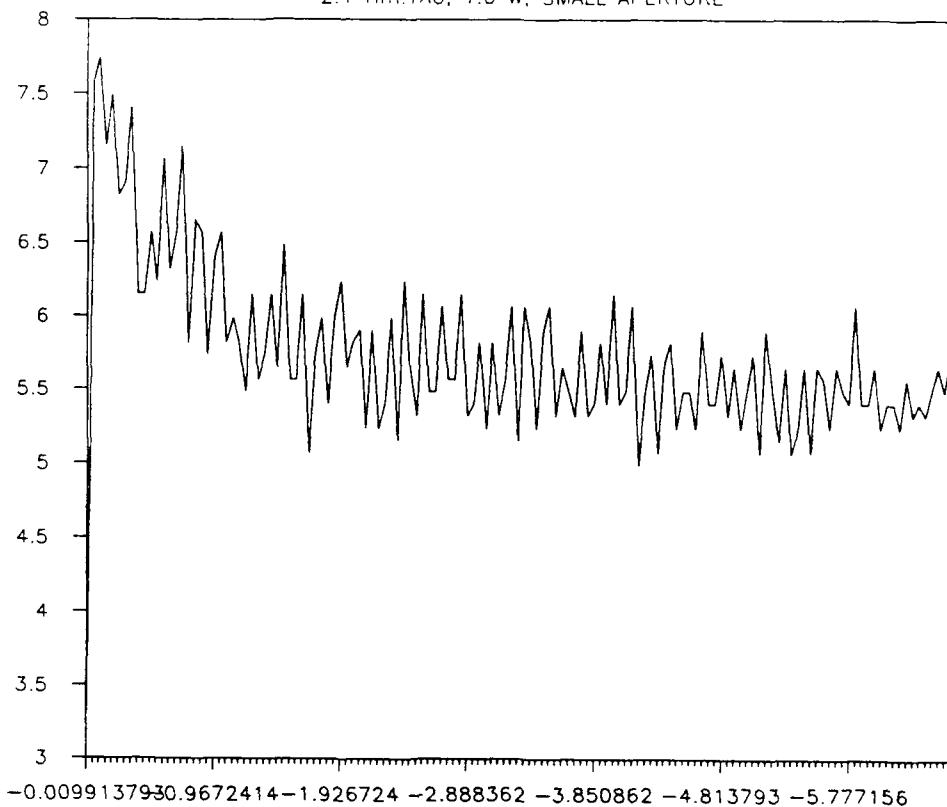
DEC17Z-BLOOD CONTROLLED AT 60, 10s

2.1 Hm:YAG, 1.0 W, SMALL APERTURE



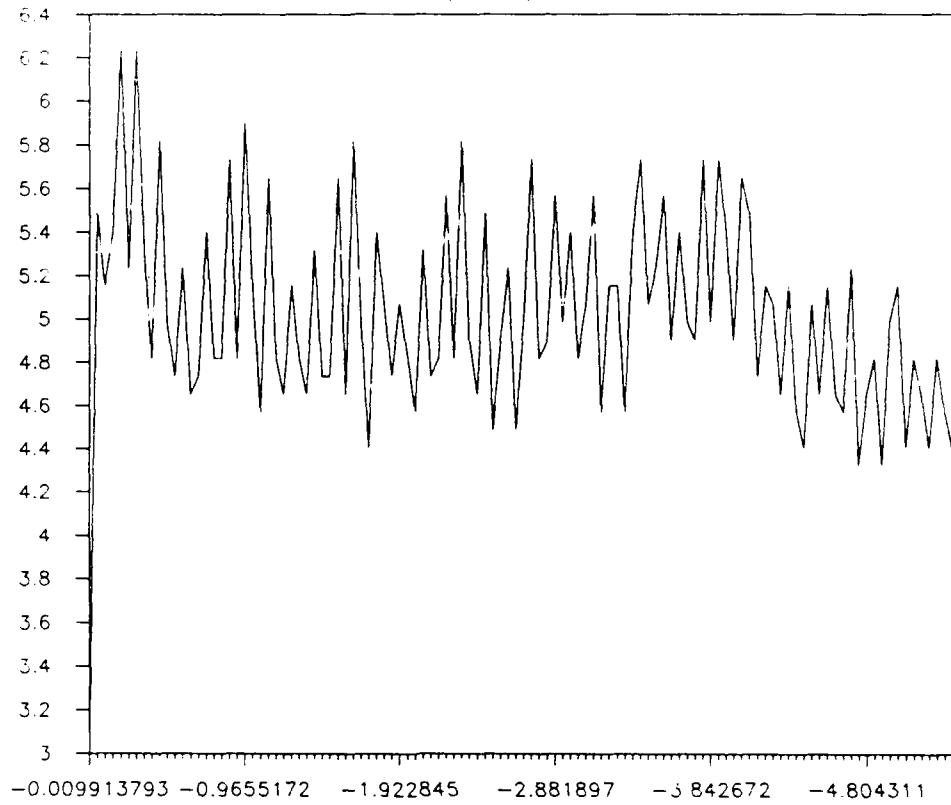
DEC17AA- BLOOD CONTROLLED AT 70, 10s

2.1 Hm:YAG, 1.0 W, SMALL APERTURE



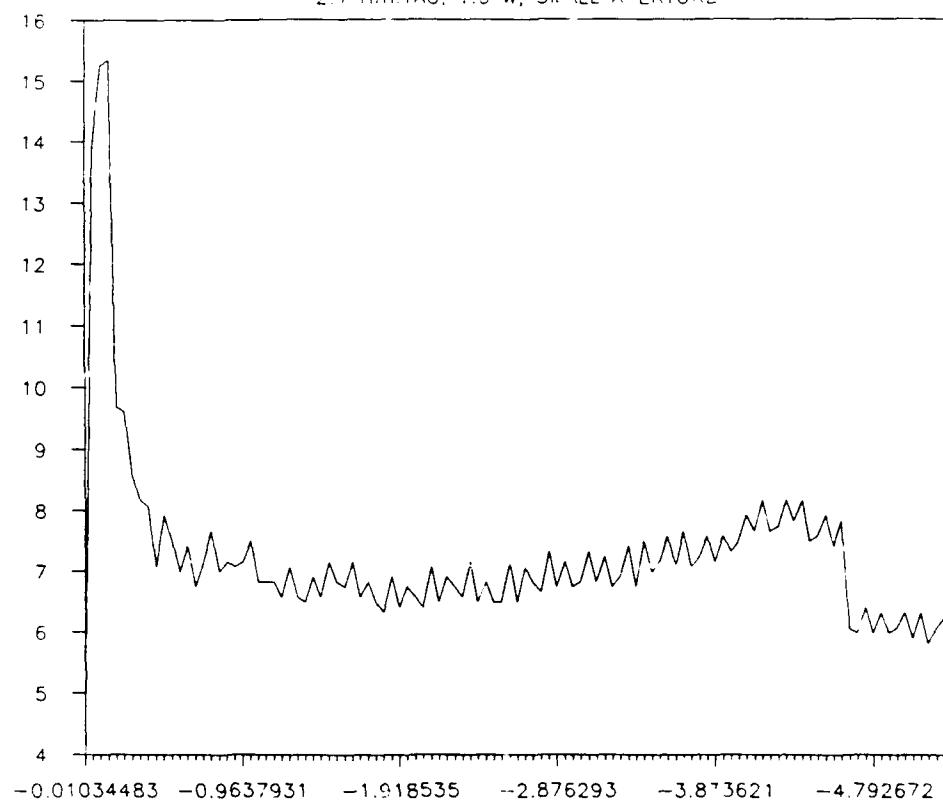
DEC17AB- BLOOD CONTROLLED AT 80, 10 s

2.1 Hm:YAG, 1.0 W, SMALL APERTURE



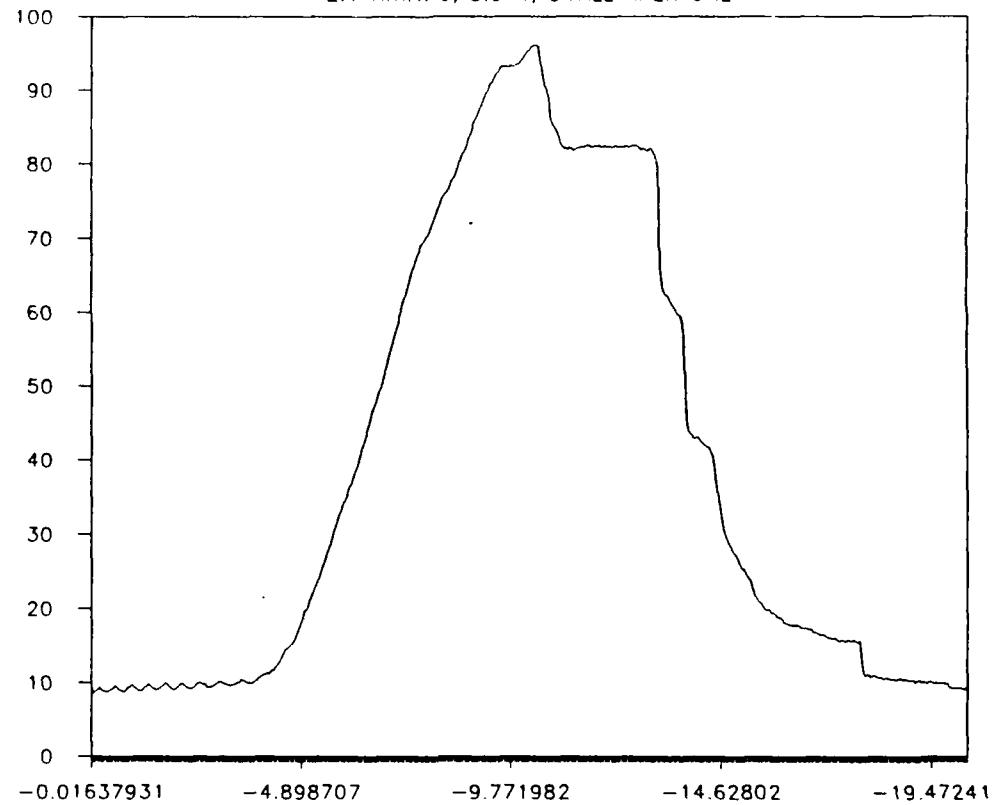
DEC17AC--BLOOD CONTROLLED AT 100, 10 s

2.1 Hz YAG, 1.0 W, SMALL APERTURE



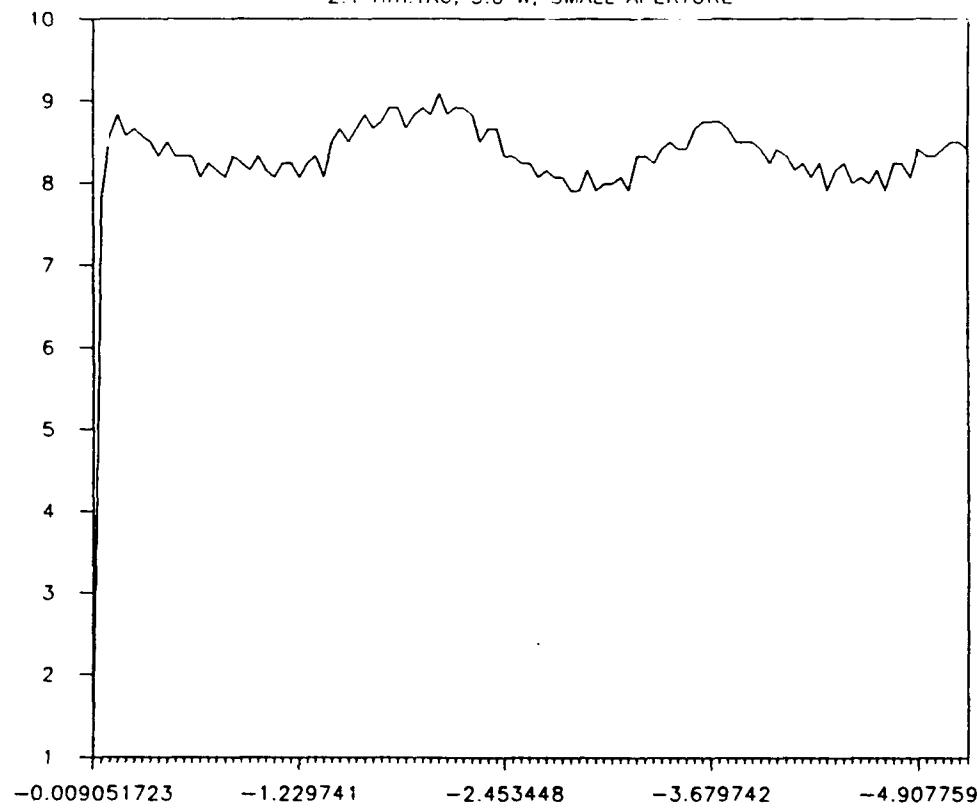
DEC18A- ONE STRIP UNWELDED CONTROL

2.1 Hm:YAG, 5.0 W, SMALL APERTURE



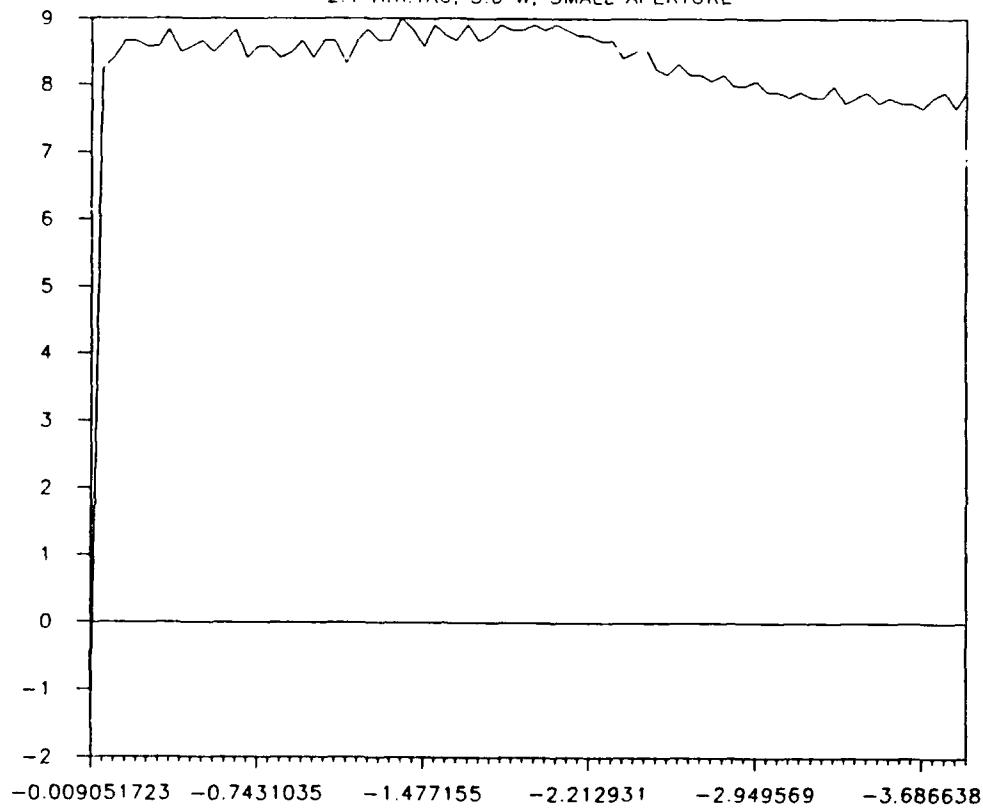
DEC18B-TWO STRIP UNWELDED CONTROL

2.1 Hm:YAG, 5.0 W, SMALL APERTURE



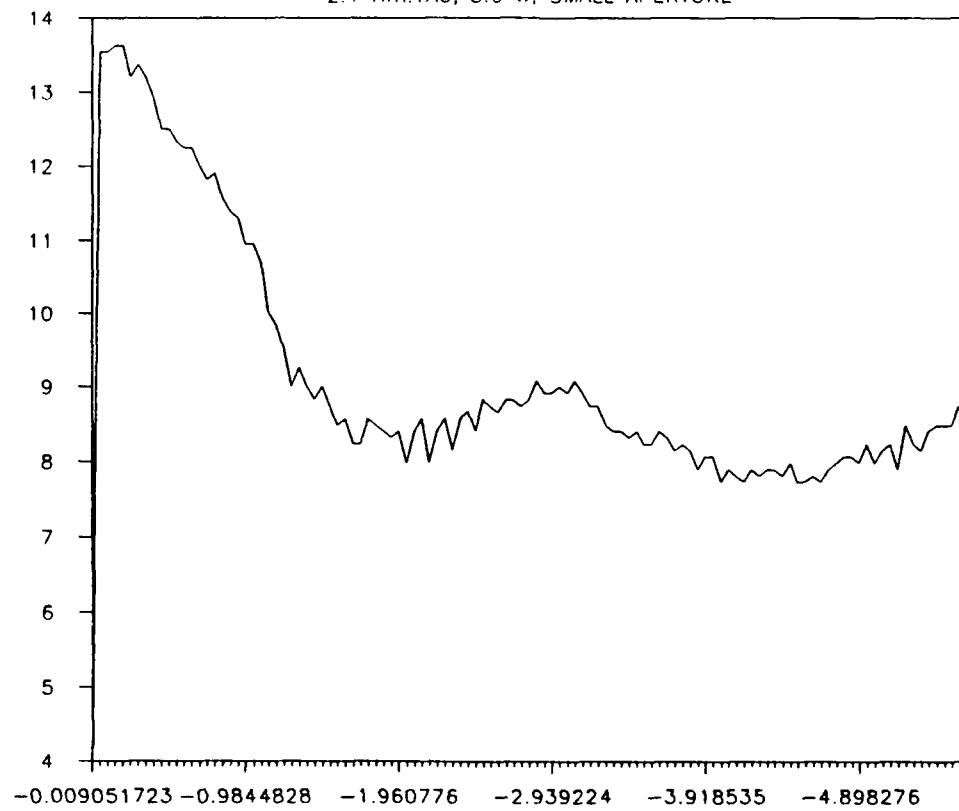
DEC18C - INDIA INK UNWELDED CONTROL

2.1 Hm:YAG, 5.0 W, SMALL APERTURE



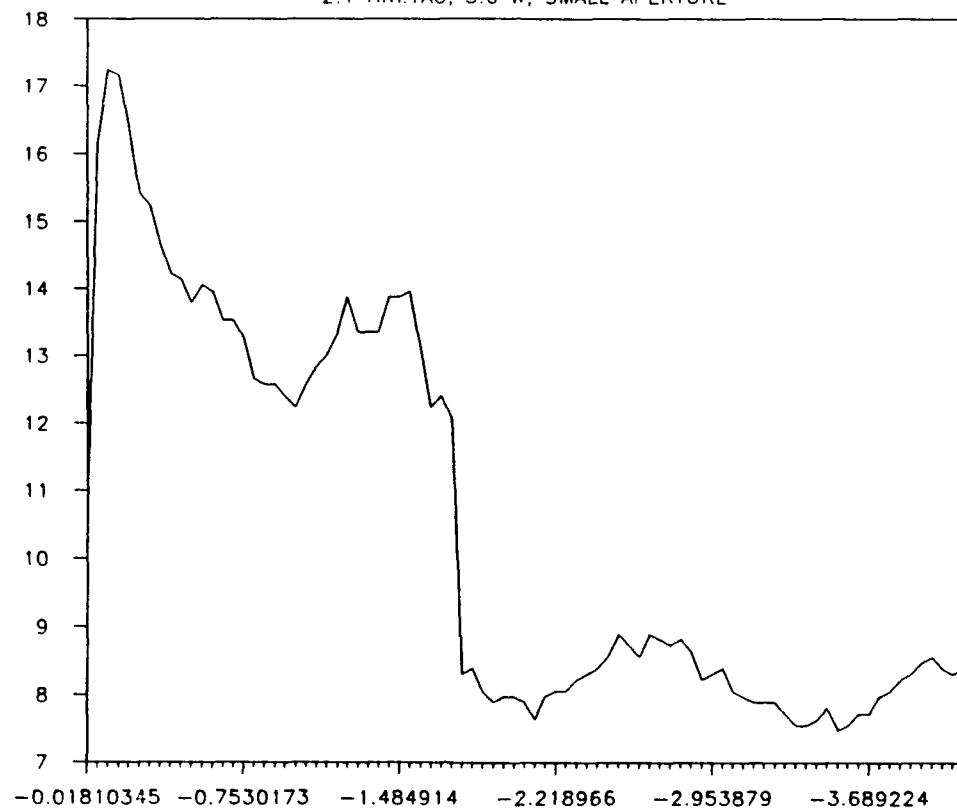
DEC18D-INDIA INK CONTROLLED AT 50, 10 s

2.1 Hm:YAG, 5.0 W, SMALL APERTURE



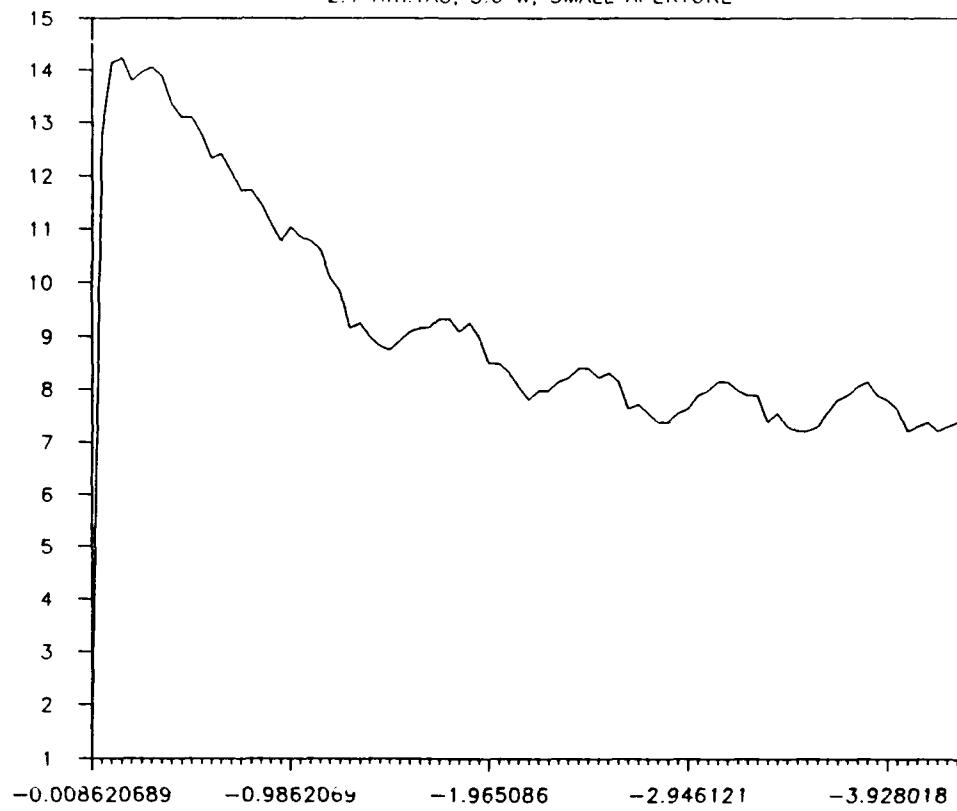
DEC18E1-INDIA INK CONTROLLED AT 60, 10s

2.1 Hm:YAG, 5.0 W, SMALL APERTURE



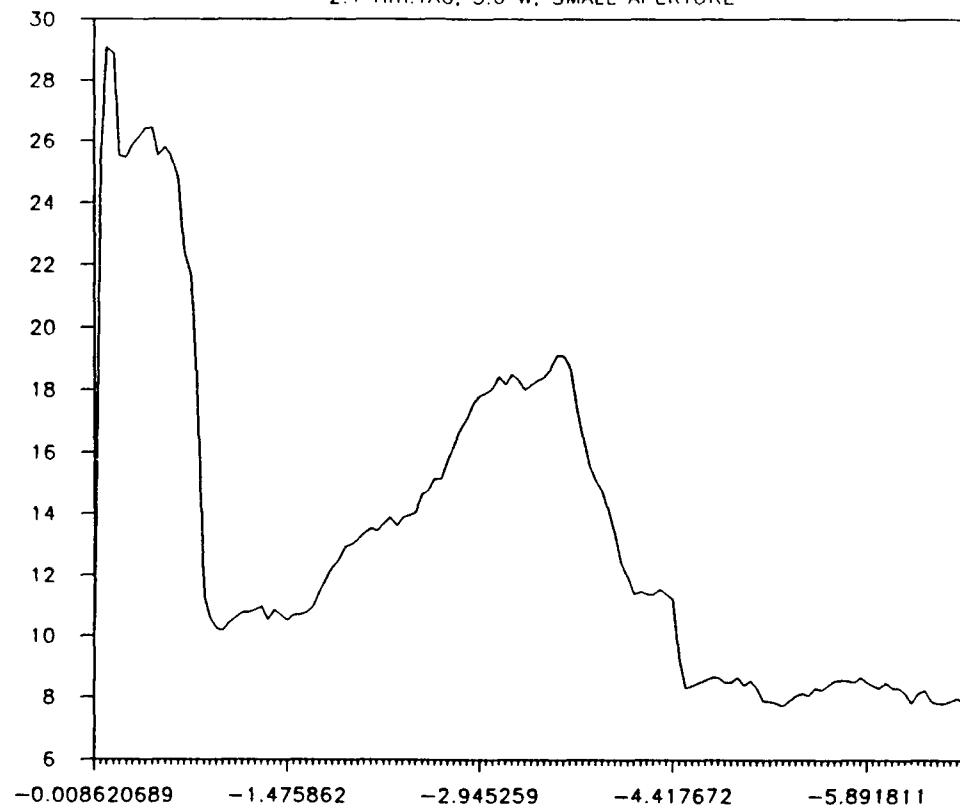
DEC18F-INDIA INK CONTROLLED AT 70, 10s

2.1 Hm:YAG, 5.0 W, SMALL APERTURE



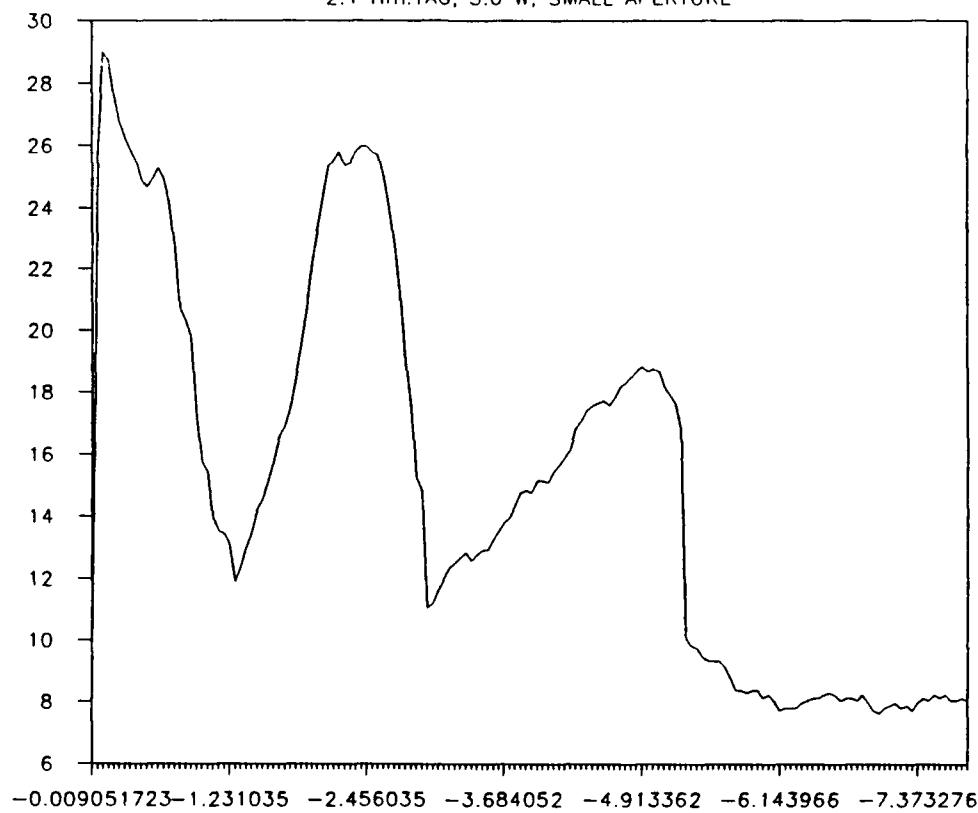
DEC18G- INDIA INK CONTROLLED AT 80, 10s

2.1 Hm:YAG, 5.0 W, SMALL APERTURE



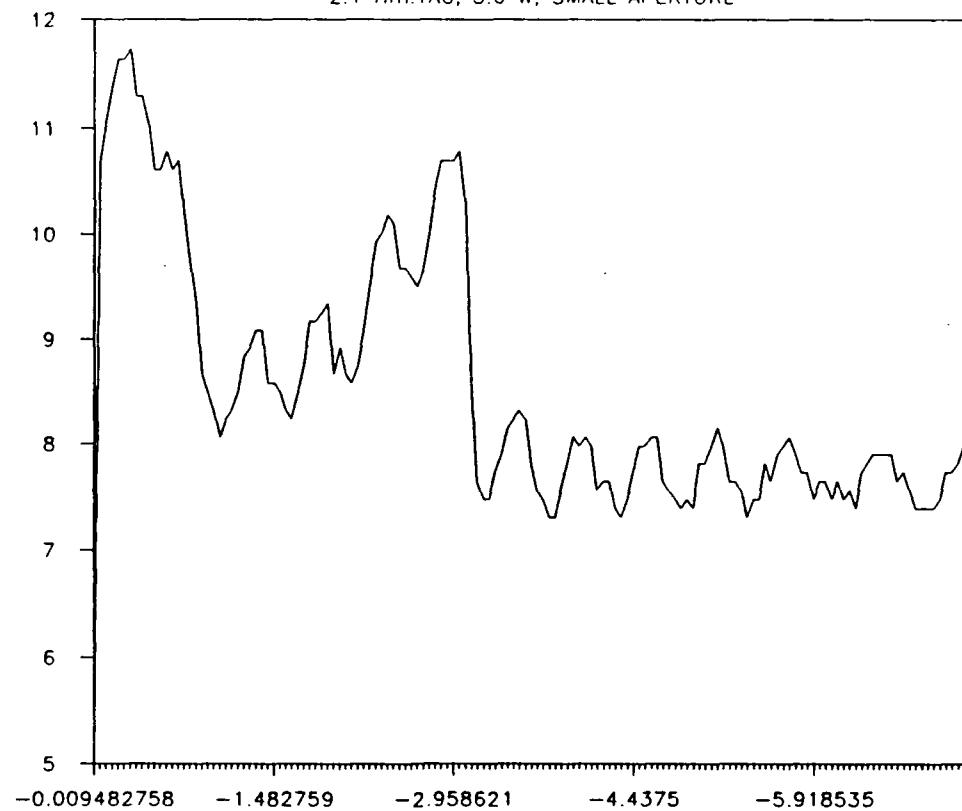
DEC18H-INDIA INK CONTROLLED AT 100, 10s

2.1 Hm:YAG, 5.0 W, SMALL APERTURE



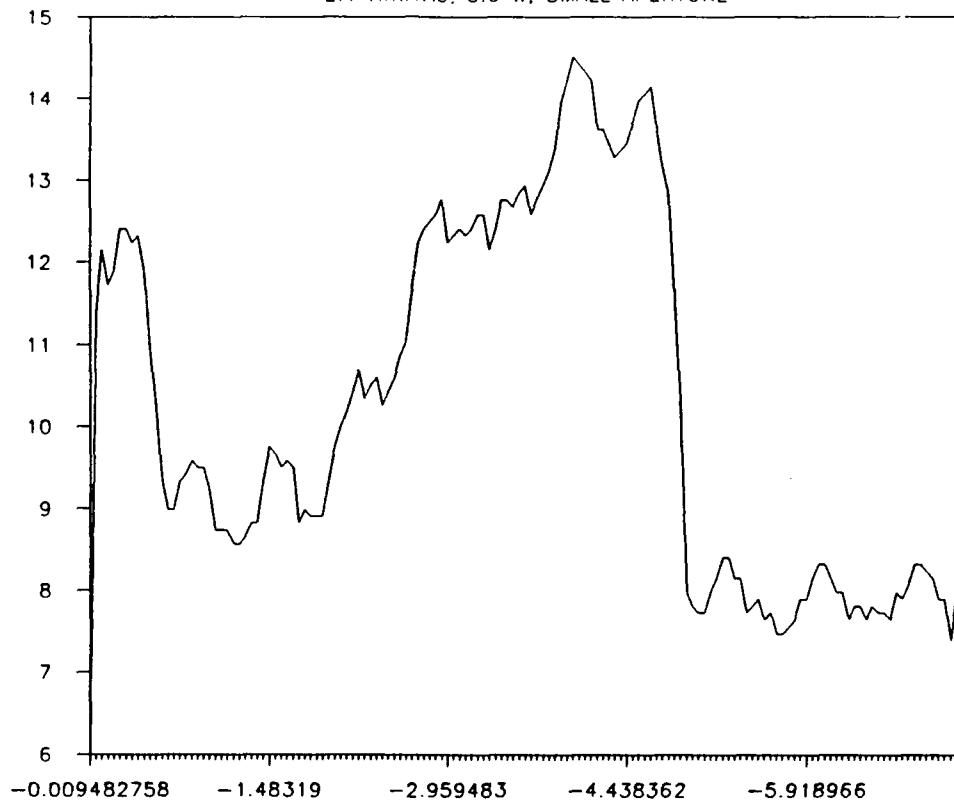
DEC18I- NO CHROM CONTROLLED AT 50, 10s

2.1 Hm:YAG, 5.0 W, SMALL APERTURE



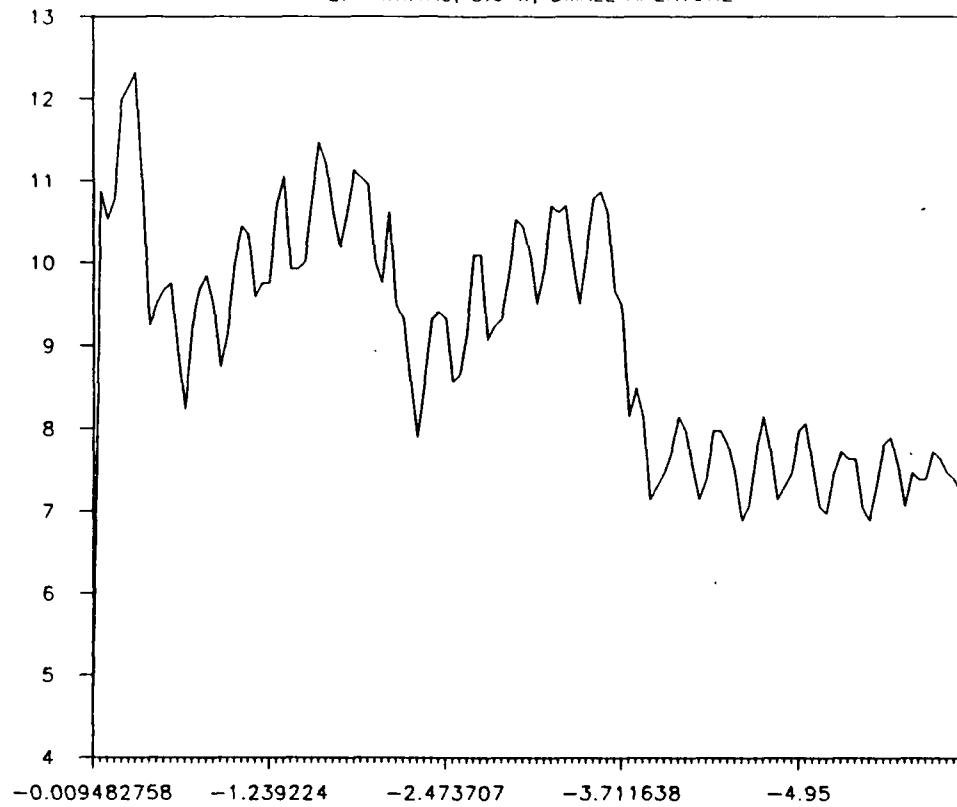
DEC18J-NO CHROMO CONTROLLED AT 60, 10s

2.1 Hm:YAG, 5.0 W, SMALL APERTURE



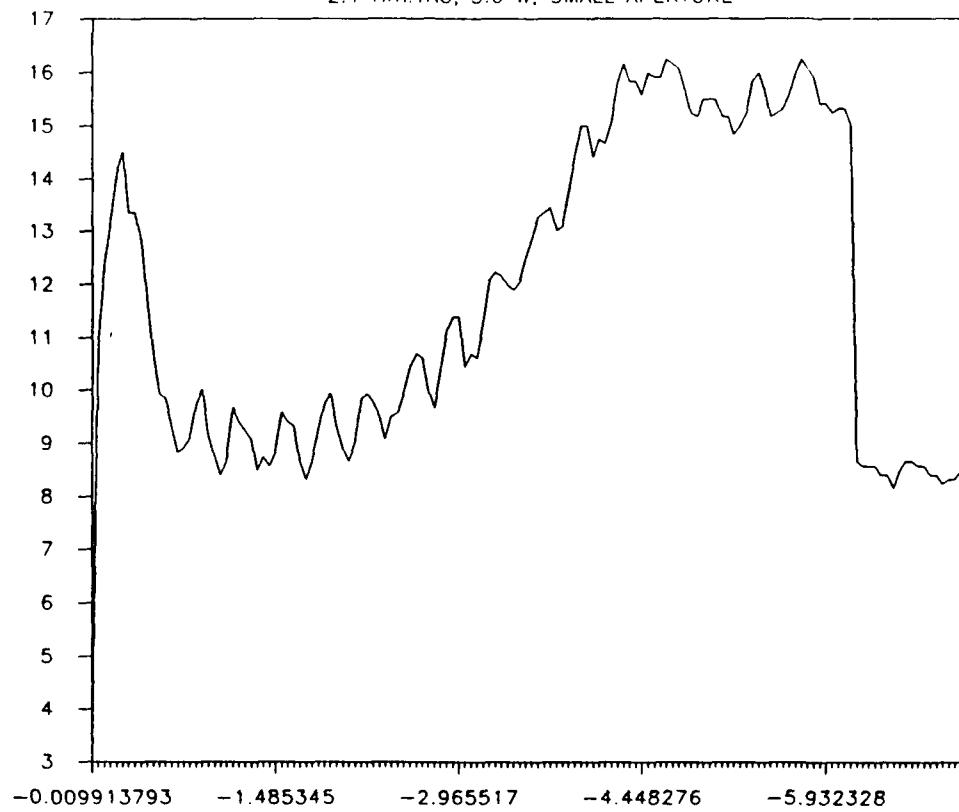
DEC18K- NO CHROMO CONTROLLED AT 60, 10s

2.1 Hm:YAG, 5.0 W, SMALL APERTURE



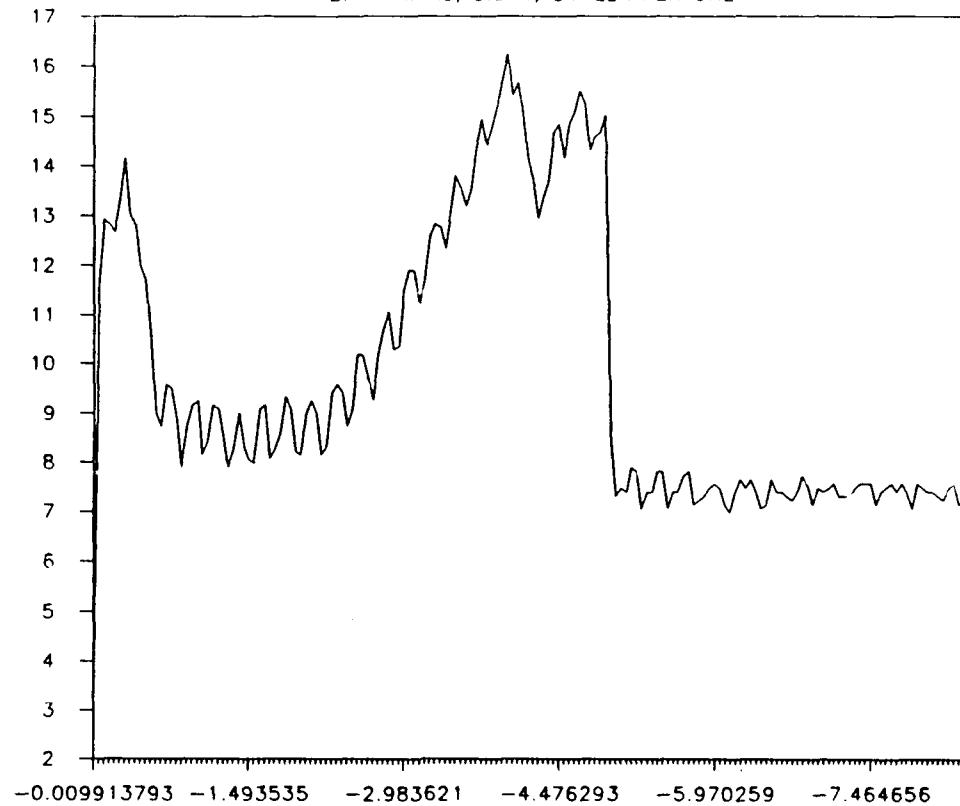
DEC18L-NO CHROMO CONTROLLED AT 70, 10 s

2.1 Hm:YAG, 5.0 W. SMALL APERTURE



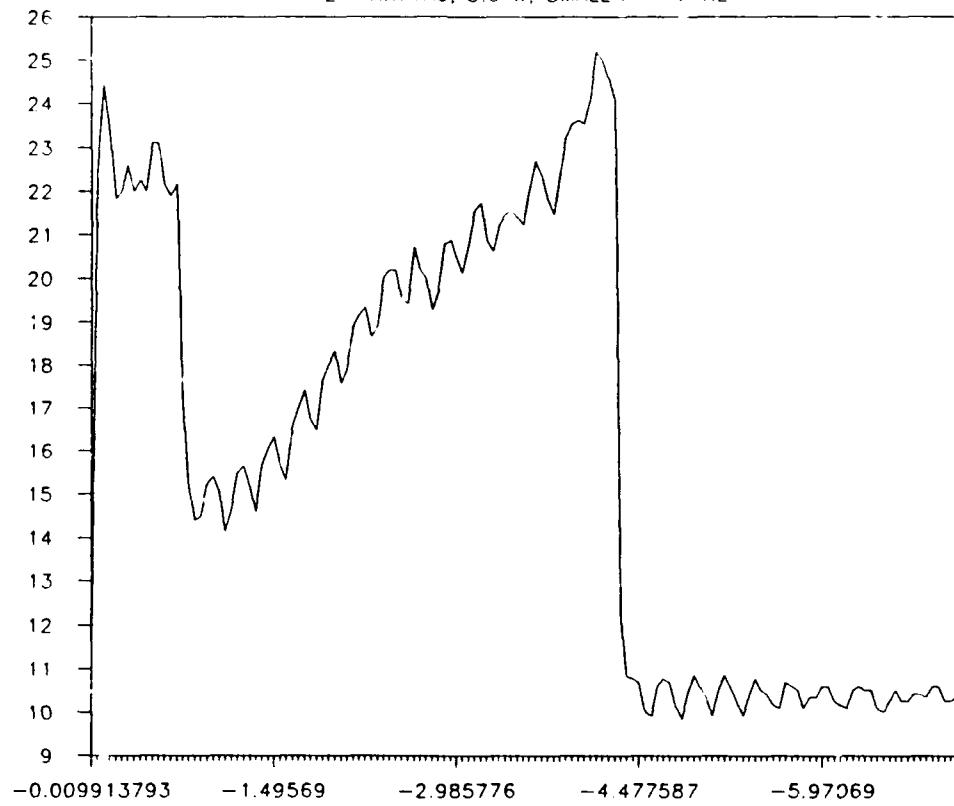
DEC18N-NO CHROMO-CONTROLLED AT 80, 10 s

2.1 Hm:YAG, 5.0 W, SMALL APERTURE



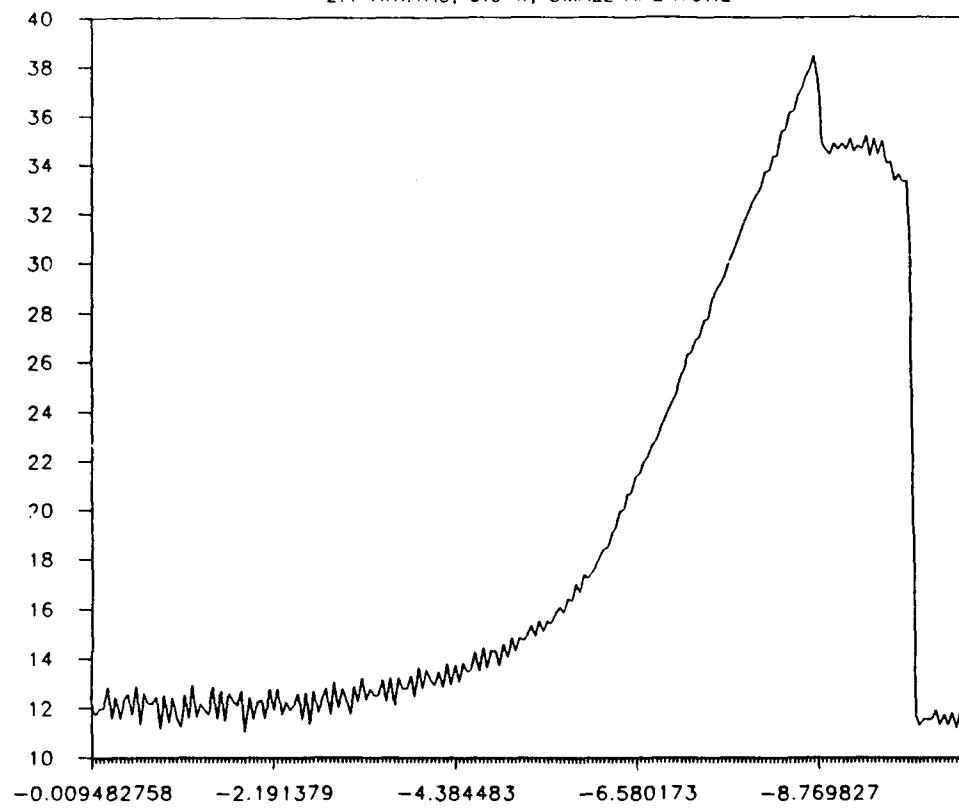
DEC'80 - NO CHROMO CONTROLLED AT 100,10s

2.1 HM:YAG, 5.0 W, SMALL APERTURE



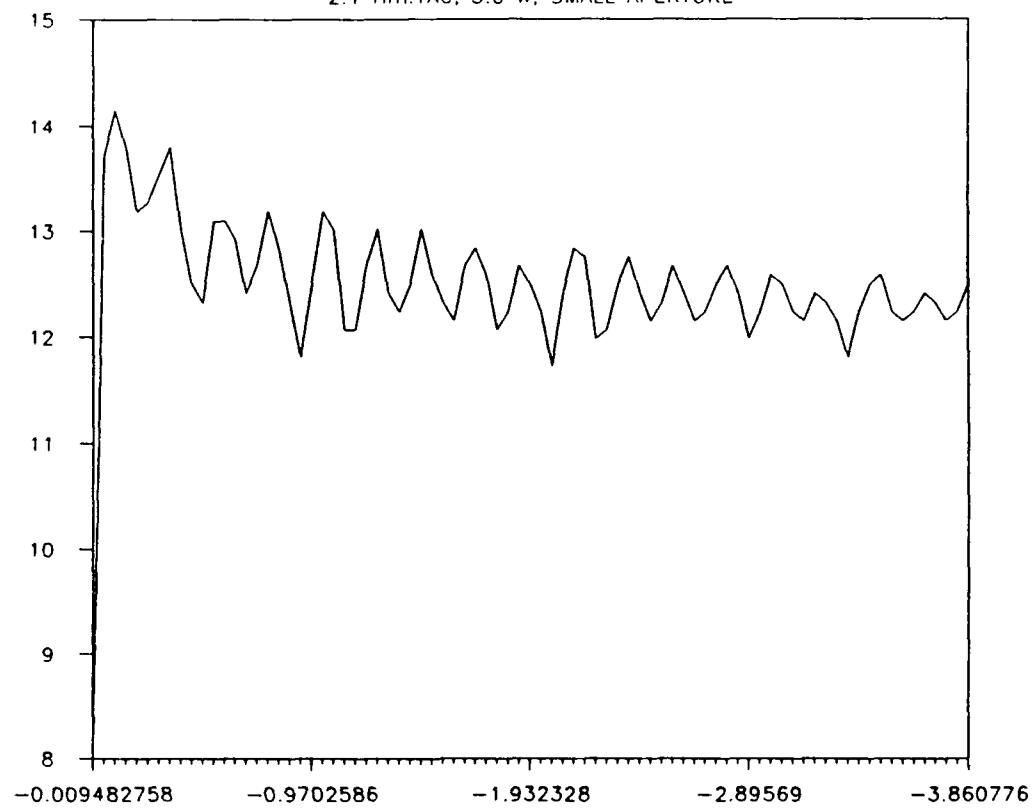
DEC18P-NO CHROMO CONTROLLED AT 100, 10s

2.1 Hm:YAG, 5.0 W, SMALL APERTURE



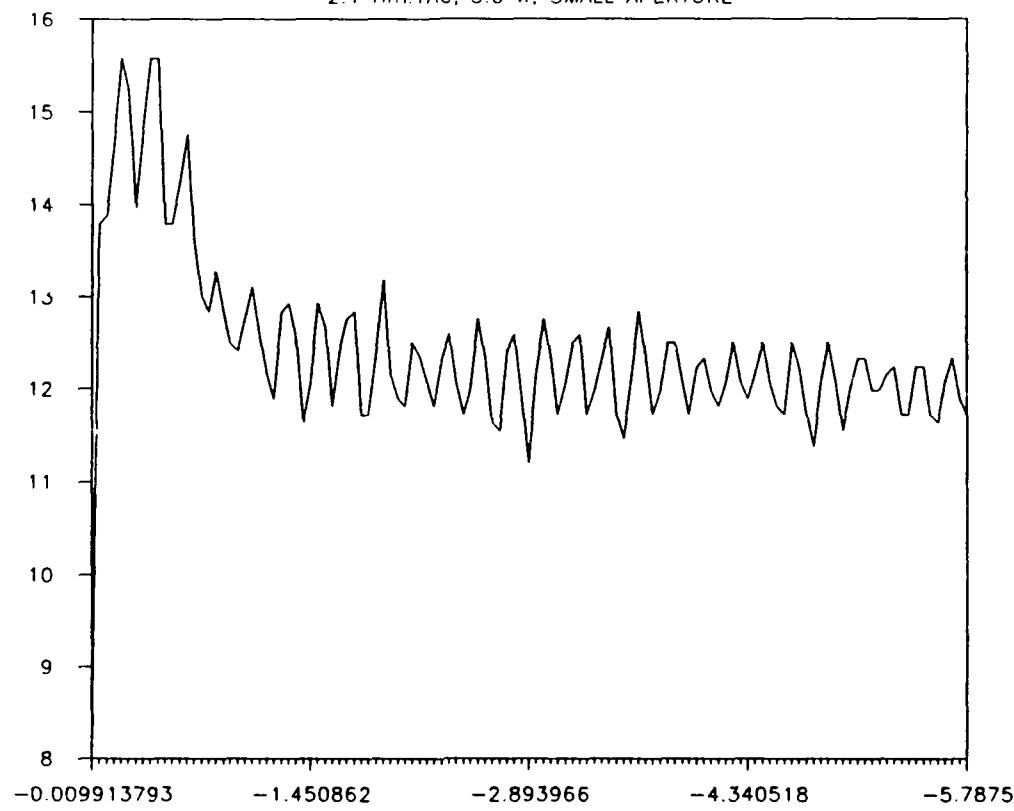
DEC18Q- 2X ICG UNWELDED CONTROL

2.1 Hm:YAG, 5.0 W, SMALL APERTURE



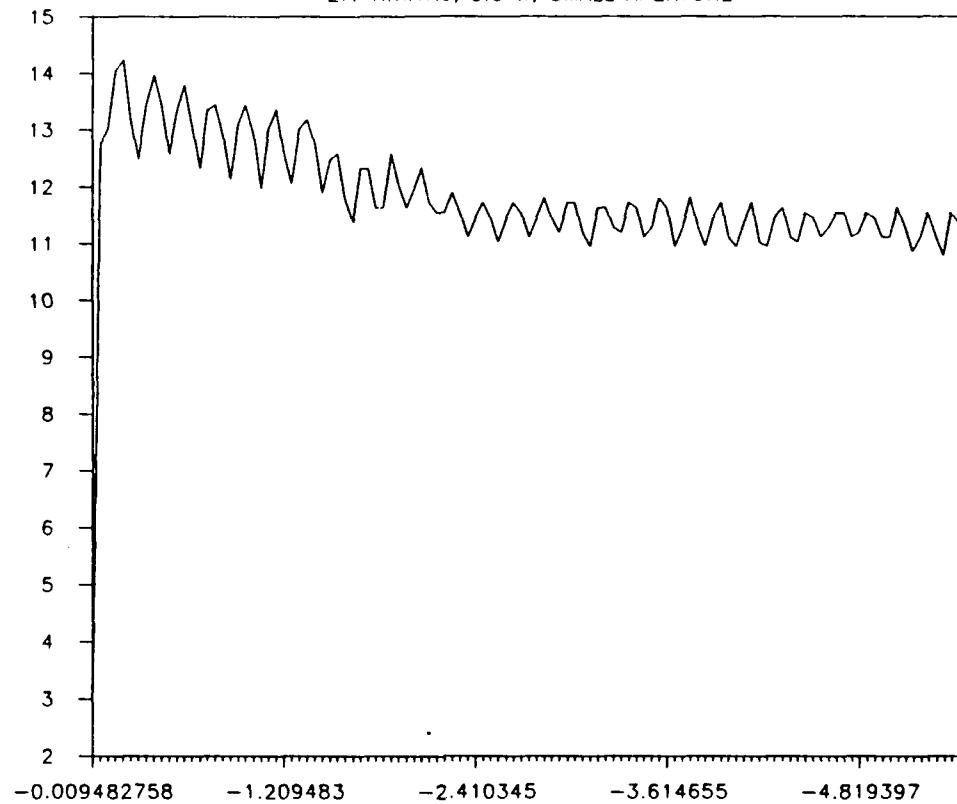
DEC18R- 2X ICG CONTROLLED AT 50,10 s

2.1 Hm:YAG, 5.0 W, SMALL APERTURE



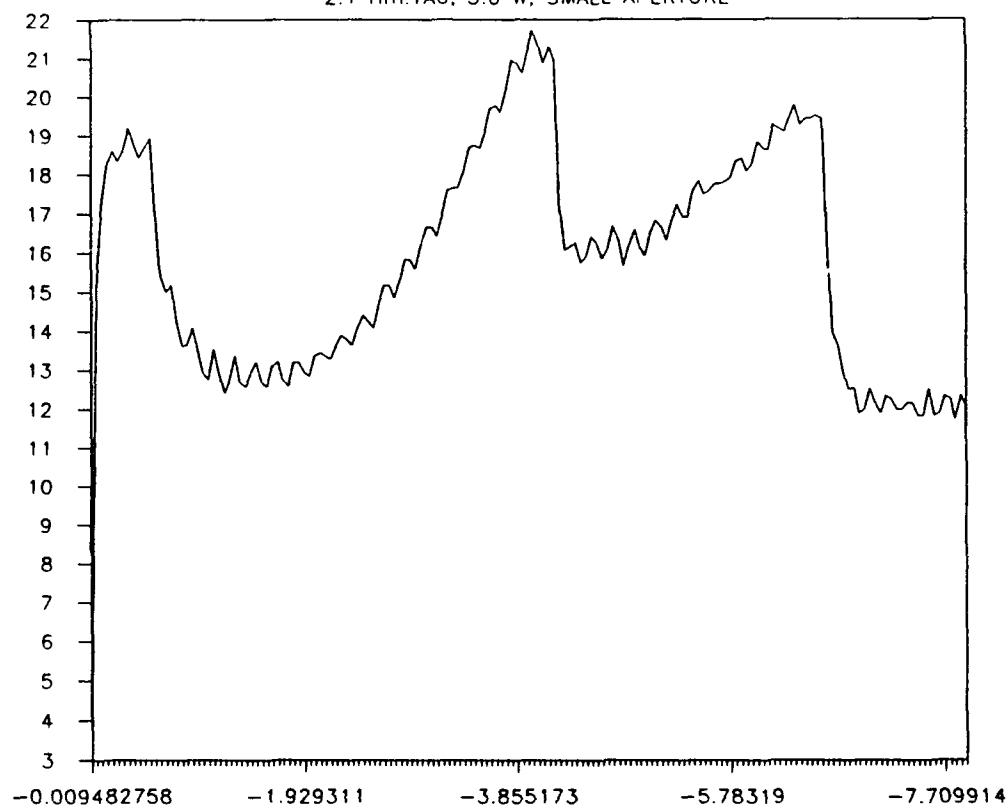
DEC18S-- 2X ICG CONTROLLED AT 60, 10 s

2.1 Hm:YAG, 5.0 W, SMALL APERTURE



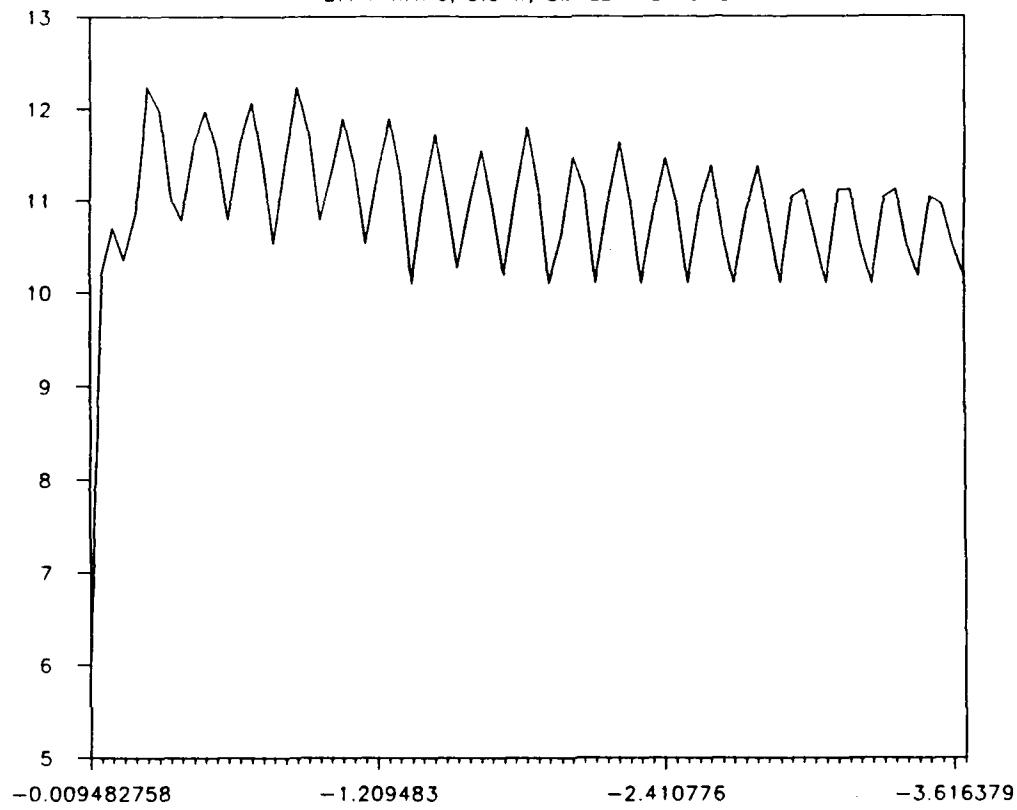
DEC18T- 2X ICG CONTROLLED AT 70, 10 s

2.1 Hm:YAG, 5.0 W, SMALL APERTURE



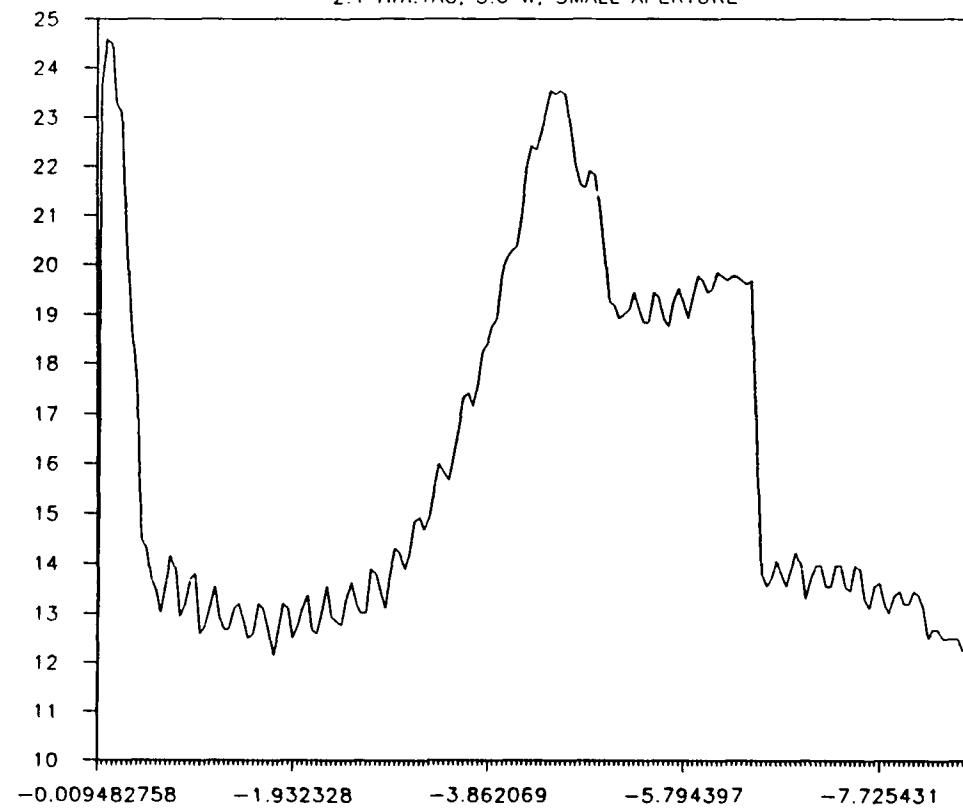
DEC18U- 2X ICG CONTROLLED AT 80, 10s

2.1 Hm:YAG, 5.0 W, SMALL APERTURE



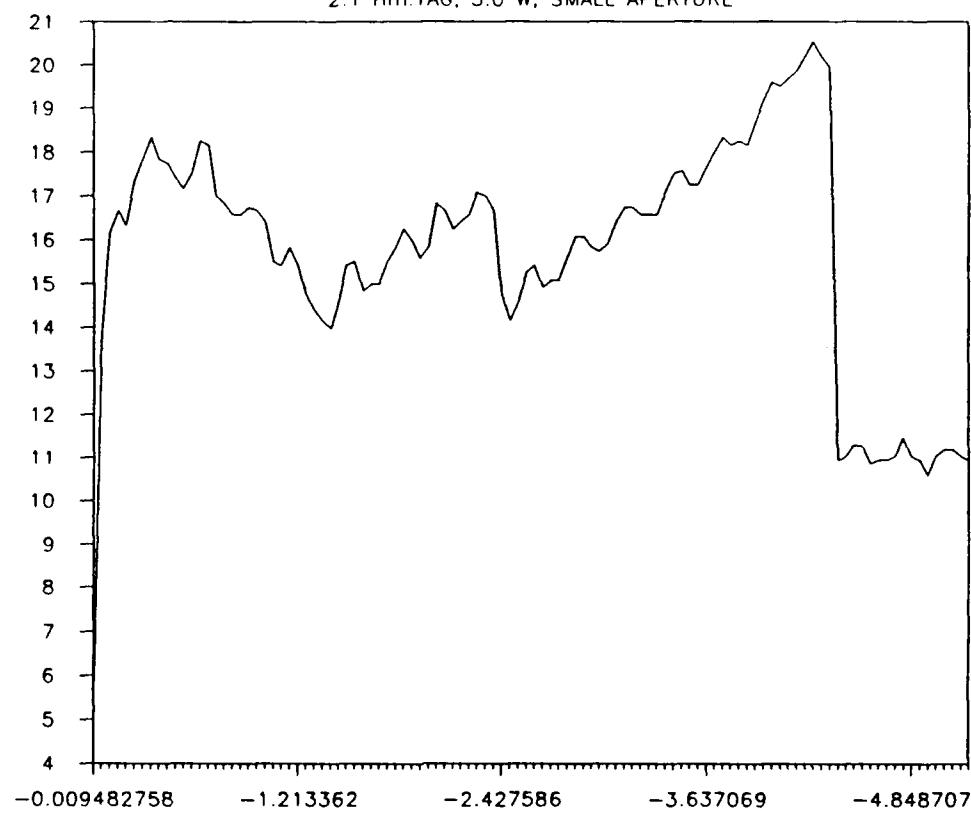
DEC18V- 2X CIG CONTROLLED AT 100, 10s

2.1 Hz:YAG, 5.0 W, SMALL APERTURE



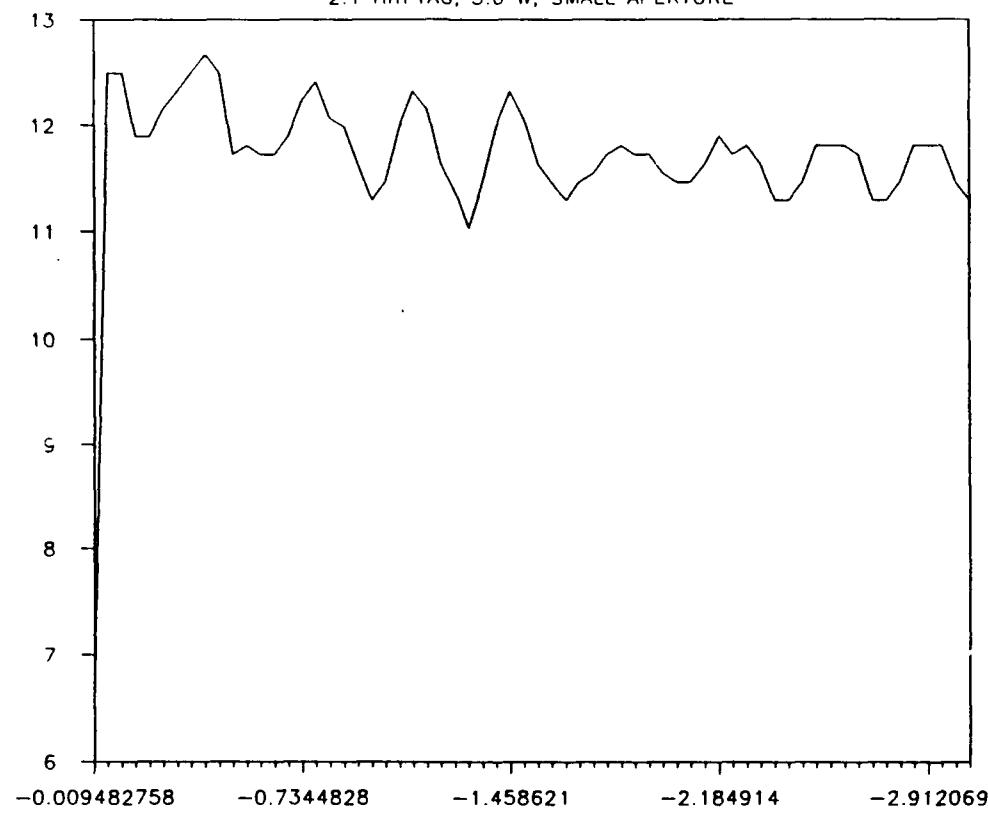
DEC18W1 - 2X ICG CONTROLLED AT 80, 10s

2.1 Hm:YAG, 5.0 W, SMALL APERTURE



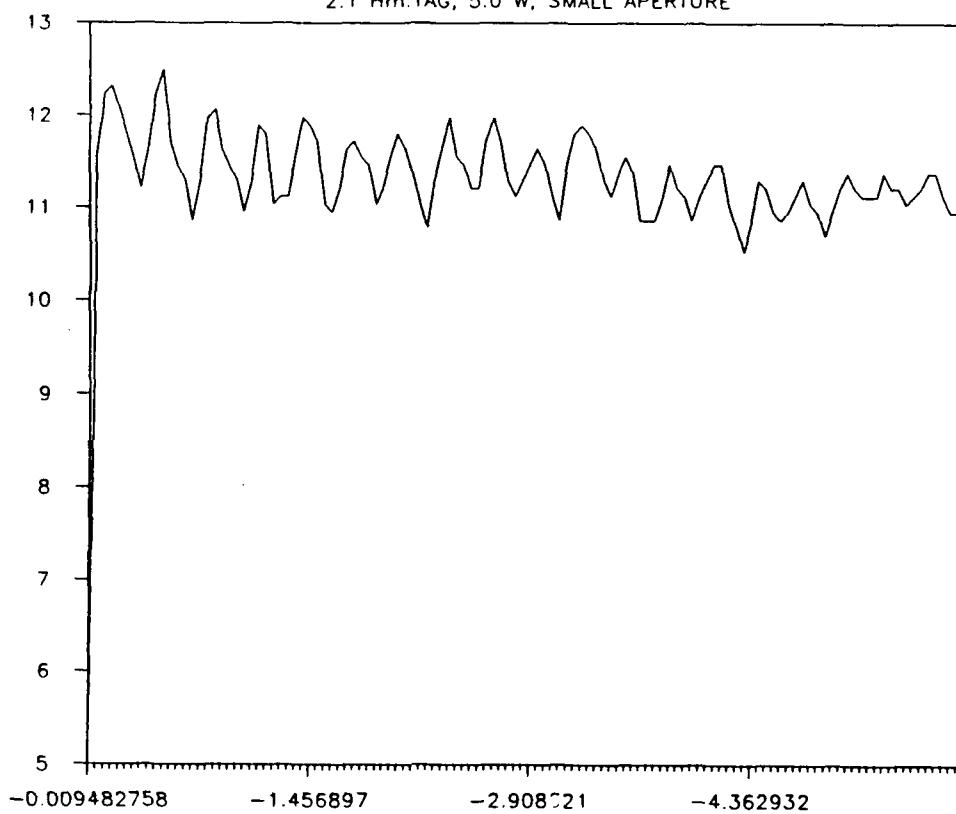
DEC18X- BLOOD UNWELDED CONTROL

2.1 Hm·YAG, 5.0 W, SMALL APERTURE



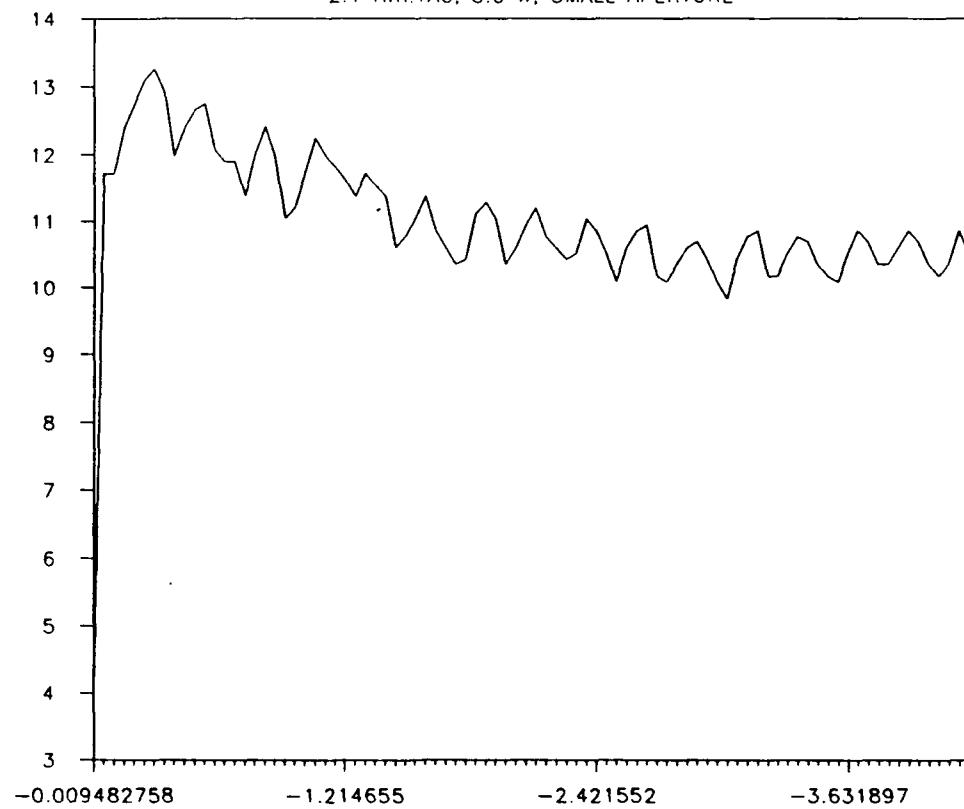
DEC18Y- BLOOD CONTROLLED AT 50, 10 s

2.1 Hm:YAG, 5.0 W, SMALL APERTURE



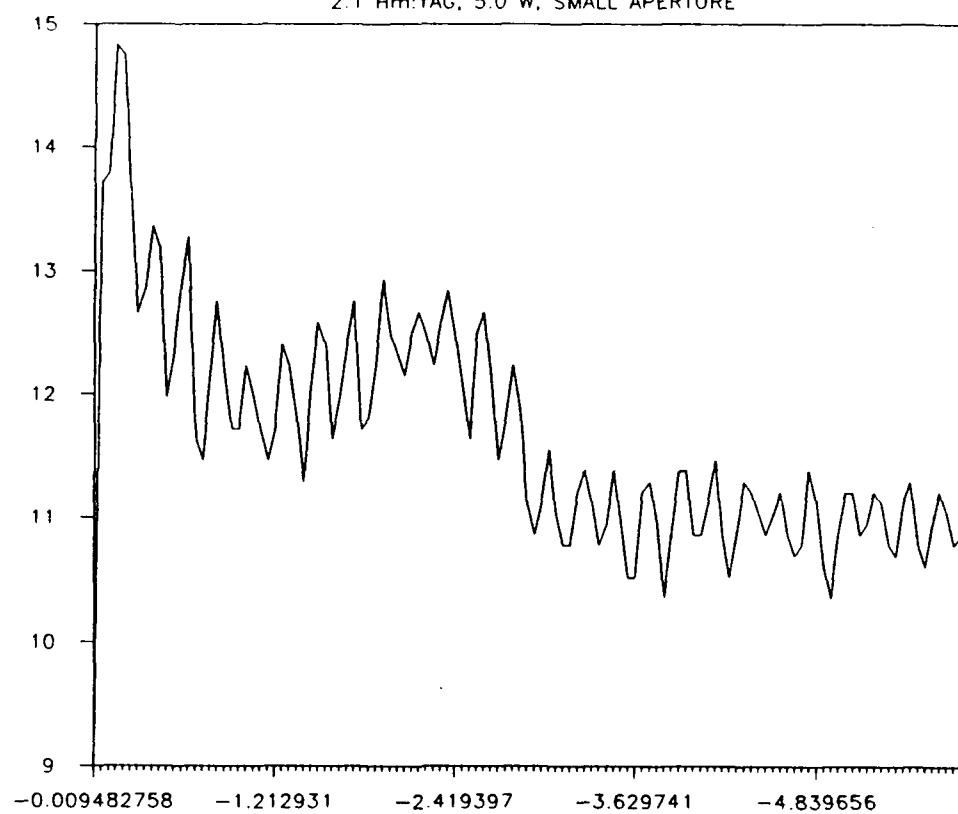
DEC18Z- BLOOD CONTROLLED AT 50, 10 s

2.1 Hm:YAG, 5.0 W, SMALL APERTURE



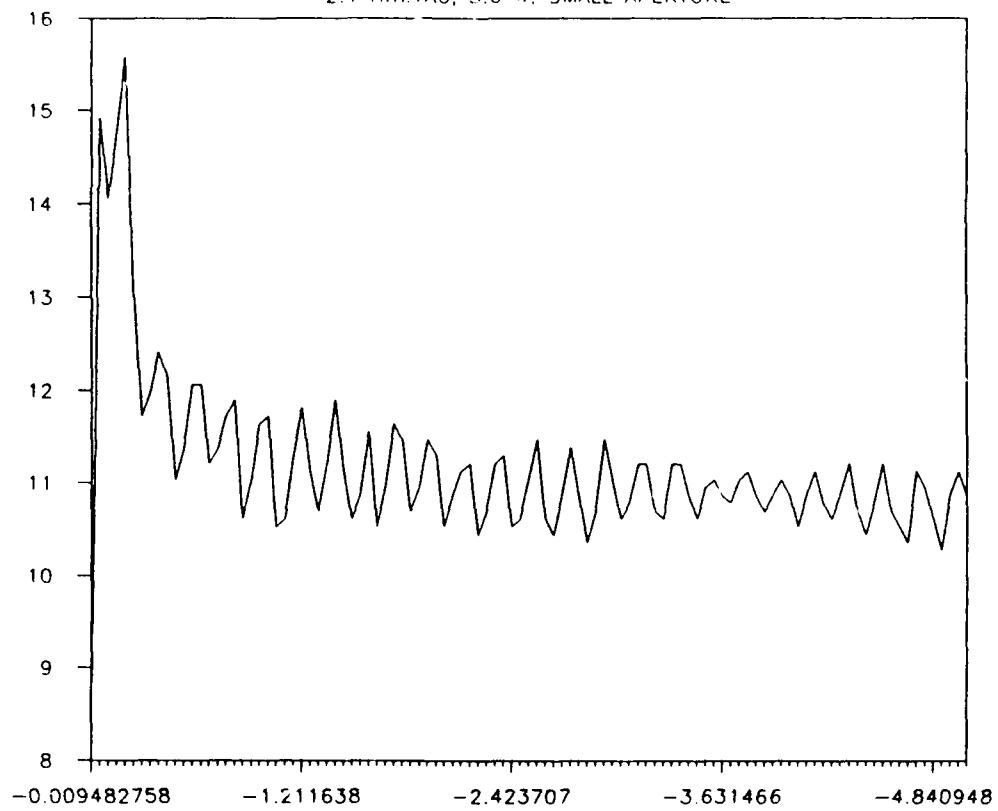
DEC18AA- BLOOD CONTROLLED AT 60, 10s

2.1 Hm:YAG, 5.0 W, SMALL APERTURE



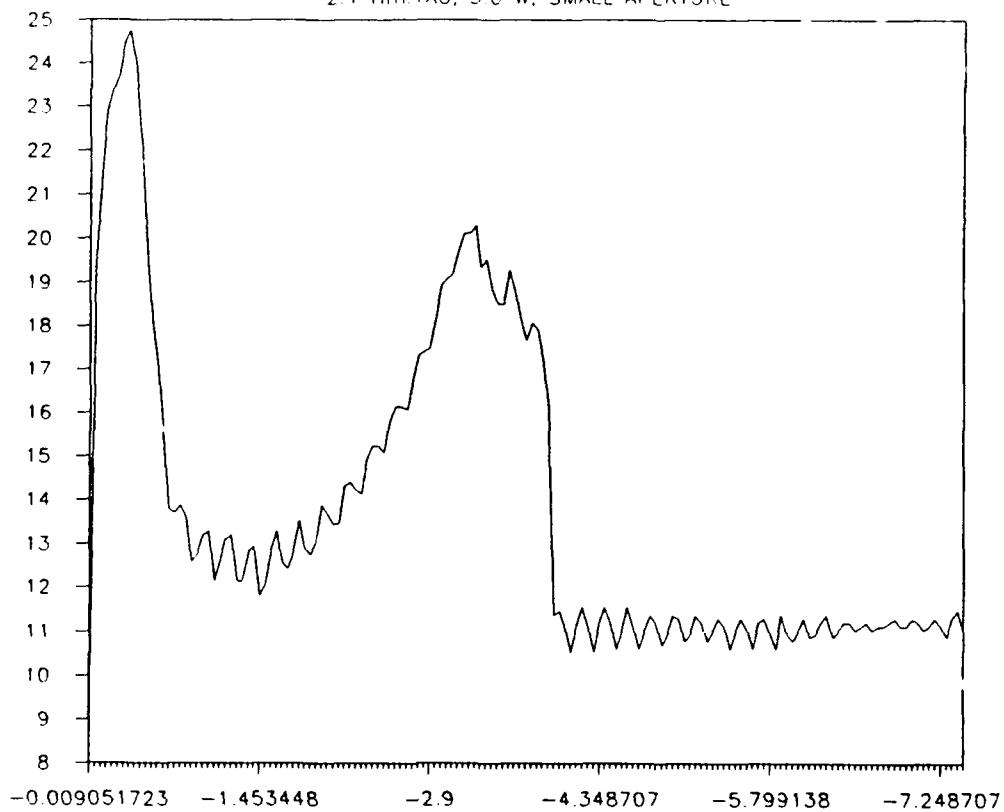
DEC18AB - BLOOD CONTROLLED AT 70, 10 s

2.1 Hm:YAG, 5.0 W. SMALL APERTURE



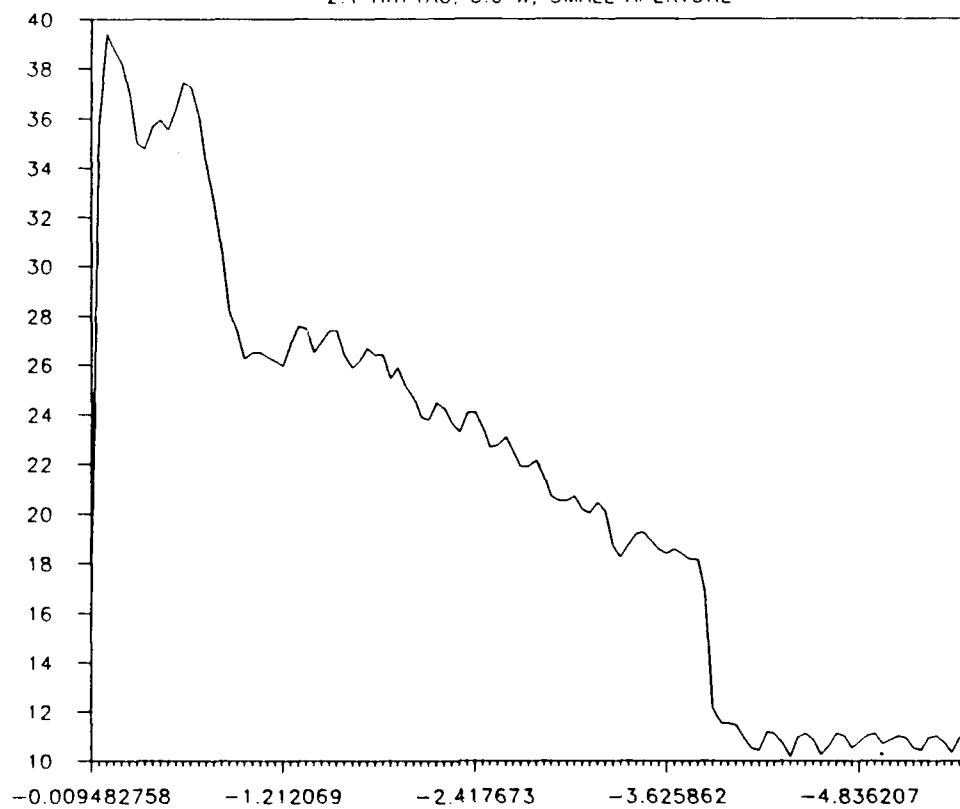
DEC18AC - BLOOD CONTROLLED AT 80, 10 s

2.1 Hm:YAG, 5.0 W, SMALL APERTURE



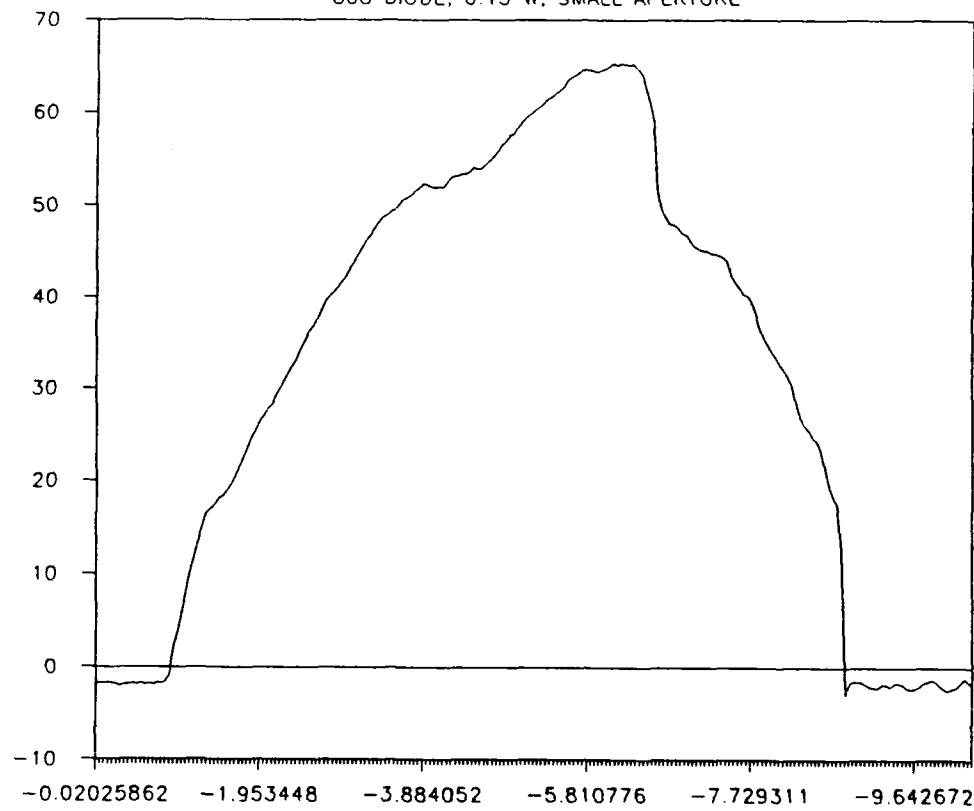
DEC18AD- BLOOD CONTROLLED AT 100, 10 s

2.1 Hm-YAG, 5.0 W, SMALL APERTURE



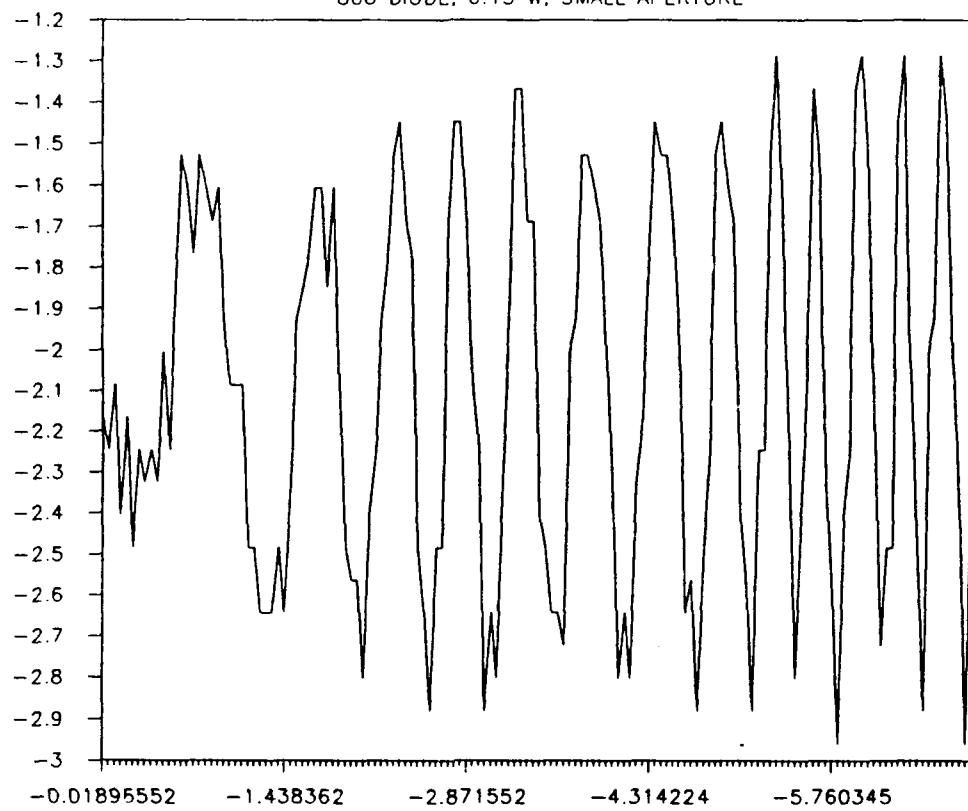
DEC19A- ONE STRIP UNWELDED CONTROL

808 DIODE, 0.15 W, SMALL APERTURE



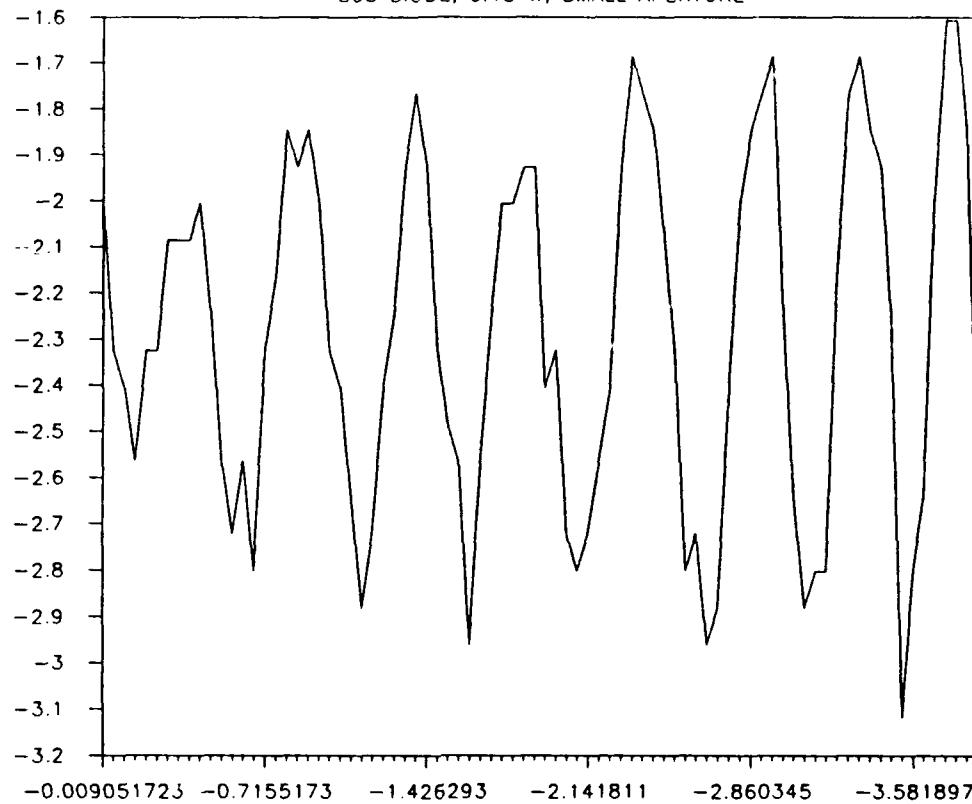
DEC1981- TWO STRIP UNWELDED CONTROL

808 DIODE, 0.15 W, SMALL APERTURE



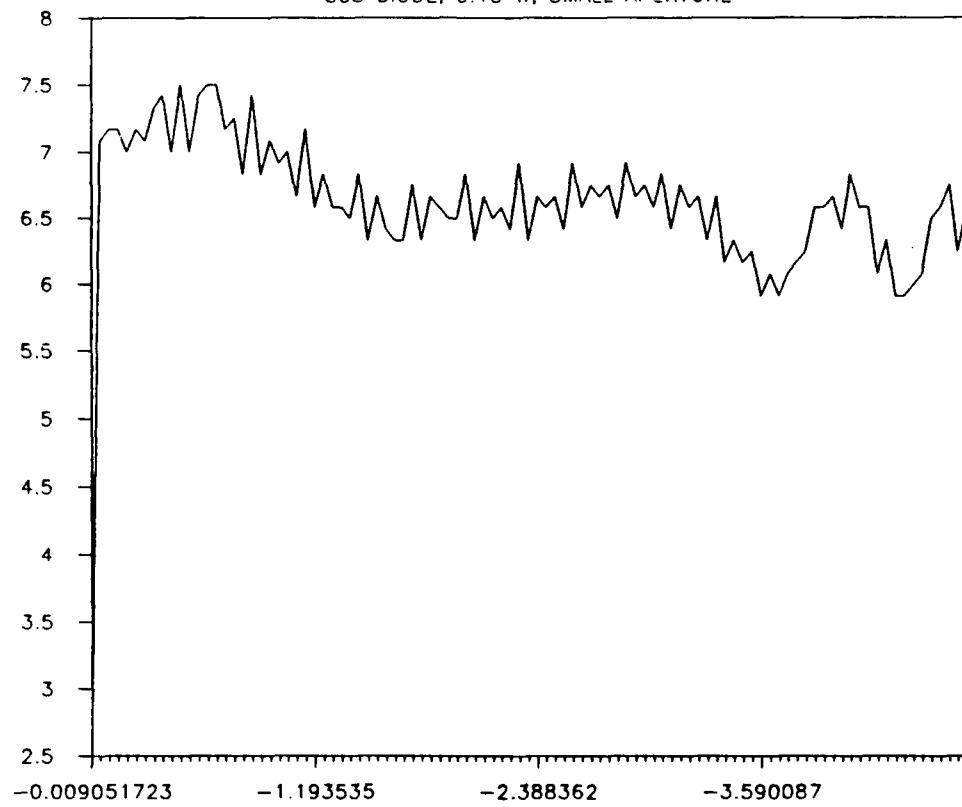
DEC19C-TWO STRIP 2X ICG UNWELDED CONTROL

808 DIODE, 0.15 W, SMALL APERTURE



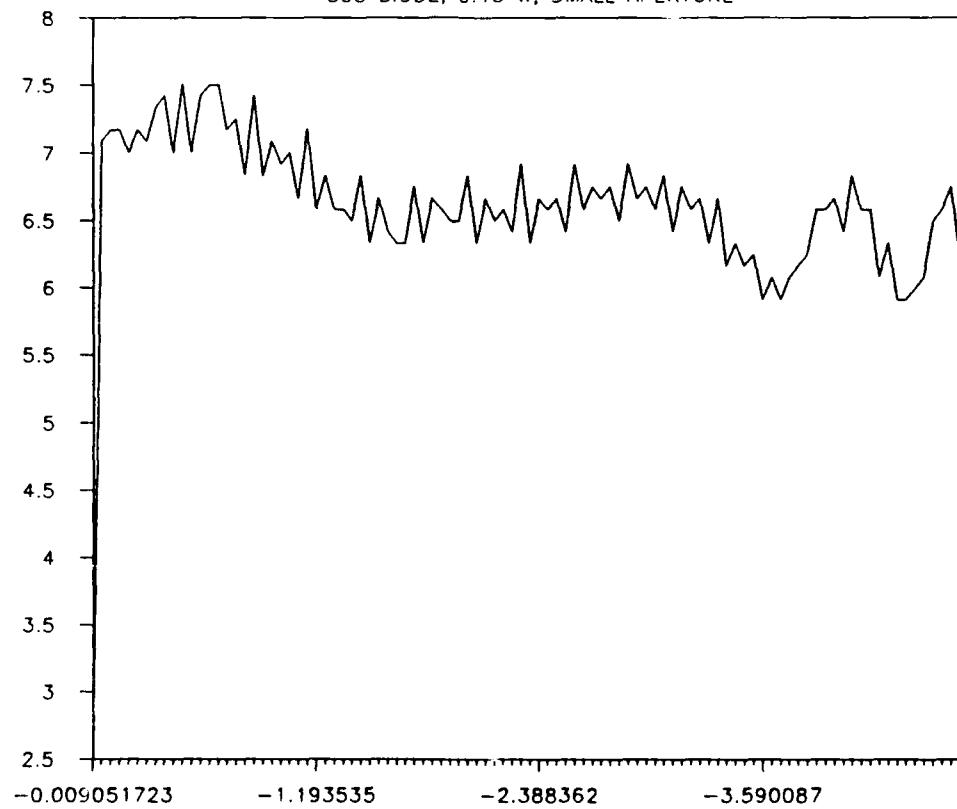
DEC19D-2X ICG SET=50, TC MAX=20

808 DIODE, 0.15 W, SMALL APERTURE



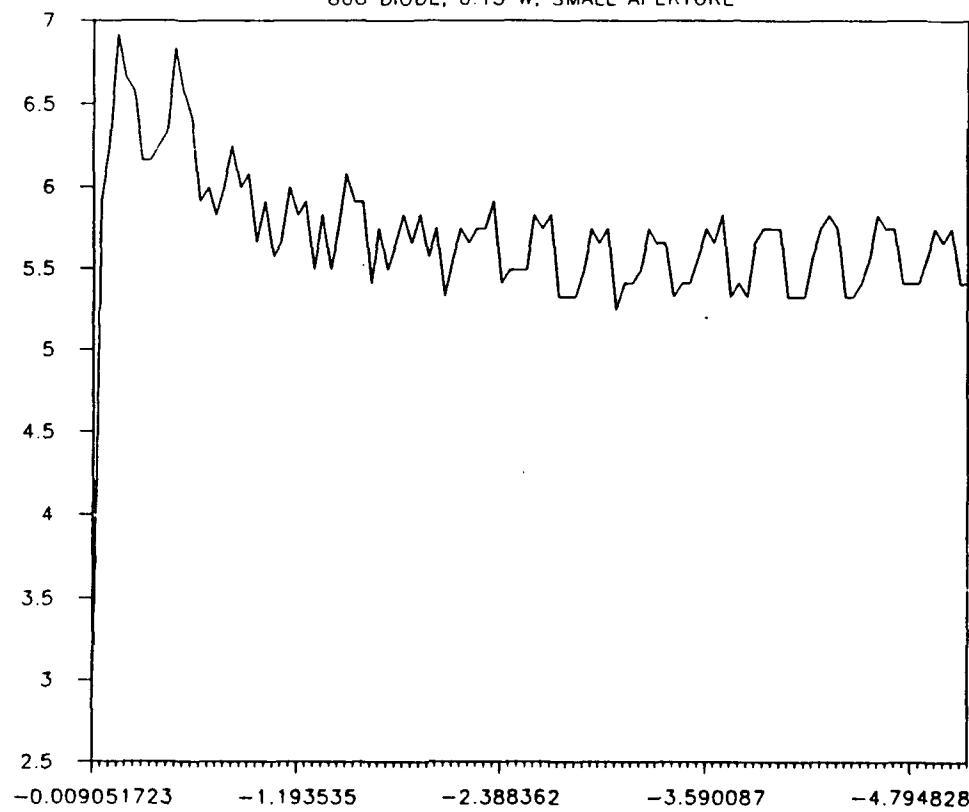
DEC19D-2X ICG SET=50, TC MAX=20

808 DIODE, 0.15 W, SMALL APERTURE



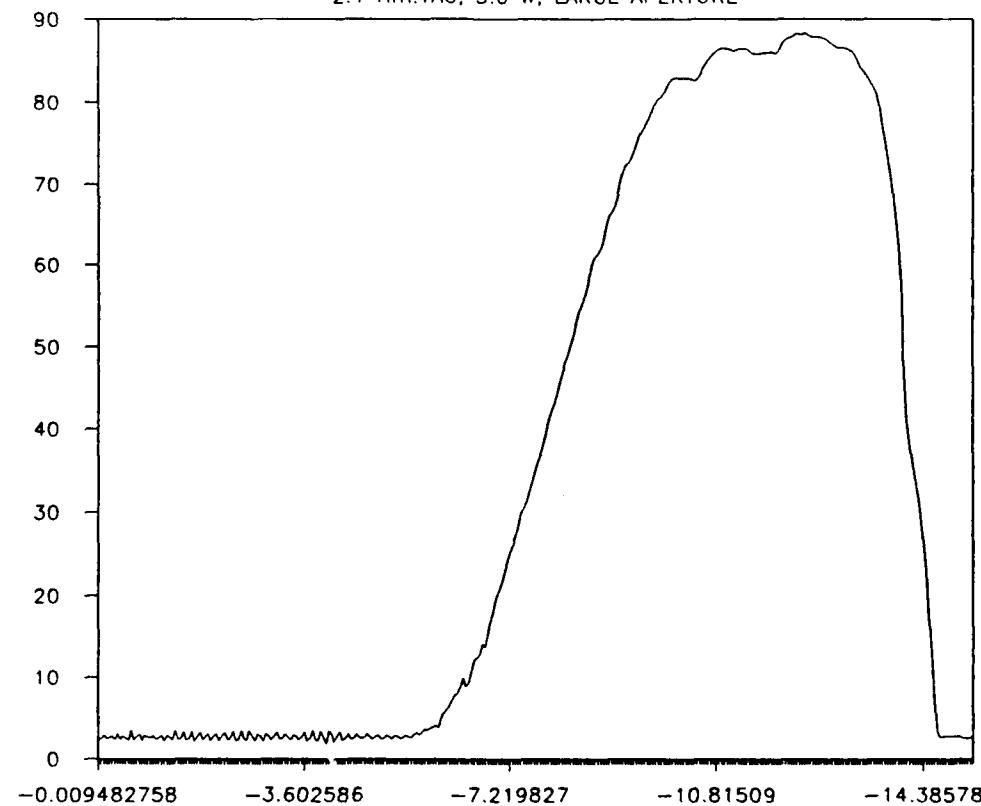
DEC19E-2X ICG SET=100, TC MAX=20

808 DIODE, 0.15 W. SMALL APERTURE



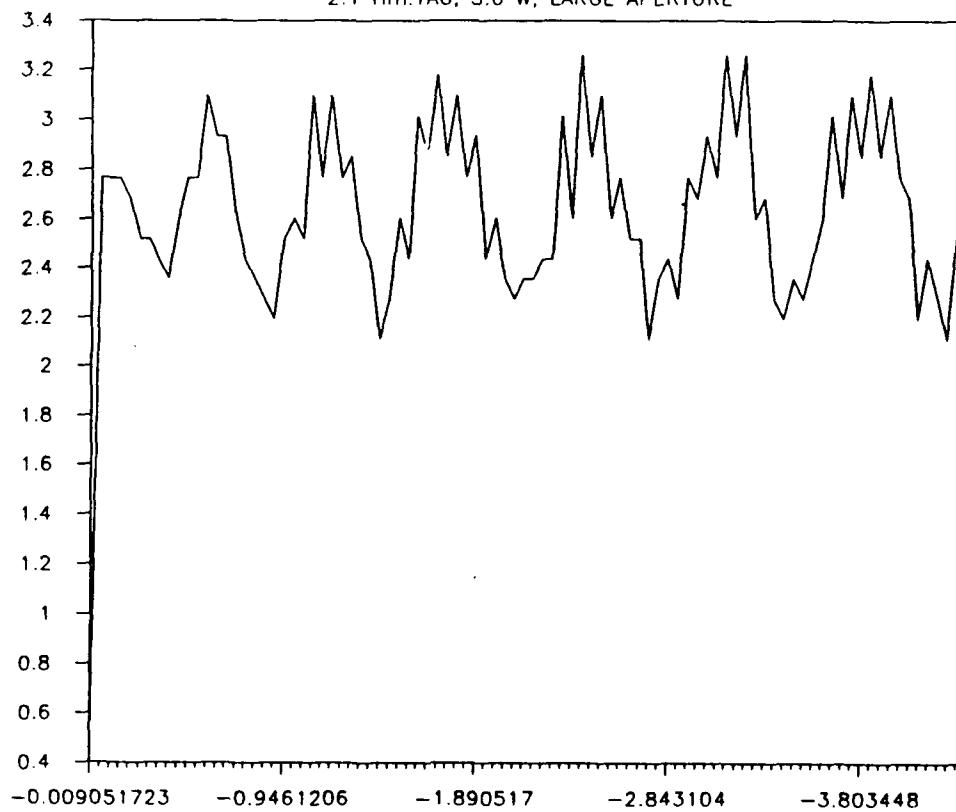
DEC19F-ONE STRIP UNWELDED CONTROL

2.1 Hm:YAG, 5.0 W, LARGE APERTURE



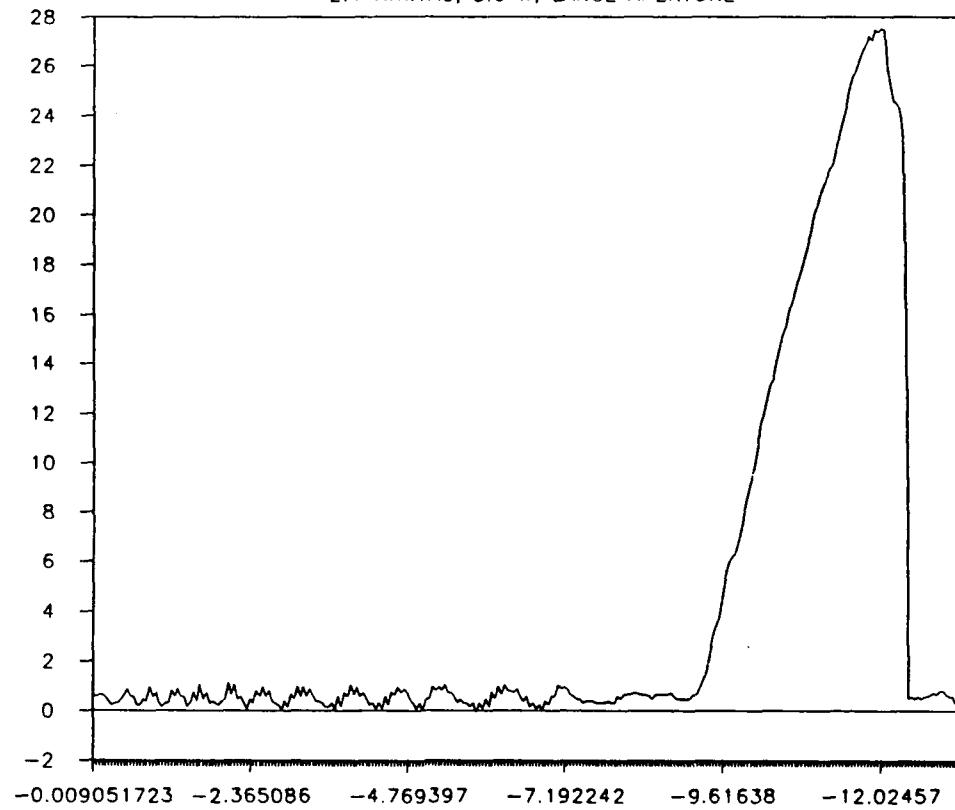
DEC19G-TWO STRIP UNWELDED CONTROL

2.1 Hm:YAG, 5.0 W, LARGE APERTURE



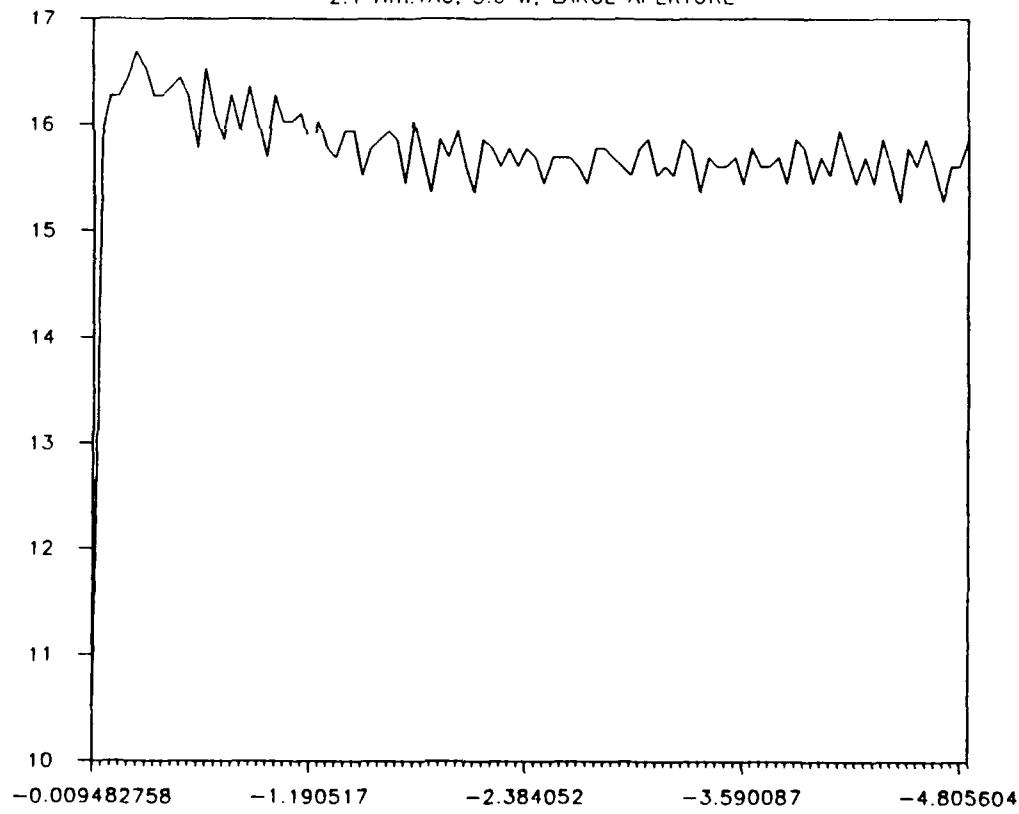
DEC19H-NO CHROMO CONTROLLED AT 50, 10 s

2.1 Hm:YAG, 5.0 W, LARGE APERTURE



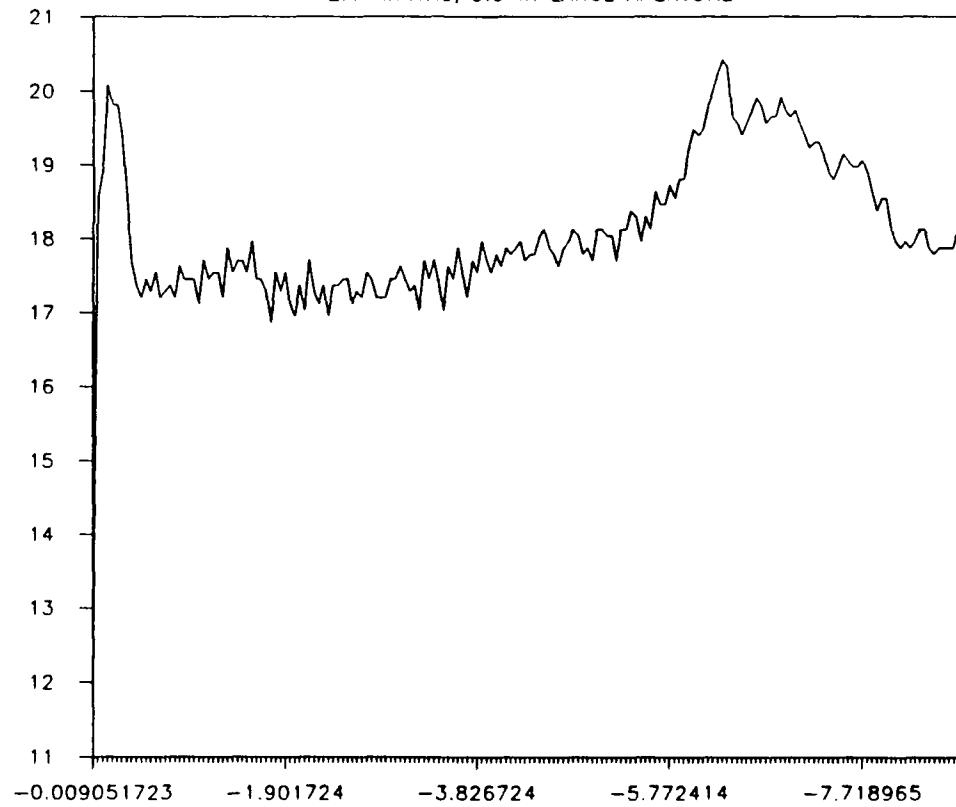
DEC19I-NO CHROMO CONTROLLED AT 60, 10s

2.1 Hm:YAG, 5.0 W, LARGE APERTURE



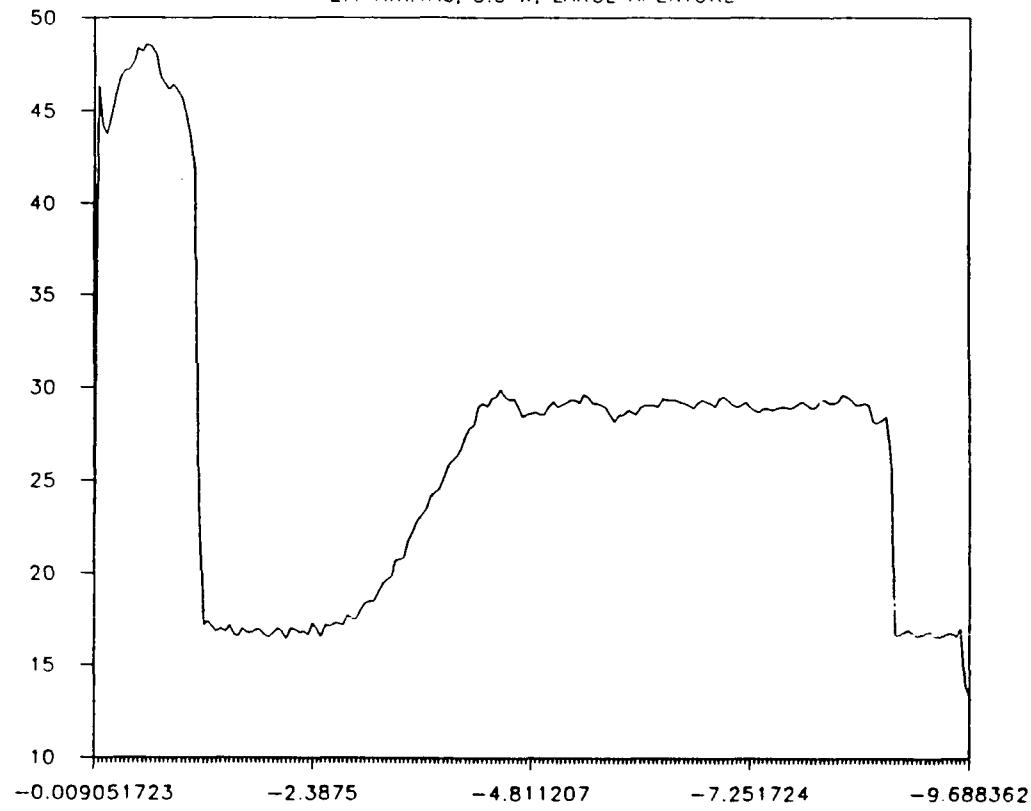
DEC19J-NO CHROMO CONTROLLED AT 70, 10 s

2.1 Hm:YAG, 5.0 W. LARGE APERTURE



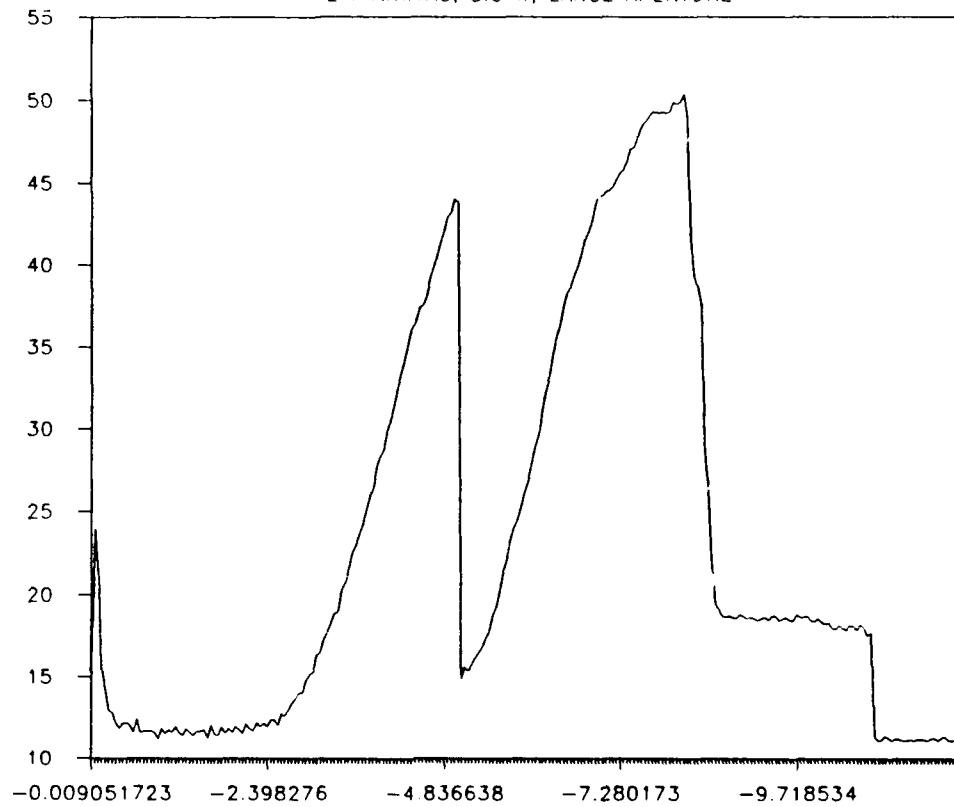
DEC19K- NO CHROMO CONTROLLED AT 80

2.1 Hm:YAG, 5.0 W, LARGE APERTURE



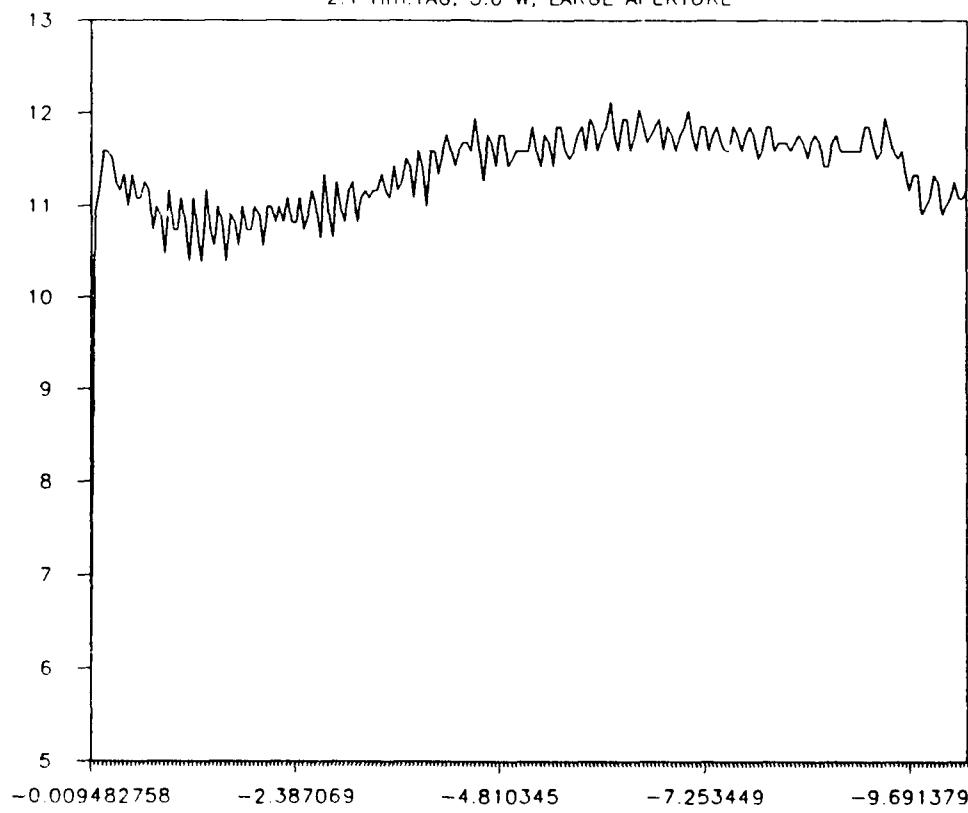
DEC19L-NO CHROMO CONTROLLED AT 100, 10s

21 Hz:YAG, 5.0 W, LARGE APERTURE



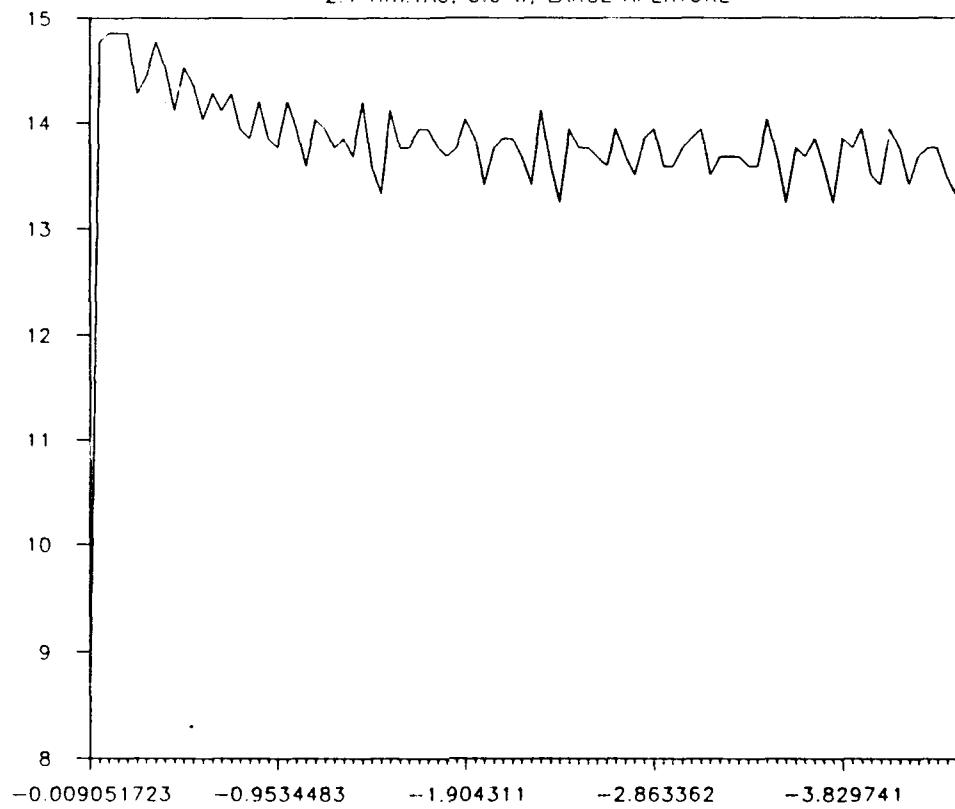
DEC19M- NO CHROMO CONTROLLED AT 50, 10s

2.1 Hm:YAG, 5.0 W, LARGE APERTURE



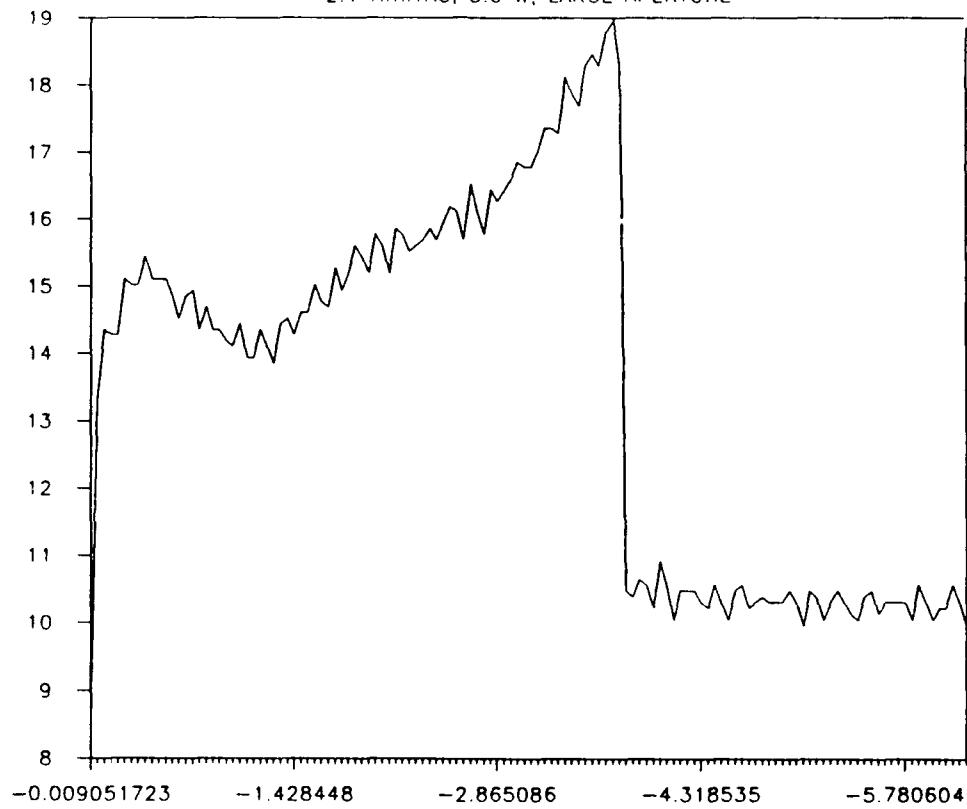
DEC19N- INDIA INK UNWELDED CONTROL

2.1 Hm:YAG, 5.0 W, LARGE APERTURE



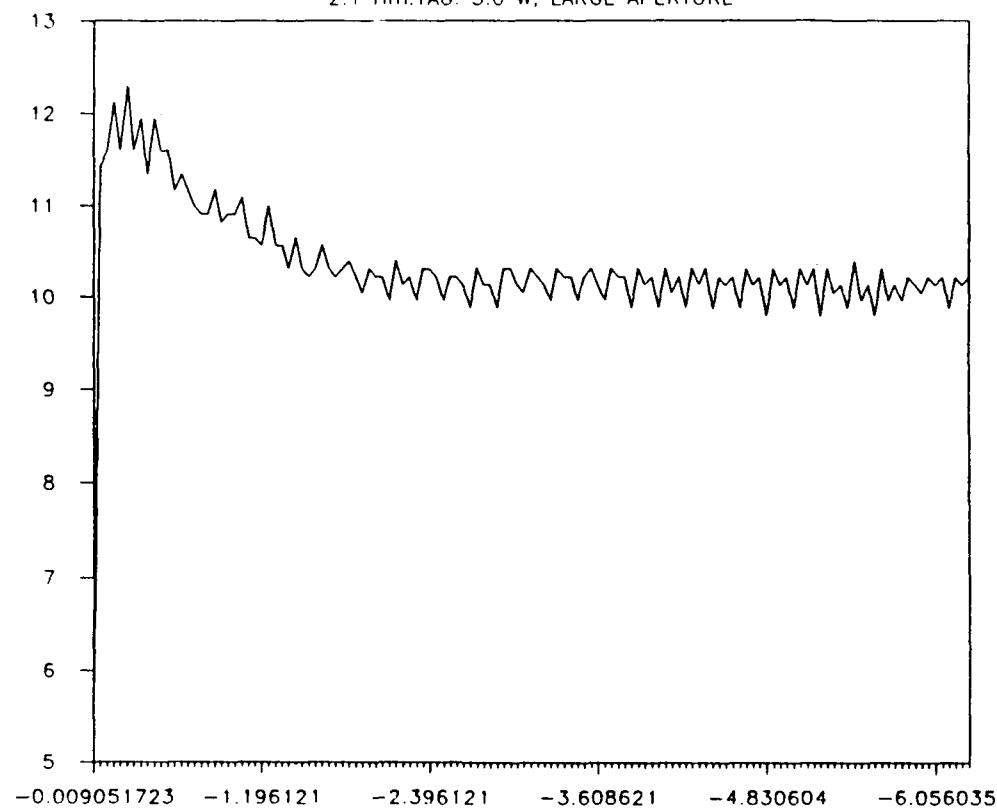
DEC190-INDIA INK CONTROLLED AT 50, 10s

2.1 Hm:YAG, 5.0 W, LARGE APERTURE



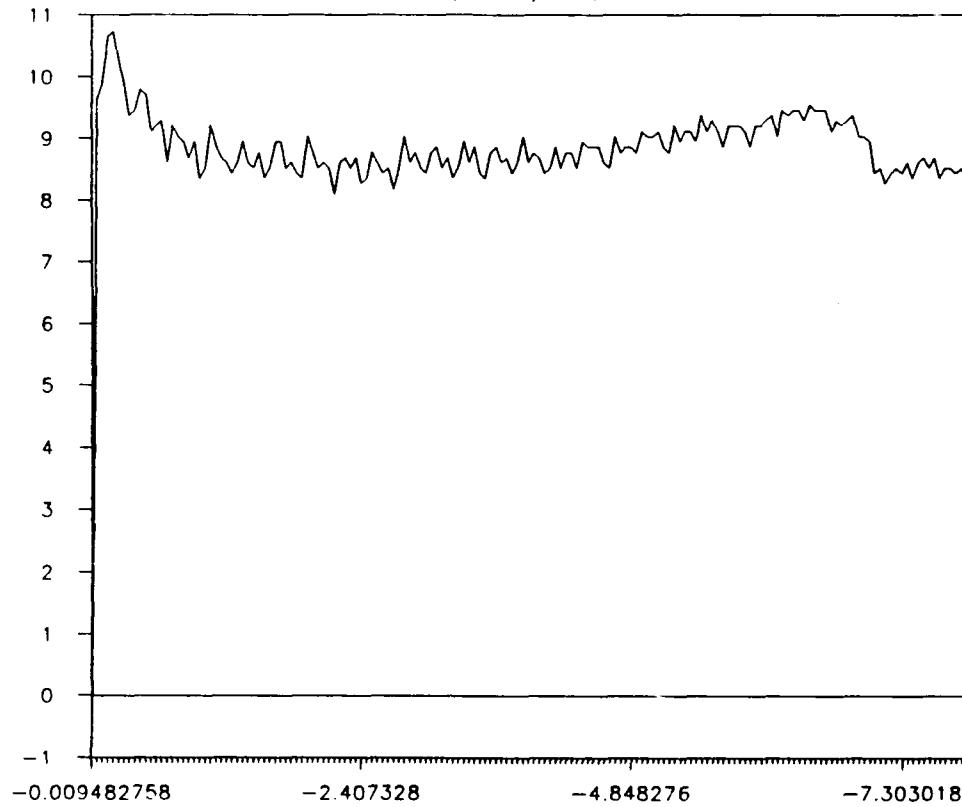
DEC19P1-INDIA INK CONTROLLED AT 50, '0s

2.1 Hm:YAG, 5.0 W, LARGE APERTURE



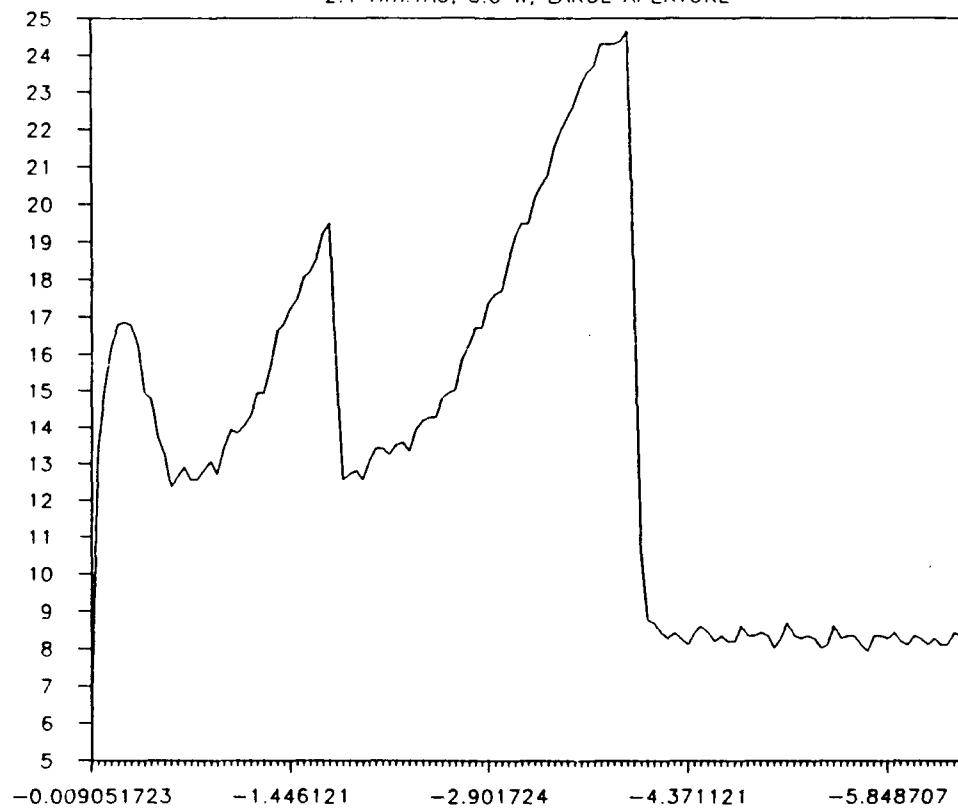
DEC19Q-INDIA INK CONTROLLED AT 60, 10s

2.1 Hm:YAG, 5.0 W, LARGE APERTURE



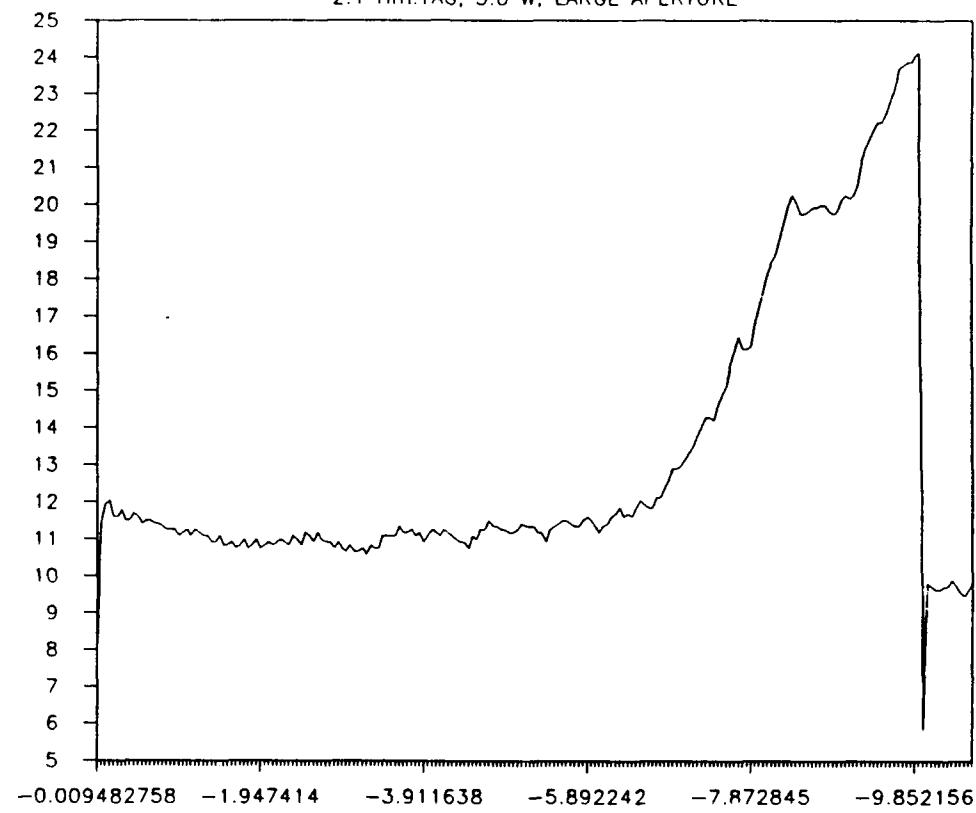
DEC19R- INDIA INK CONTROLLED AT 70, 10s

2.1 Hm:YAG, 5.0 W, LARGE APERTURE



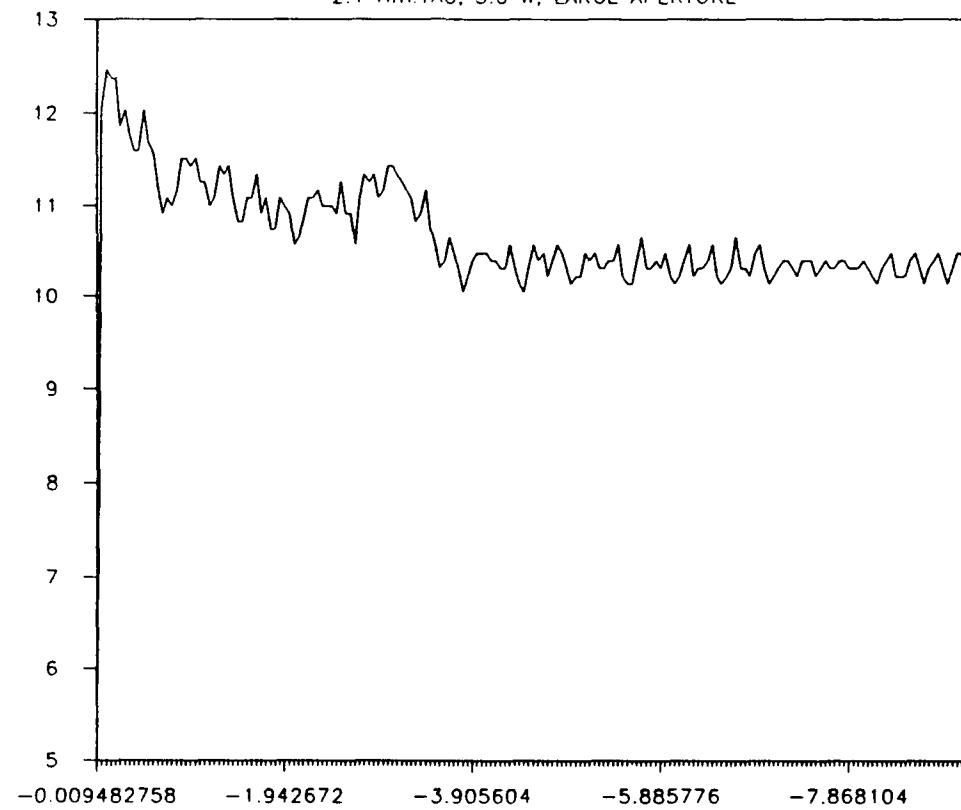
DEC19S- INDIA INK CONTROLLED AT 80, 10s

2.1 Hm:YAG, 5.0 W, LARGE APERTURE



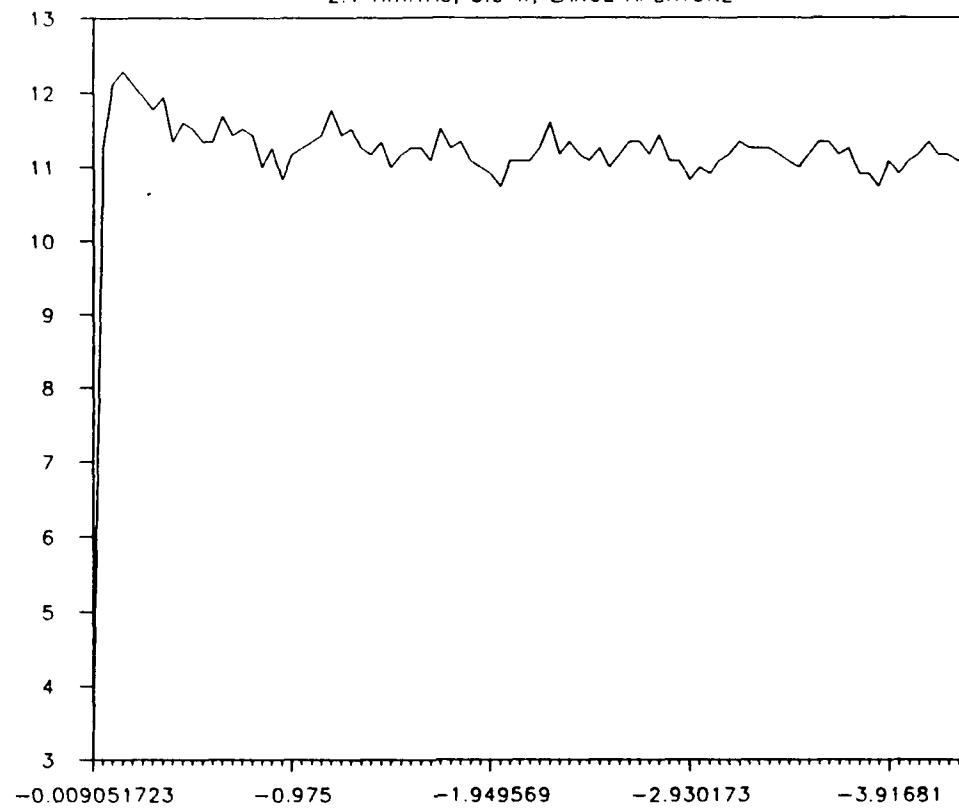
DEC19T-INDIA INK CONTROLLED AT 100, 10s

2.1 Hz YAG, 5.0 W, LARGE APERTURE



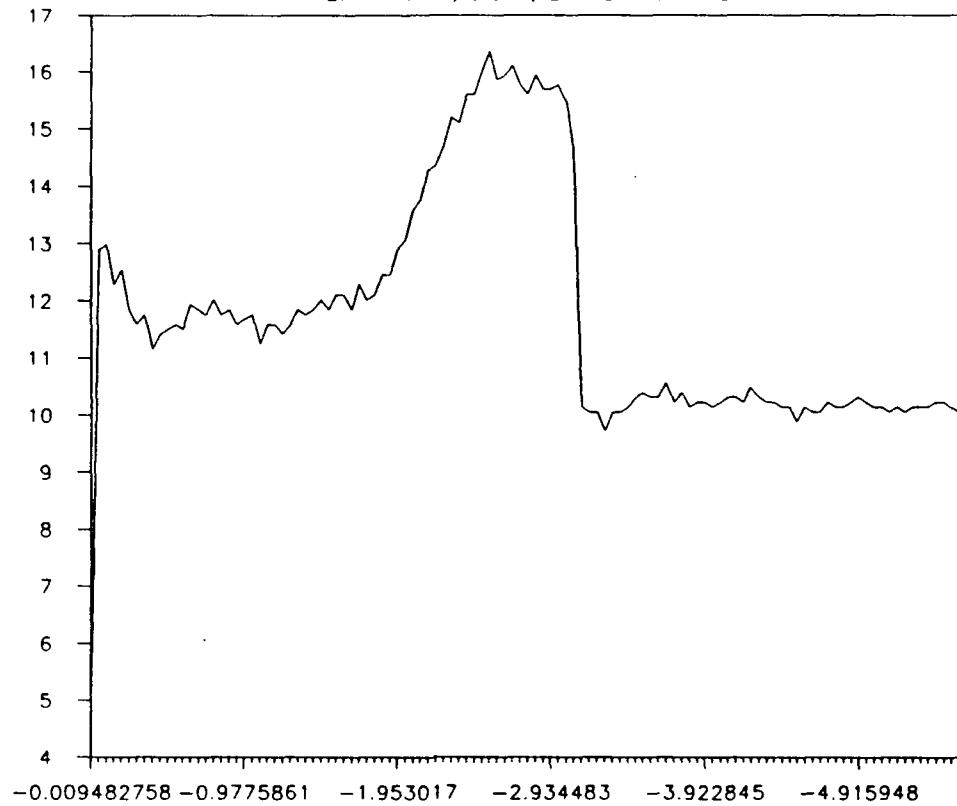
DEC19U-2X iCG UNWELDED CONTROL

2.1 Hm:YAG, 5.0 W, LARGE APERTURE



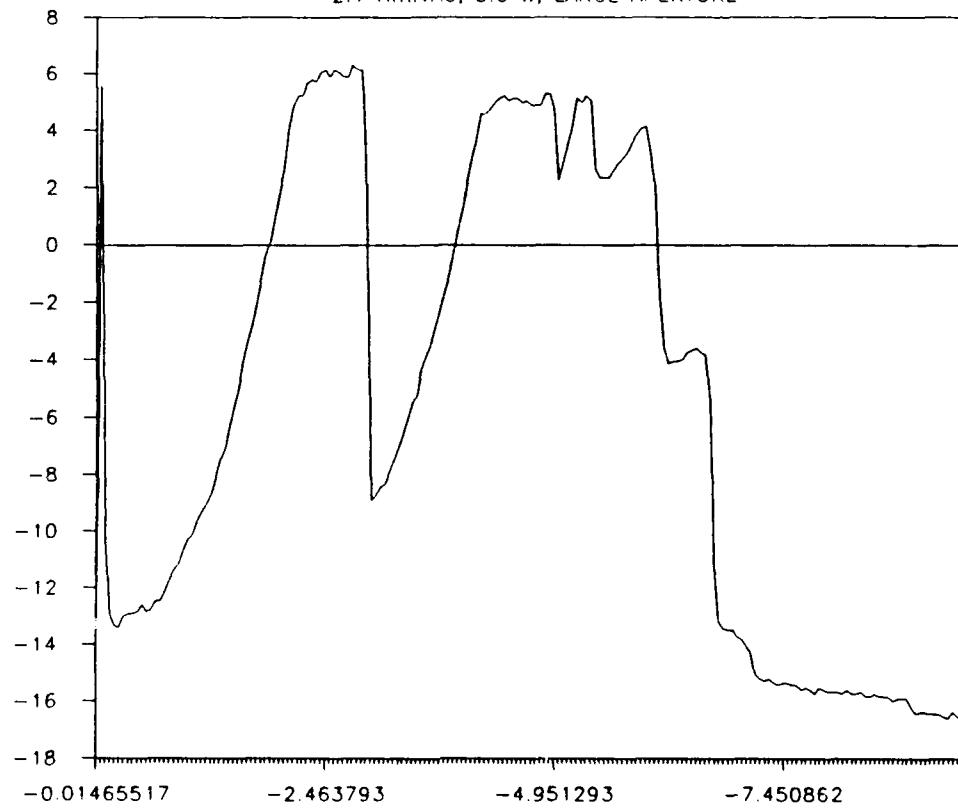
DEC19V- 2X ICG CONTROLLED AT 50, 10s

2.1 Hm:YAG, 5.0 W, LARGE APERTURE

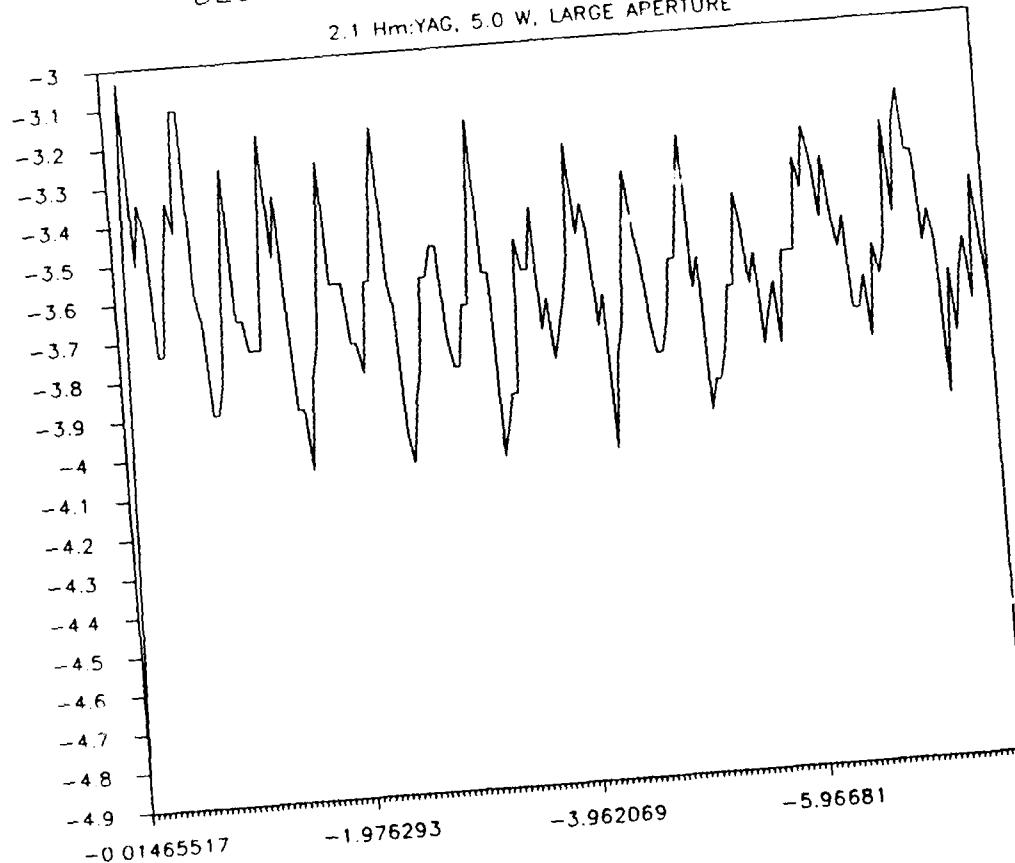


DEC19W-2X ICG CONTROLLED AT 60, 10s

2.1 Hm:YAG, 5.0 W, LARGE APERTURE

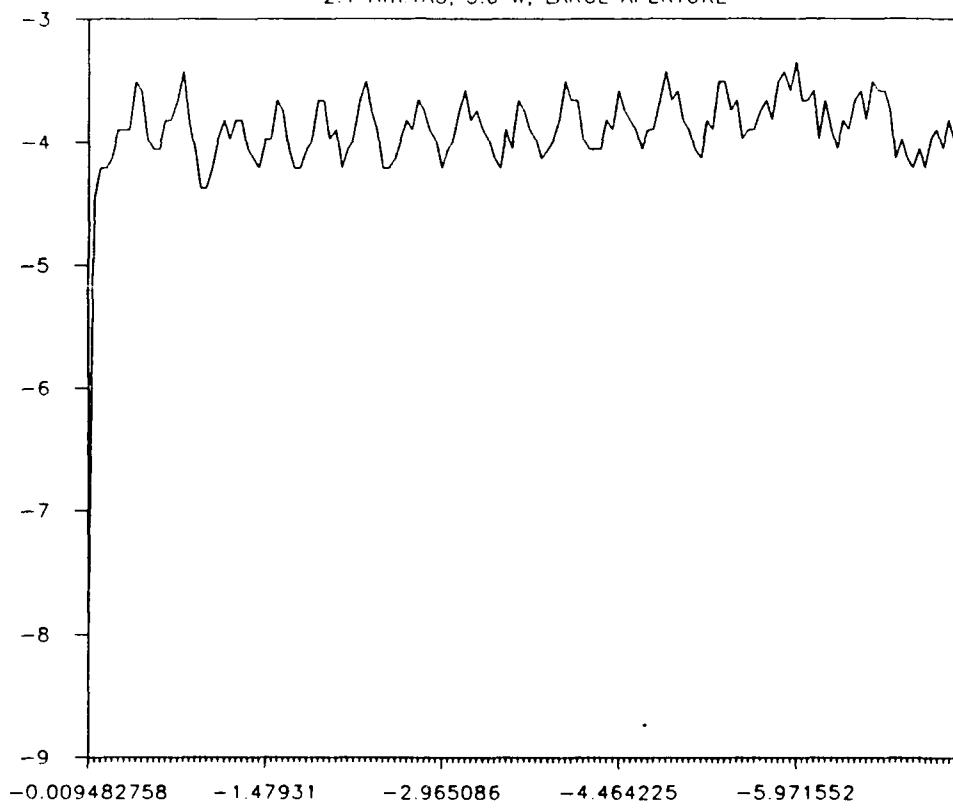


DEC19X-2X ICG CONTROLLED AT 70, 10s
2.1 Hm:YAG, 5.0 W, LARGE APERTURE



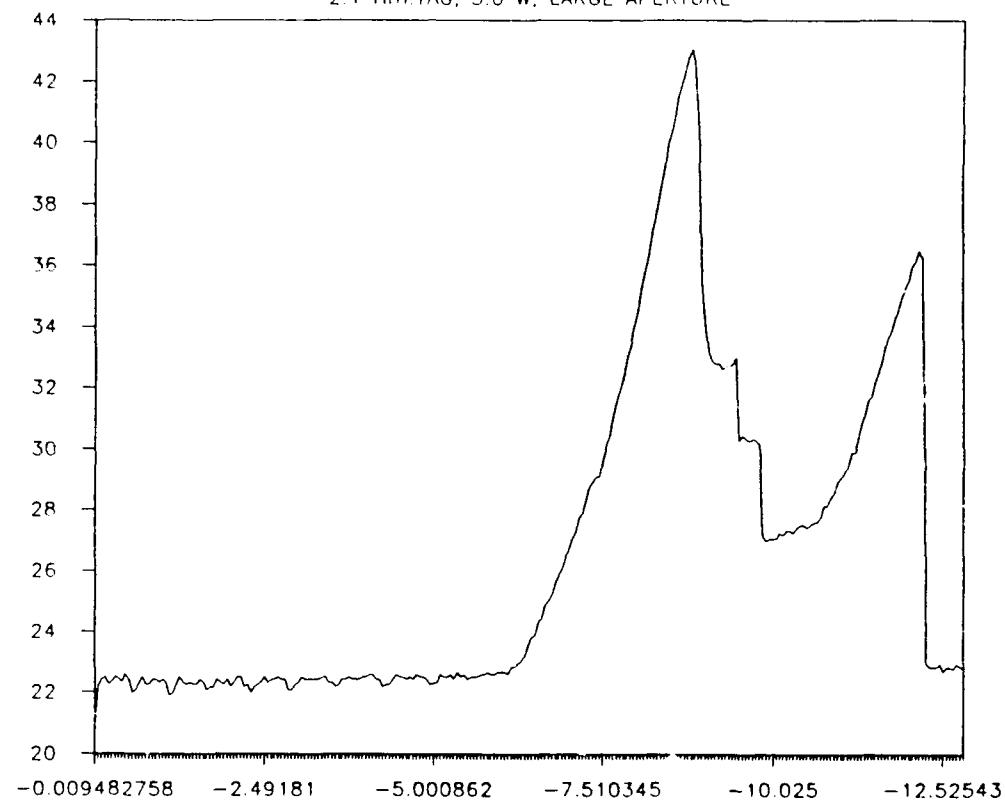
DEC19Y- 2X ICG CONTROLLED AT 80, 10s

2.1 Hm:YAG, 5.0 W, LARGE APERTURE



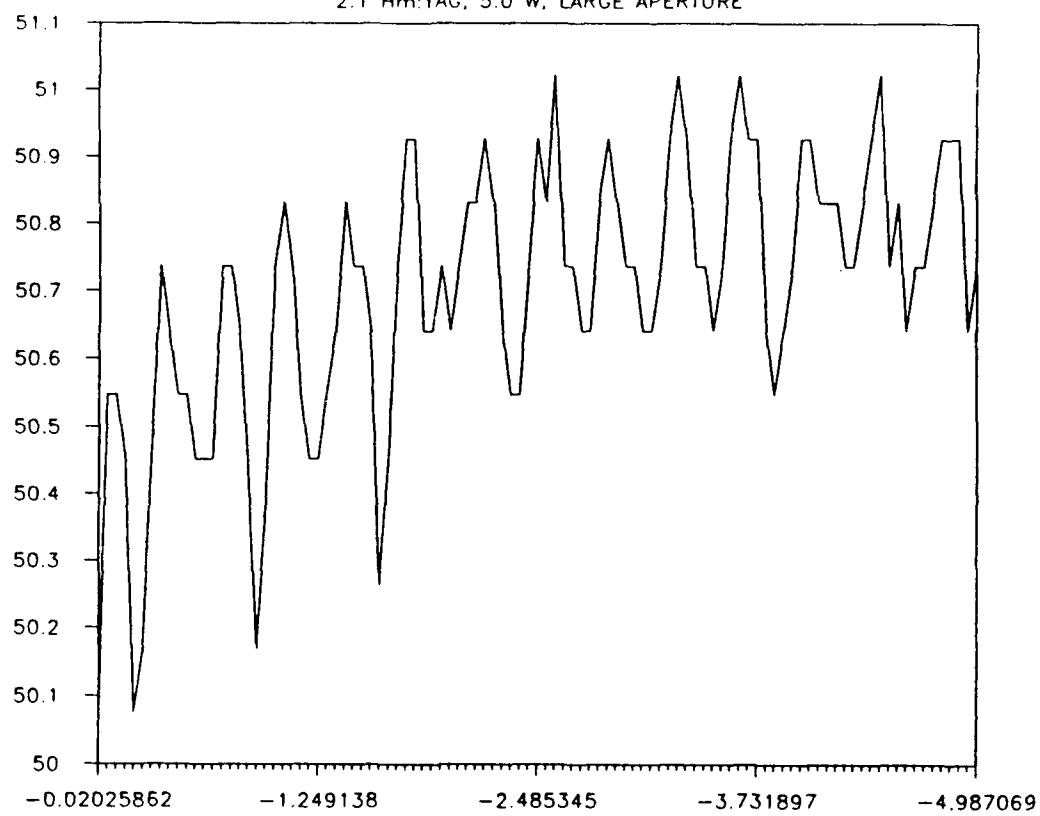
DEC19Z-2X ICG CONTROLLED AT 100, 10s

2.1 Hm:YAG, 5.0 W, LARGE APERTURE



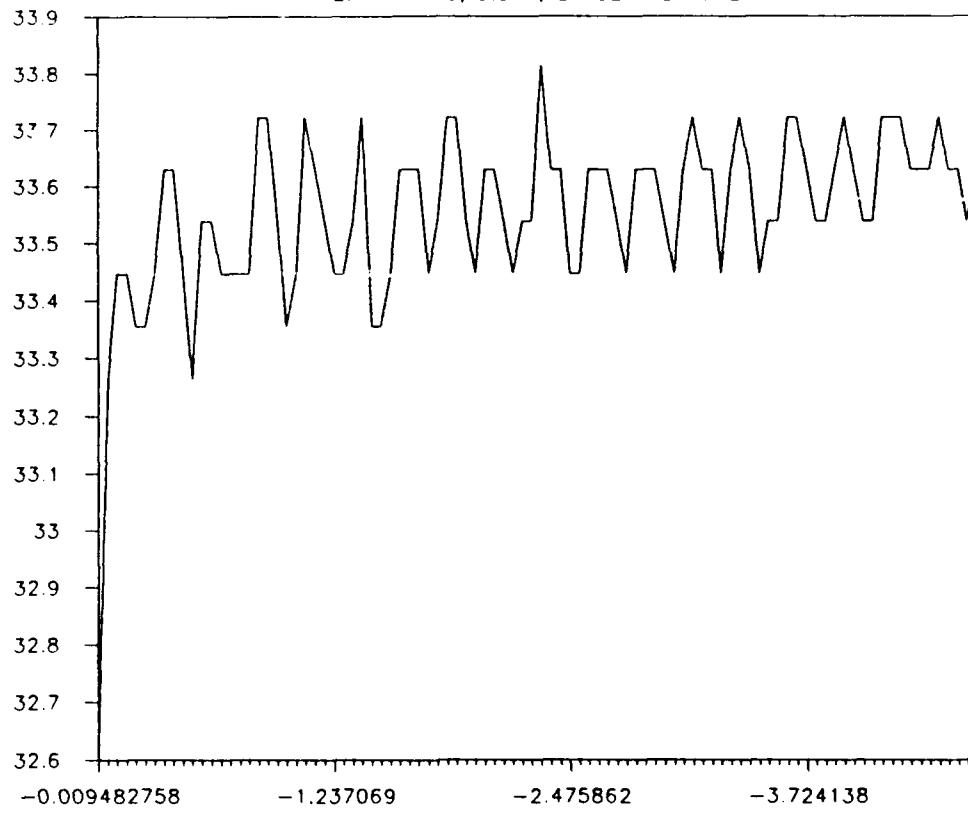
DEC19AA-BLOOD UNWELDED CONTROL

2.1 Hm:YAG, 5.0 W, LARGE APERTURE



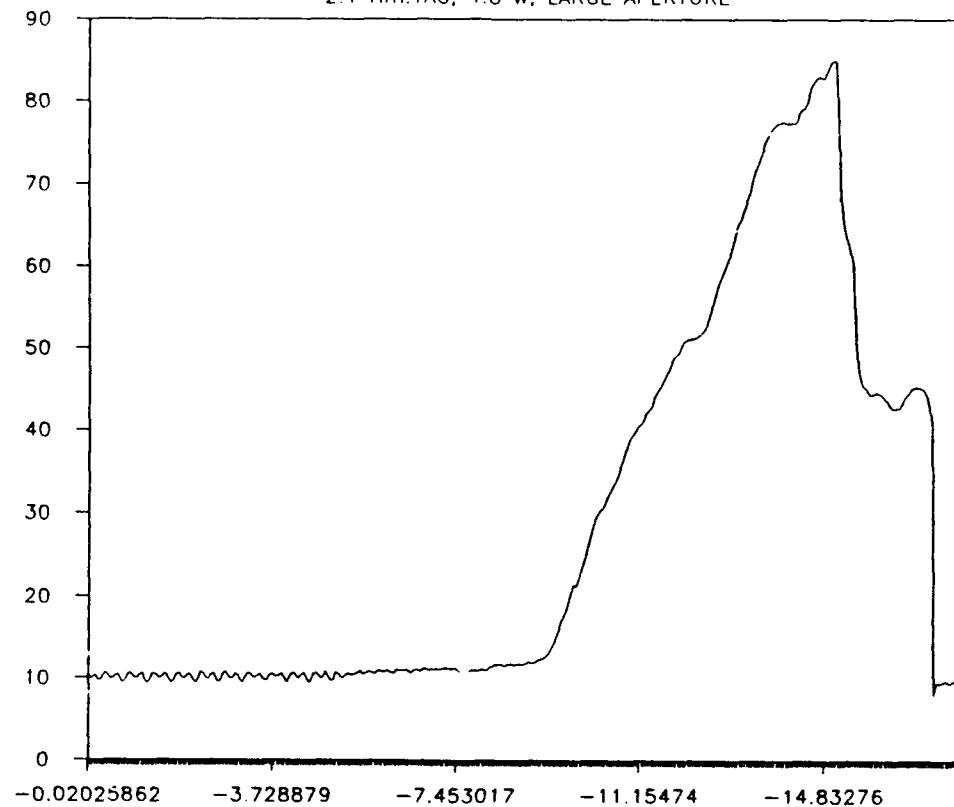
DEC19AB-BLOOD CONTROLLED AT 50, 10s

2.1 Hm:YAG, 5.0 W, LARGE APERTURE



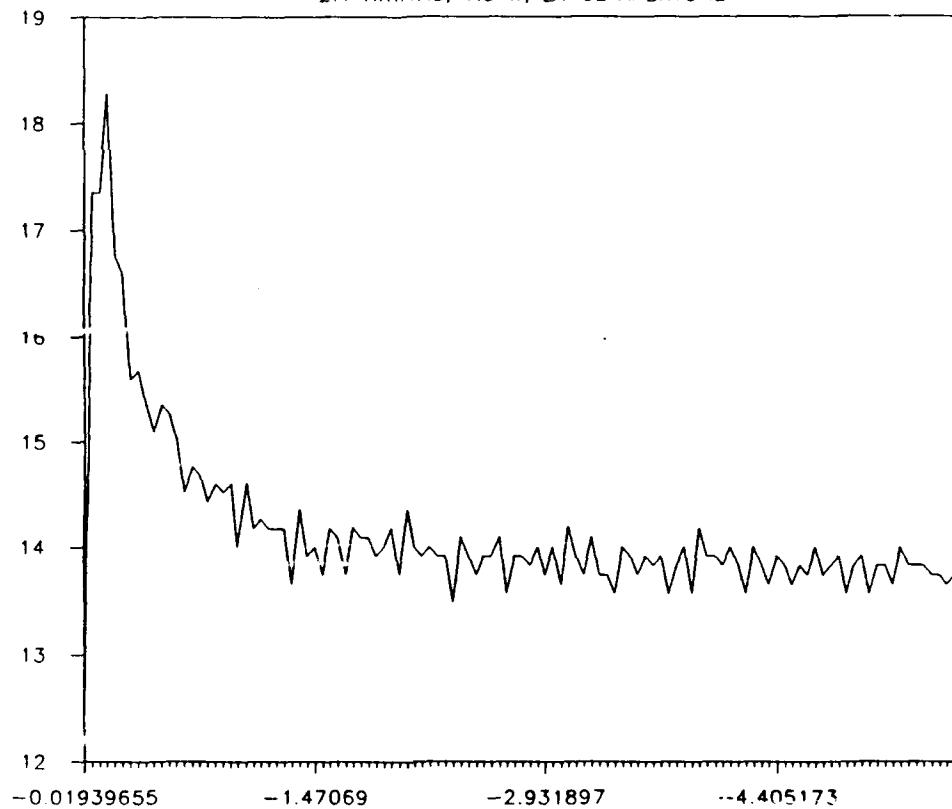
DEC21A-ONE STRIP UNWELDED CONTROL

2.1 Hm:YAG, 4.8 W, LARGE APERTURE



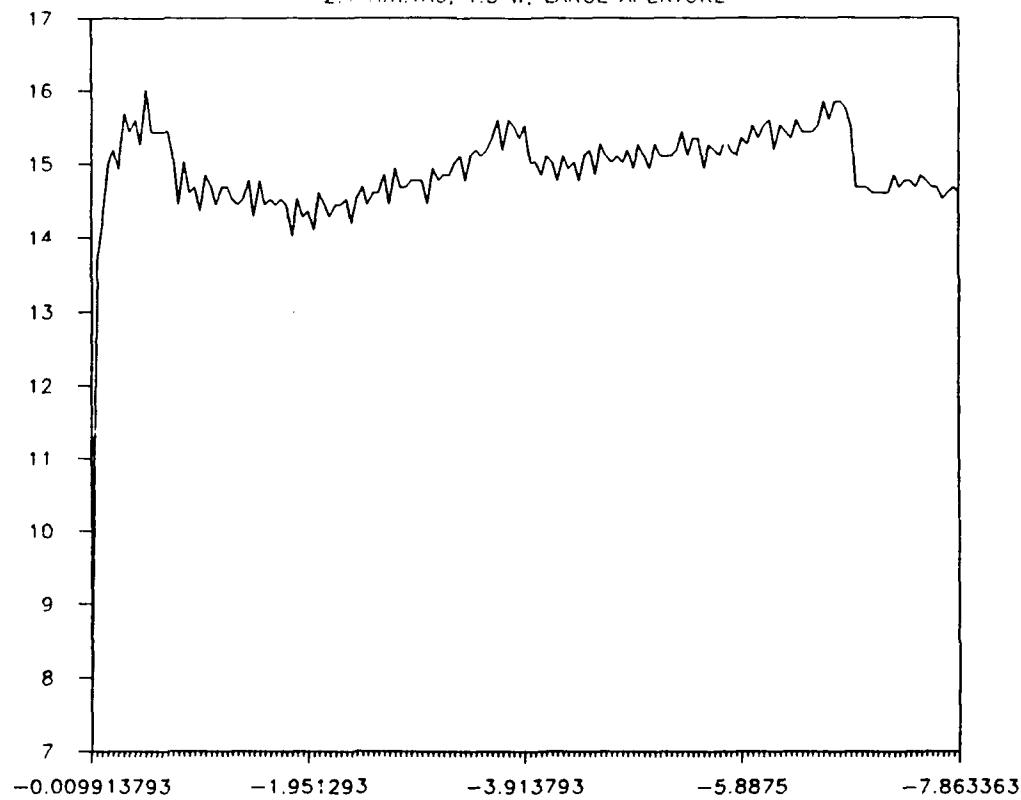
DEC21AA1-NO CHROMO CONTROLLED AT 70,10s

2.1 Hm:YAG, 1.5 W, LARGE APERTURE



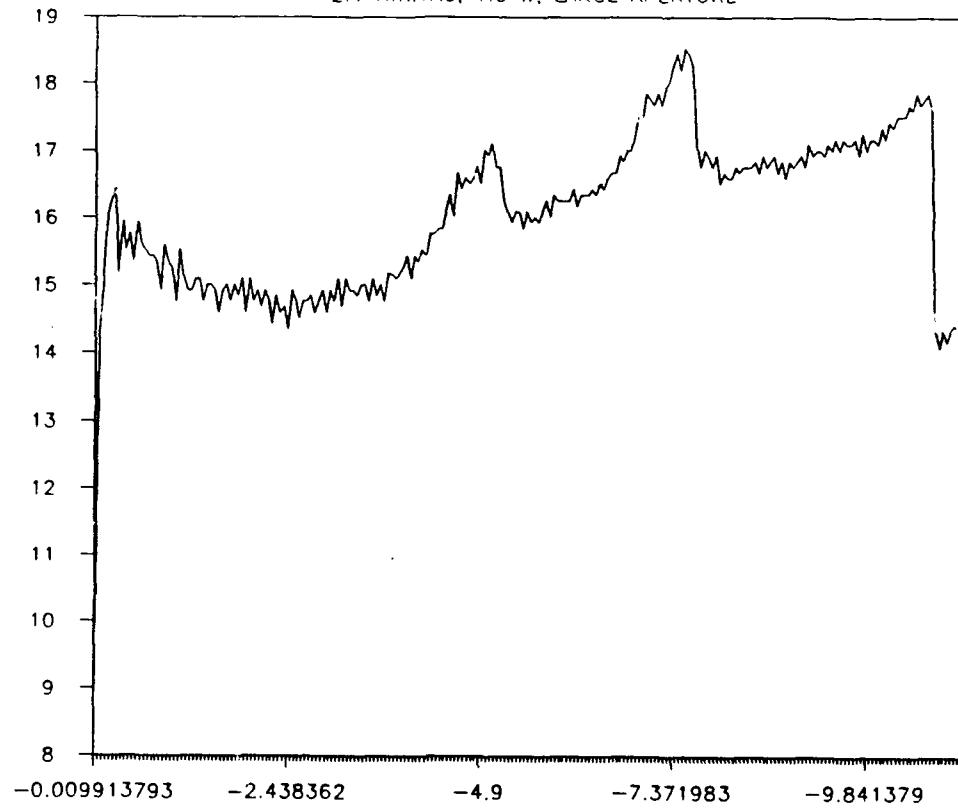
DEC21AB-NO CHROMO CONTROLLED AT 80, 10s

2.1 Hm:YAG, 1.5 W, LARGE APERTURE



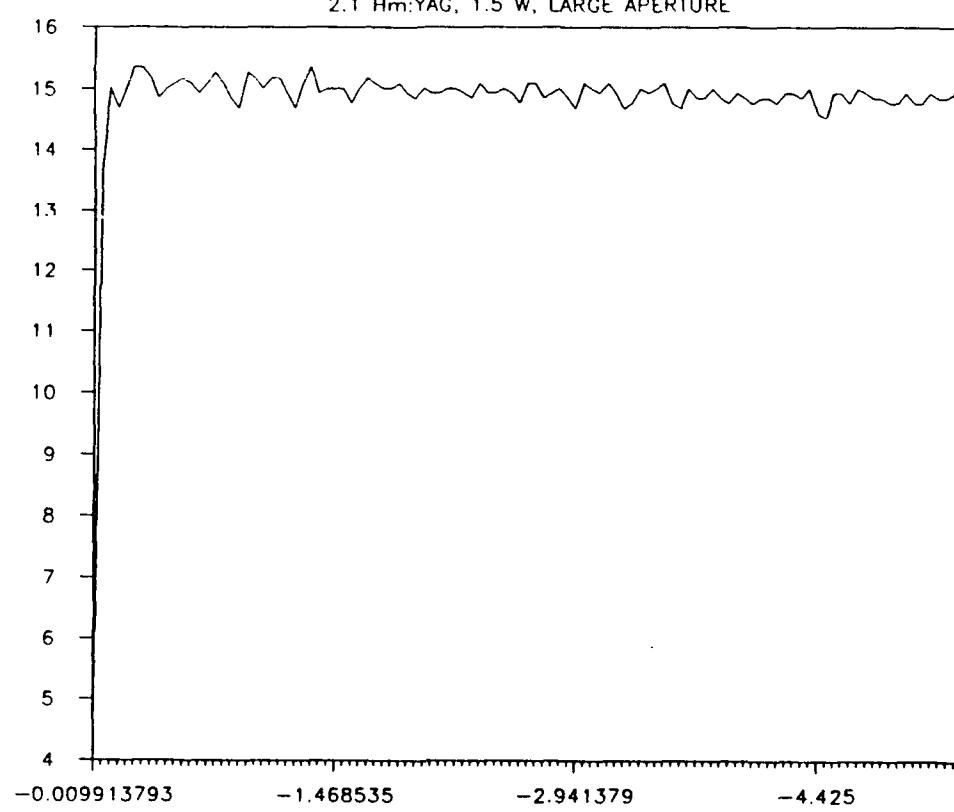
DEC21AC-NO CHROMO CONTROLLED AT 100,10s

2.1 Hm:YAG, 1.5 W, LARGE APERTURE



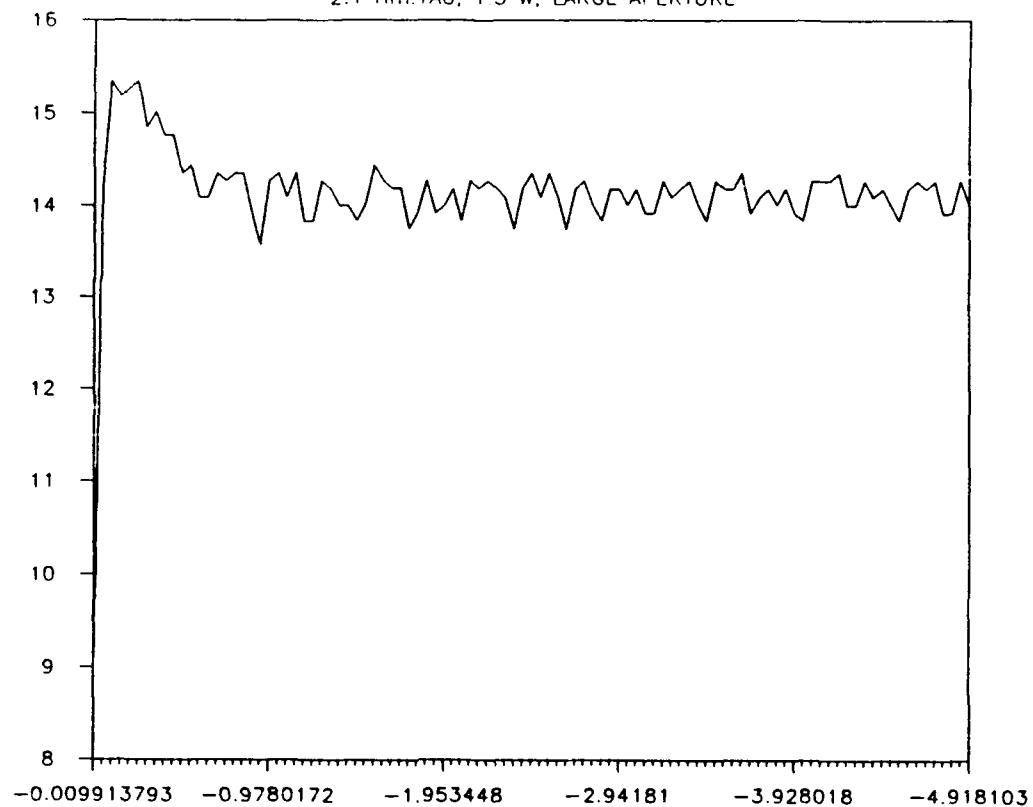
DEC21AD-INDIA INK UNWELDED CONTROL

2.1 Hm:YAG, 1.5 W, LARGE APERTURE



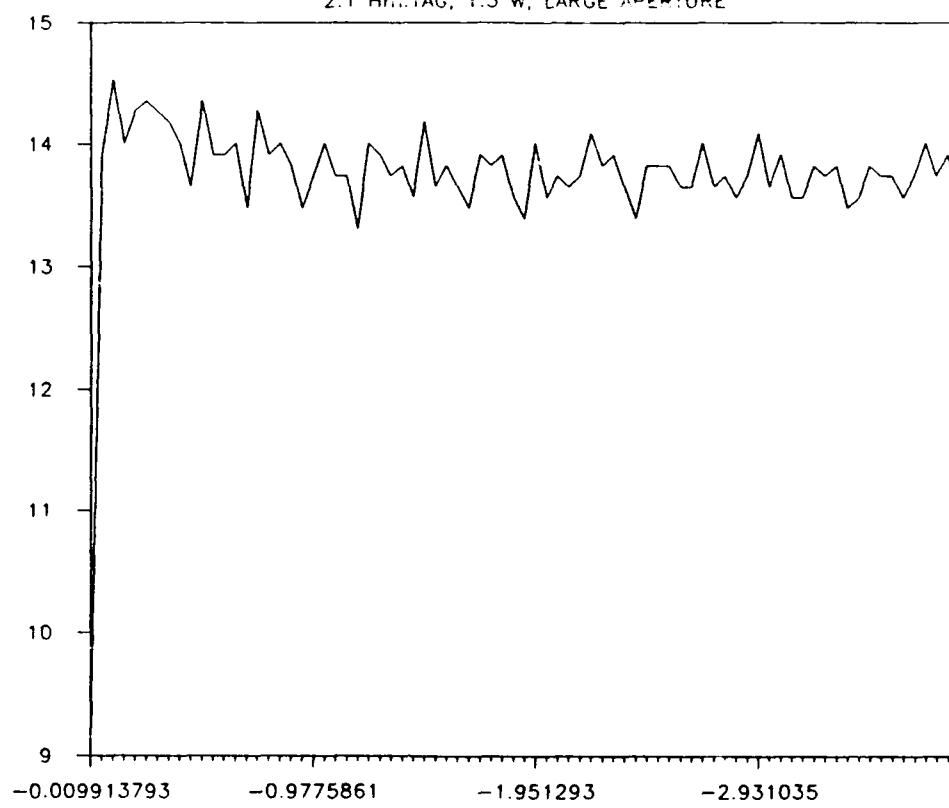
DEC21AE-INDIA INK CONTROLLED AT 50,10s

2.1 Hz:YAG, 1.5 W, LARGE APERTURE



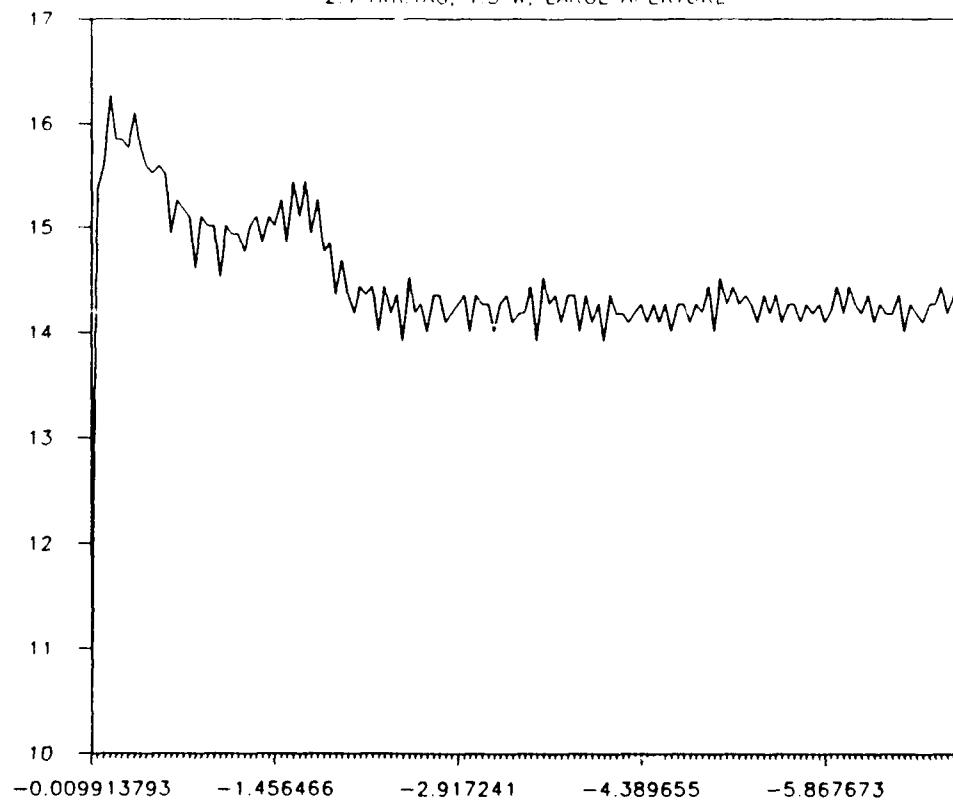
DEC21AF-INDIA INK CONTROLLED AT 60, 10s

2.1 Hz, YAG, 1.5 W, LARGE APERATURE



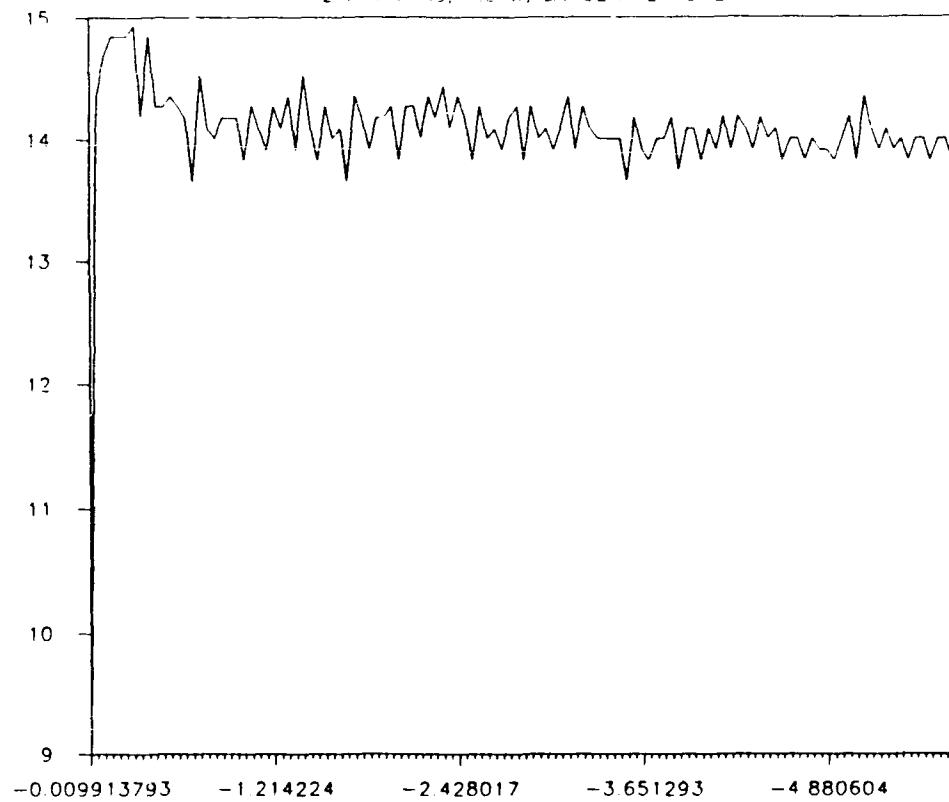
DEC21AG-INDIA INK CONTROLLED AT 70, 10s

2.1 Hz YAG, 1.5 W, LARGE APERTURE



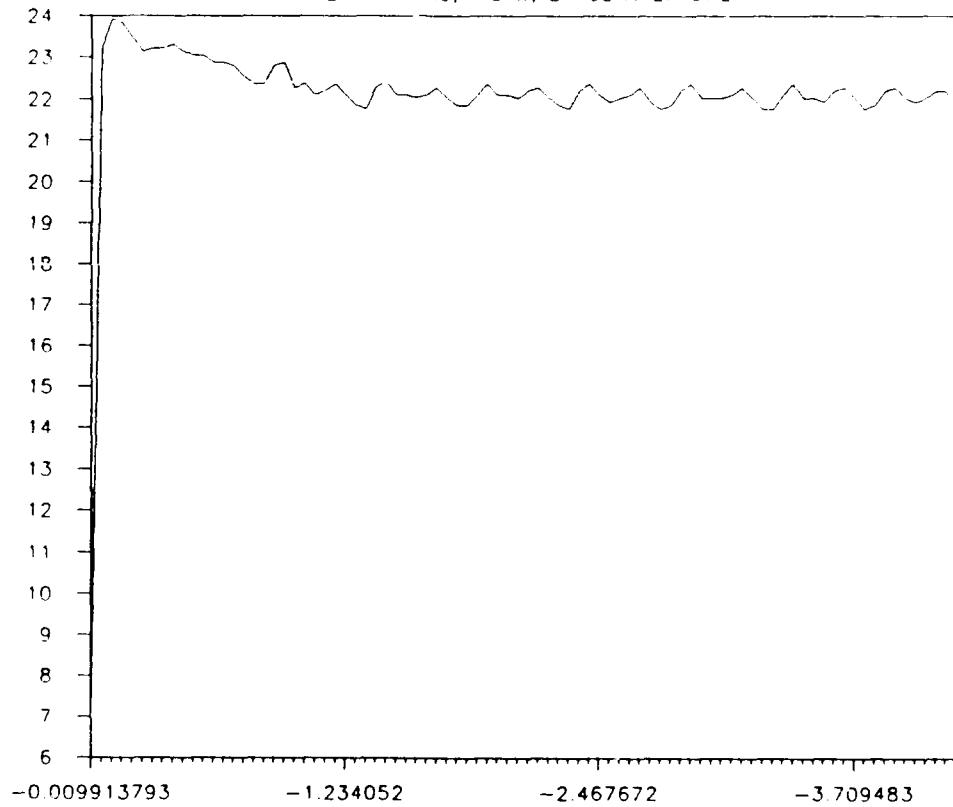
DEC21AH-INDIA INK CONTROLLED AT 80, 10s

21 Hz YAG, 1.5 W, LARGE APERTURE



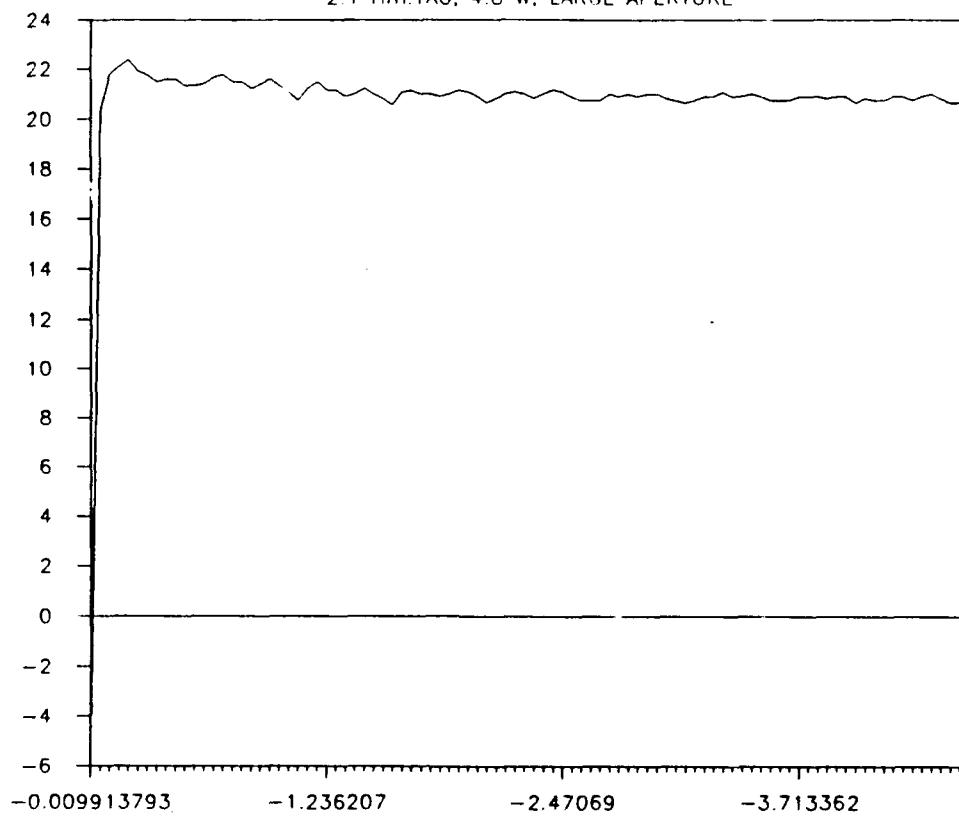
DEC21B-TWO STRIP UNWELDED CONTROL

21 Hm:YAG, 48 W, LARGE APERTURE



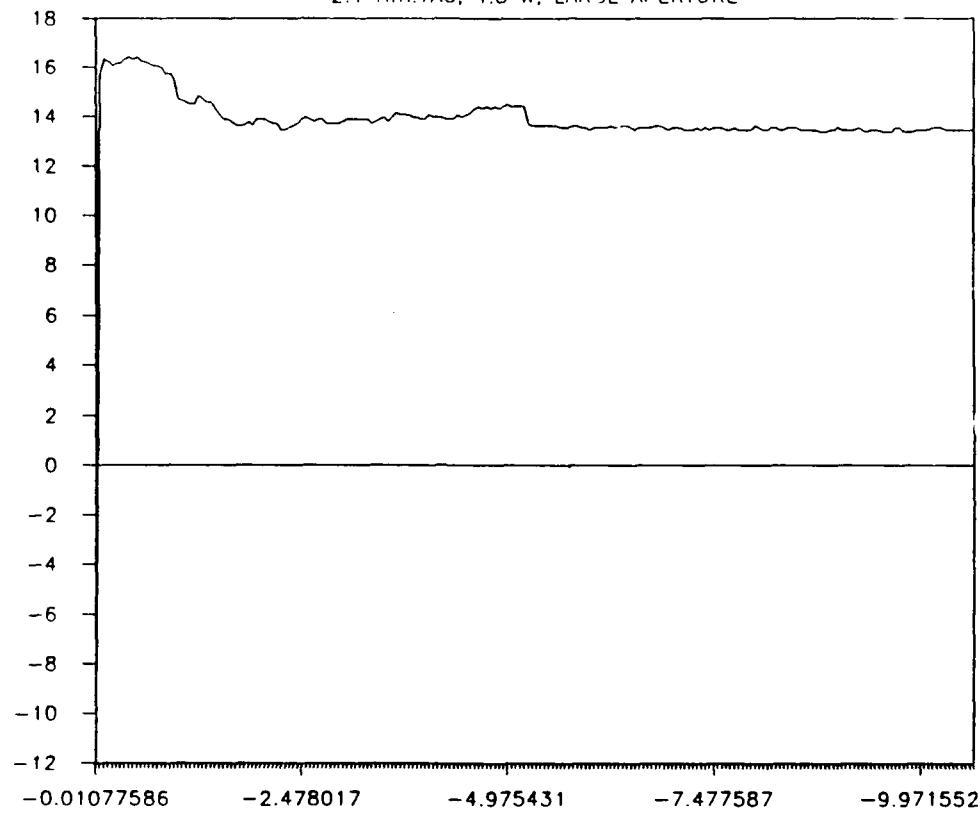
DEC21C-TWO STRIP 2X ICG UNWELDED CONTROL

2.1 Hm:YAG, 4.8 W, LARGE APERTURE



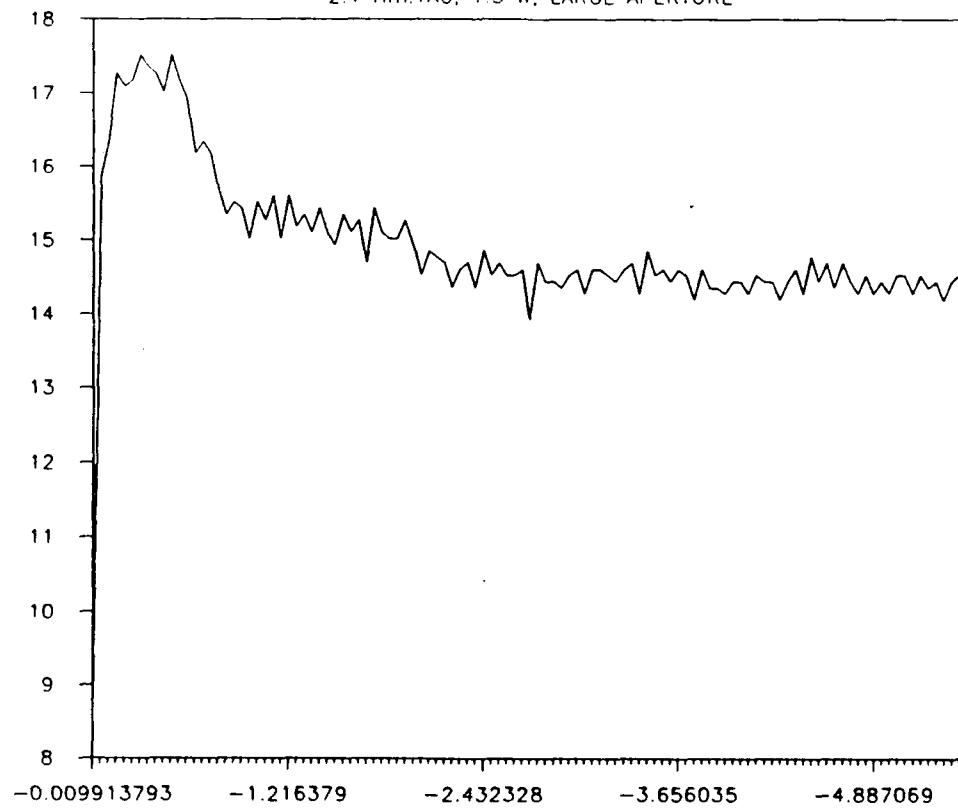
DEC21D-2X ICG CONTROLLED AT 50, 10s

2.1 Hm:YAG, 4.8 W, LARGE APERTURE



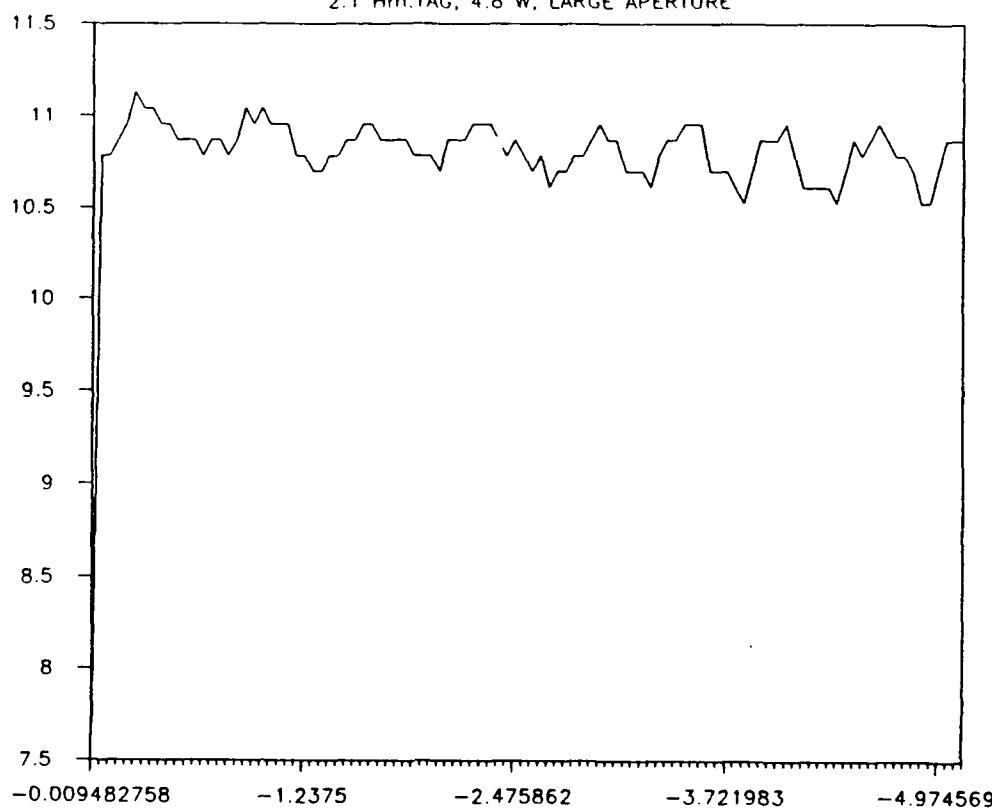
DEC21AI-INDIA INK CONTROLLED AT 100,10s

2.1 Hm:YAG, 1.5 W, LARGE APERTURE



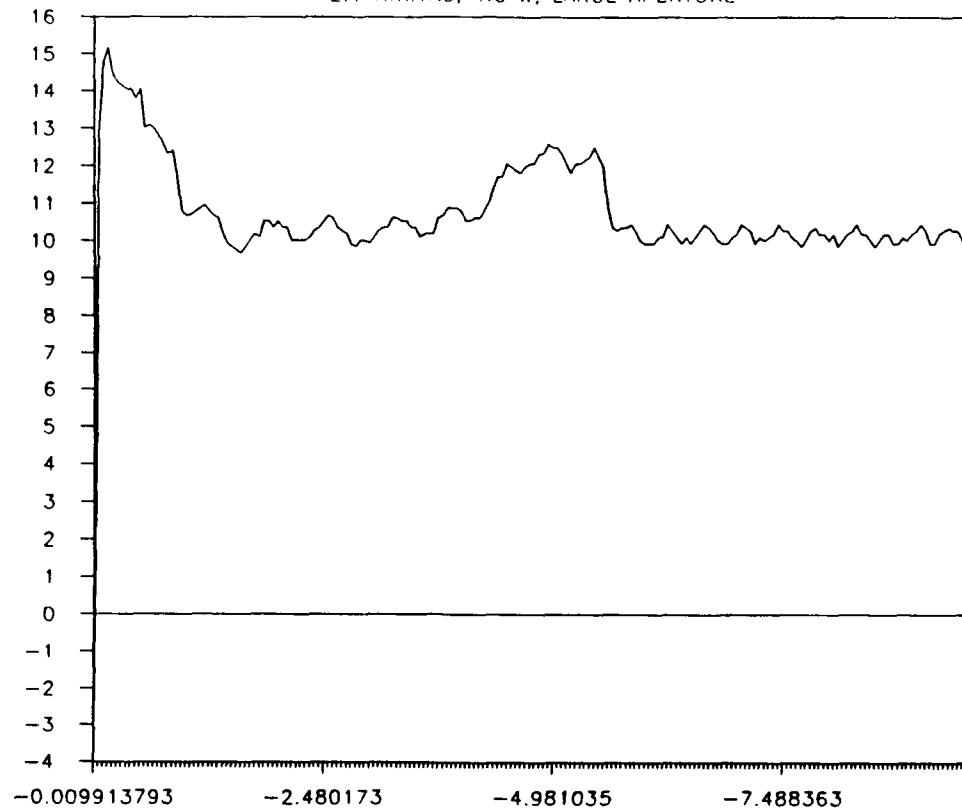
DEC21E-2X ICG CONTROLLED AT 60, 10s

2.1 Hm:YAG, 4.8 W, LARGE APERTURE



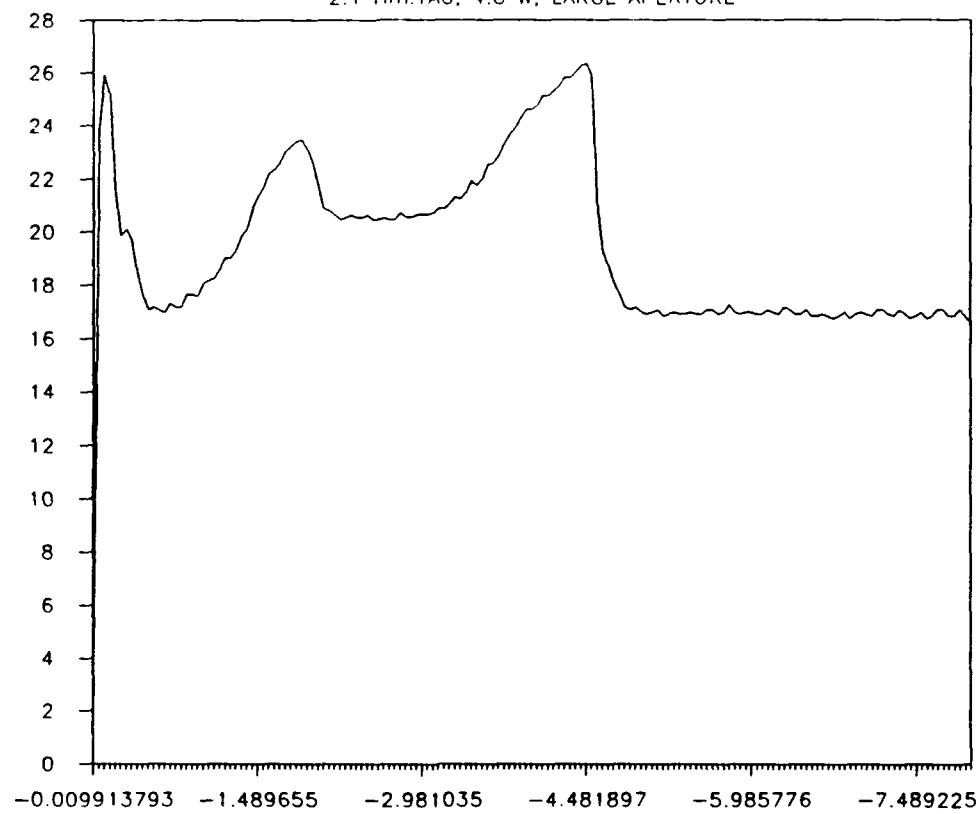
DEC21F-2X ICG CONTROLLED AT 70, 10s

2.1 Hm:YAG, 4.8 W, LARGE APERTURE



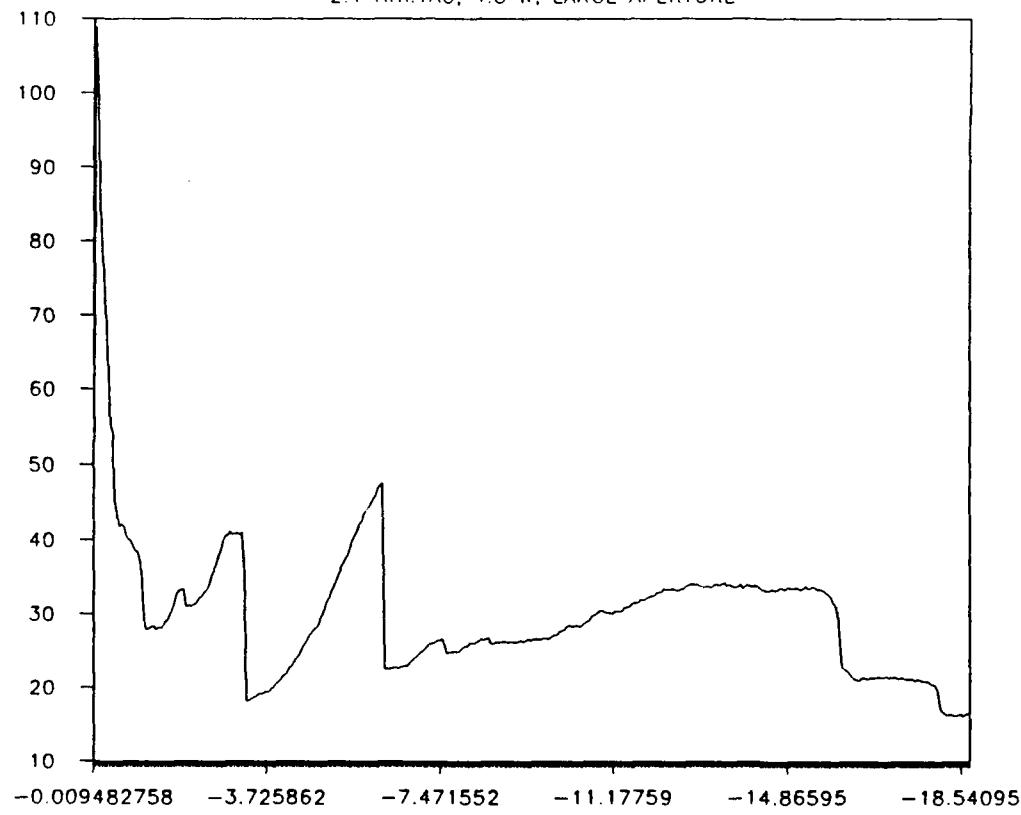
DEC21G-2X ICG CONTROLLED AT 80, 10s

2.1 Hm:YAG, 4.8 W, LARGE APERTURE



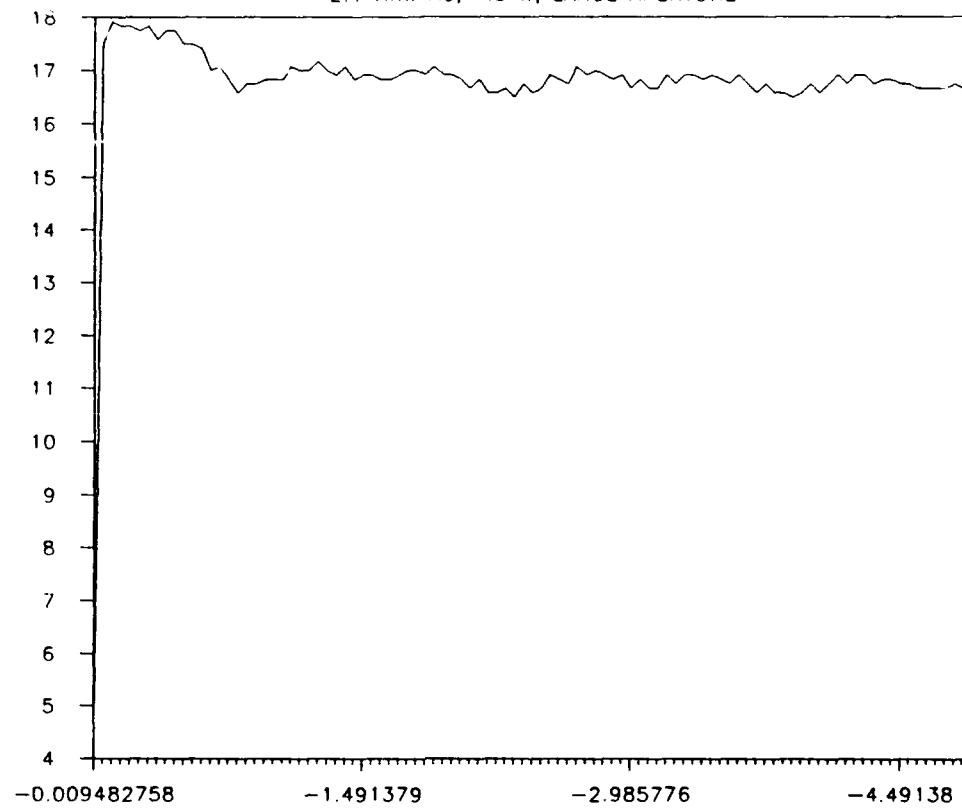
DEC21H-2X ICC CONTROLLED AT 100, 10s

2.1 Hm:YAG, 4.8 W, LARGE APERTURE



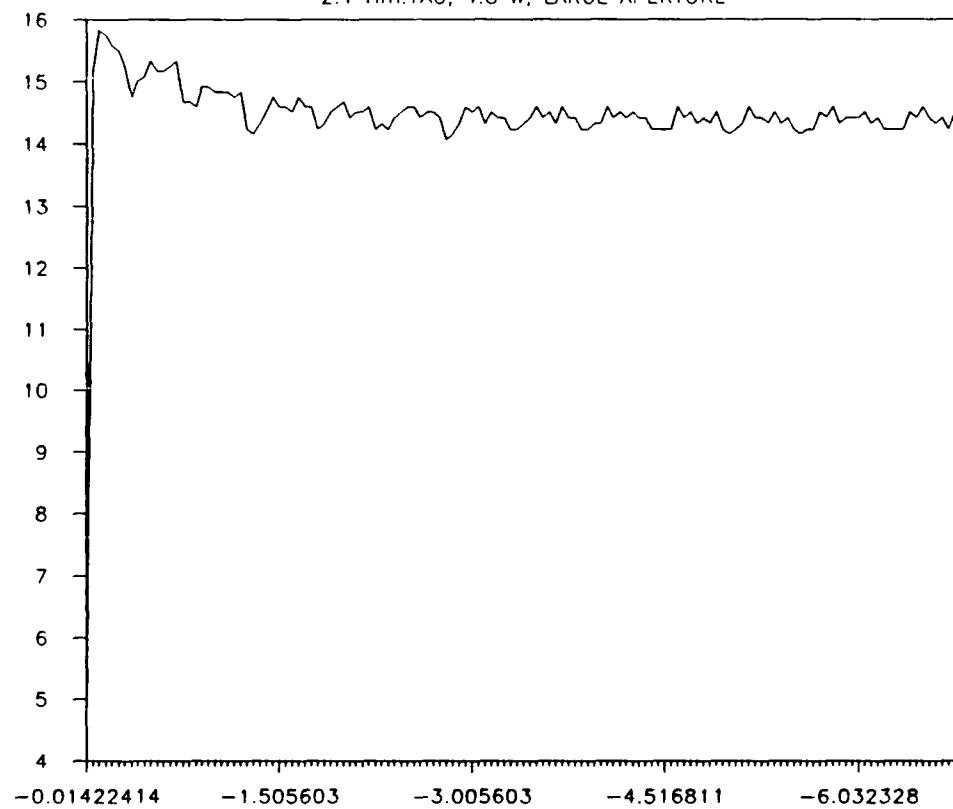
DEC21H-BLOOD UNWELDED CONTROL

2.1 Hm:YAG, 4.8 W, LARGE APERTURE



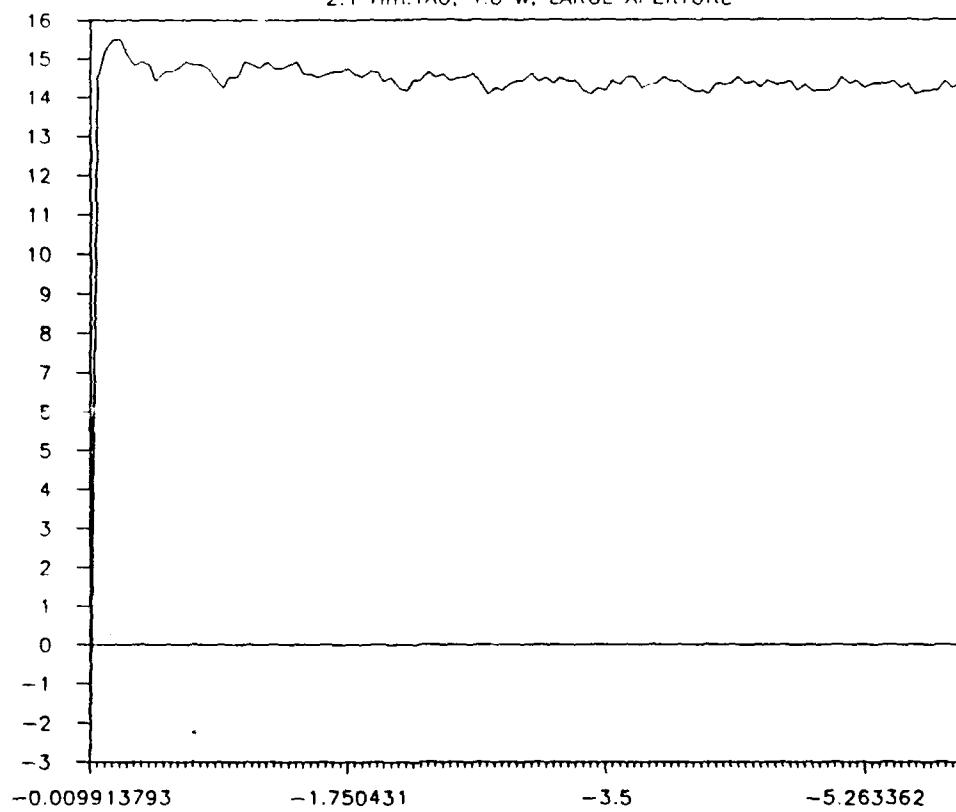
DEC21J-BLOOD CONTROLLED AT 50, 10s

2.1 Hm:YAG, 4.8 W, LARGE APERTURE



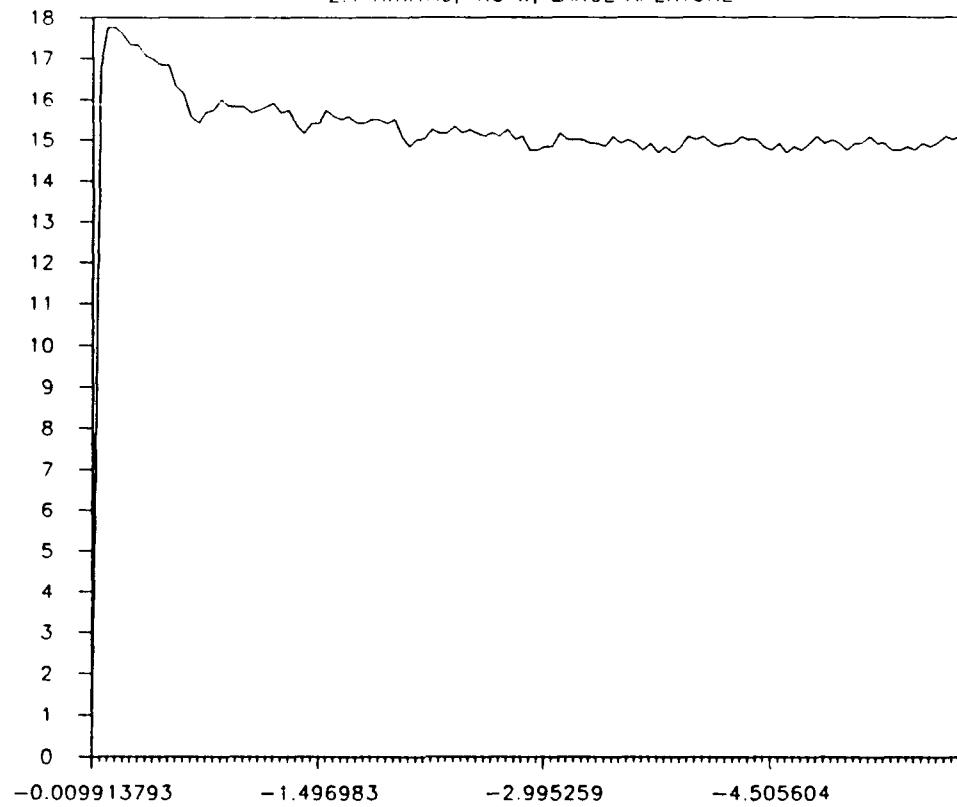
DEC21K-BLOOD CONTROLLED AT 60, 10s

2.1 Hm:YAG, 4.8 W, LARGE APERTURE



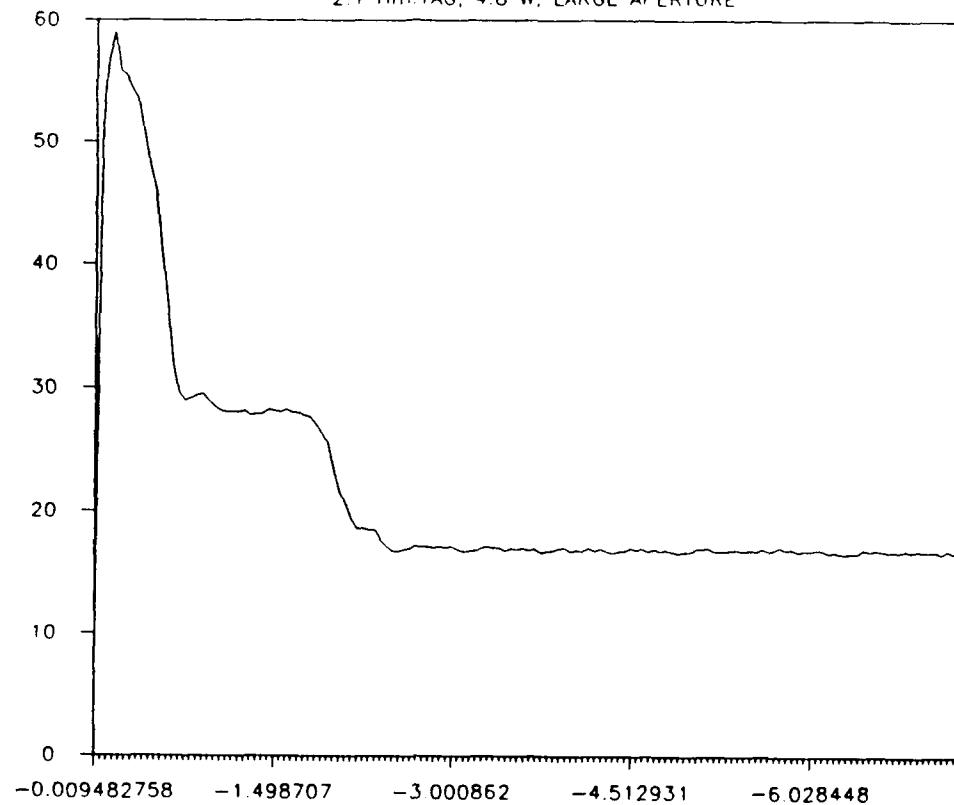
DEC21L - BLOOD CONTROLLED AT 70, 10s

2.1 Hm:YAC, 4.8 W, LARGE APERTURE



DEC21M- BLOOD CONTROLLED AT 80, 10s

2.1 Hm:YAG, 4.8 W, LARGE APERTURE



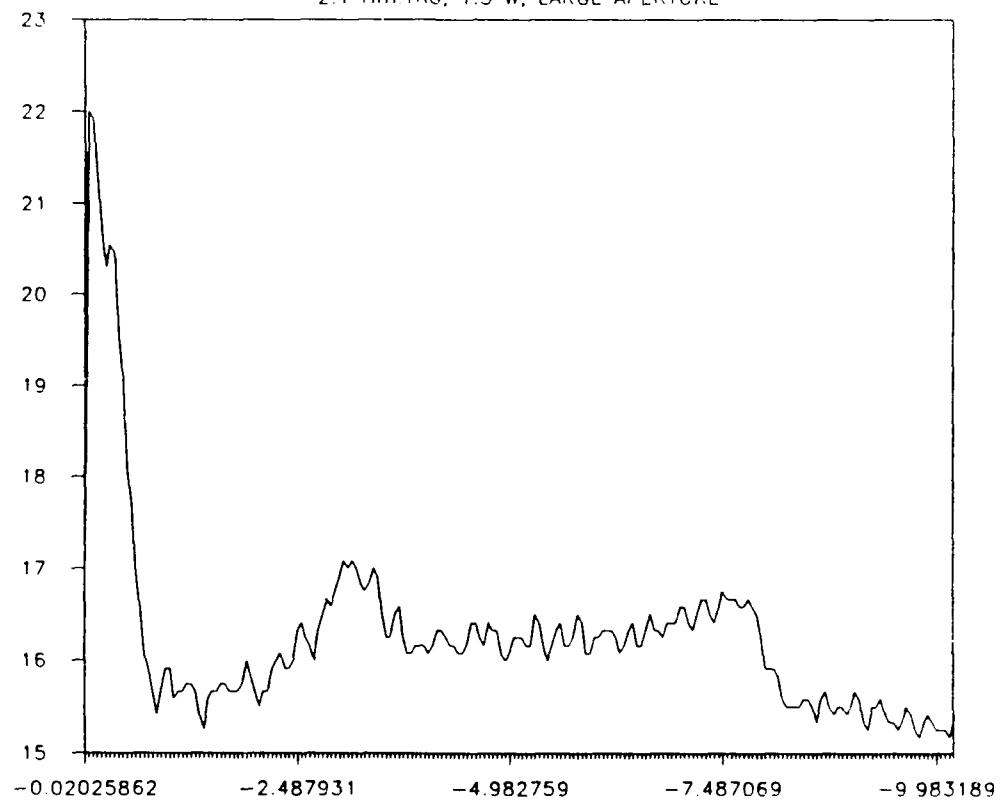
DEC21N-BLOOD CONTROLLED AT 100, 10s

2.1 Hm·YAG, 4.8 W, LARGE APERTURE



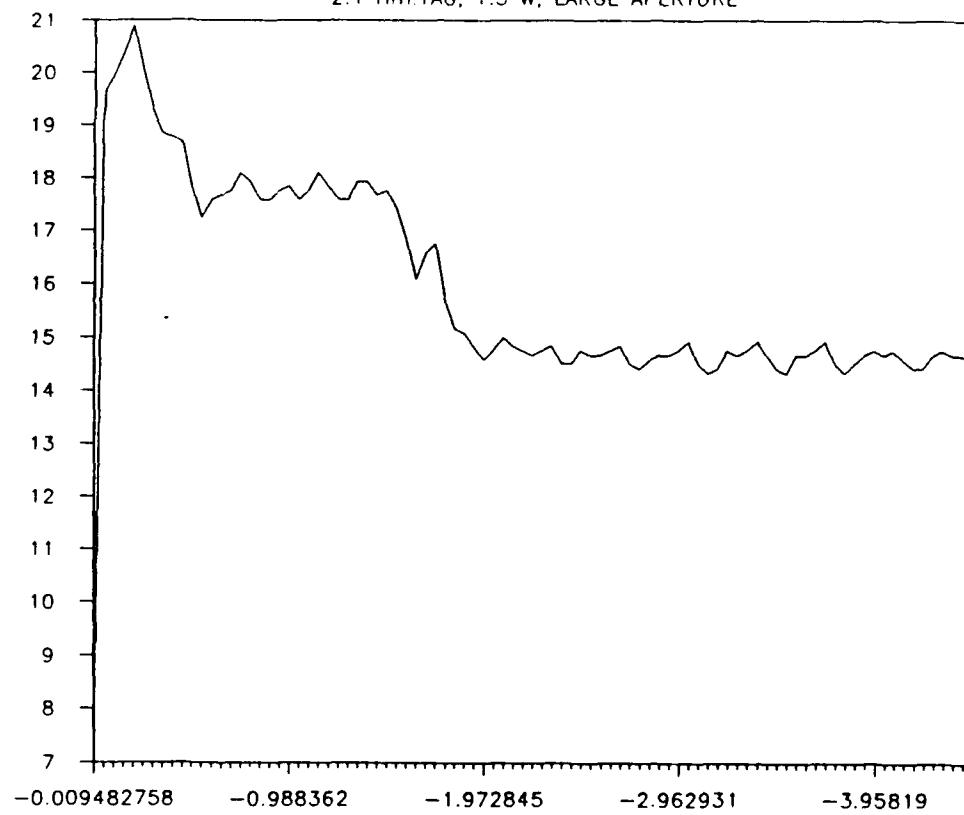
DEC2101-BLOOD CONTROLLED AT 50, 10s

2.1 Hm:YAG, 1.5 W, LARGE APERTURE



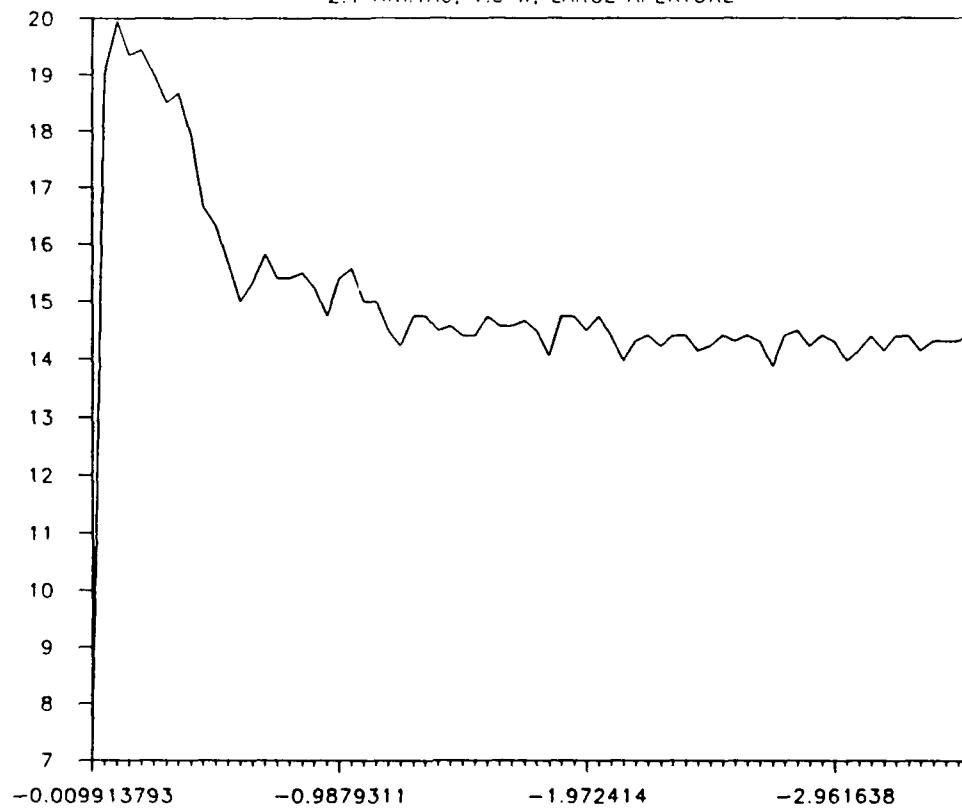
DEC21P-BLOOD CONTROLLED AT 60, 10s

2.1 Hm:YAG, 1.5 W, LARGE APERTURE



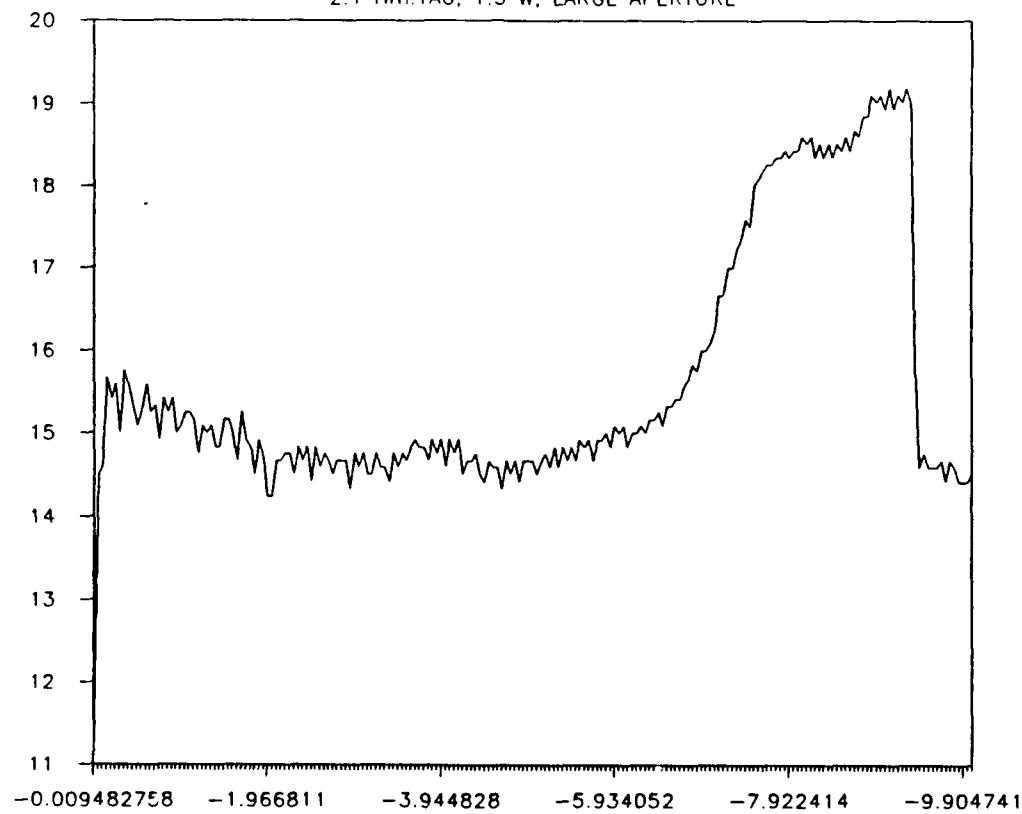
DEC21Q-BLOOD CONTROLLED AT 70, 10s

2.1 Hm:YAG, 1.5 W, LARGE APERTURE



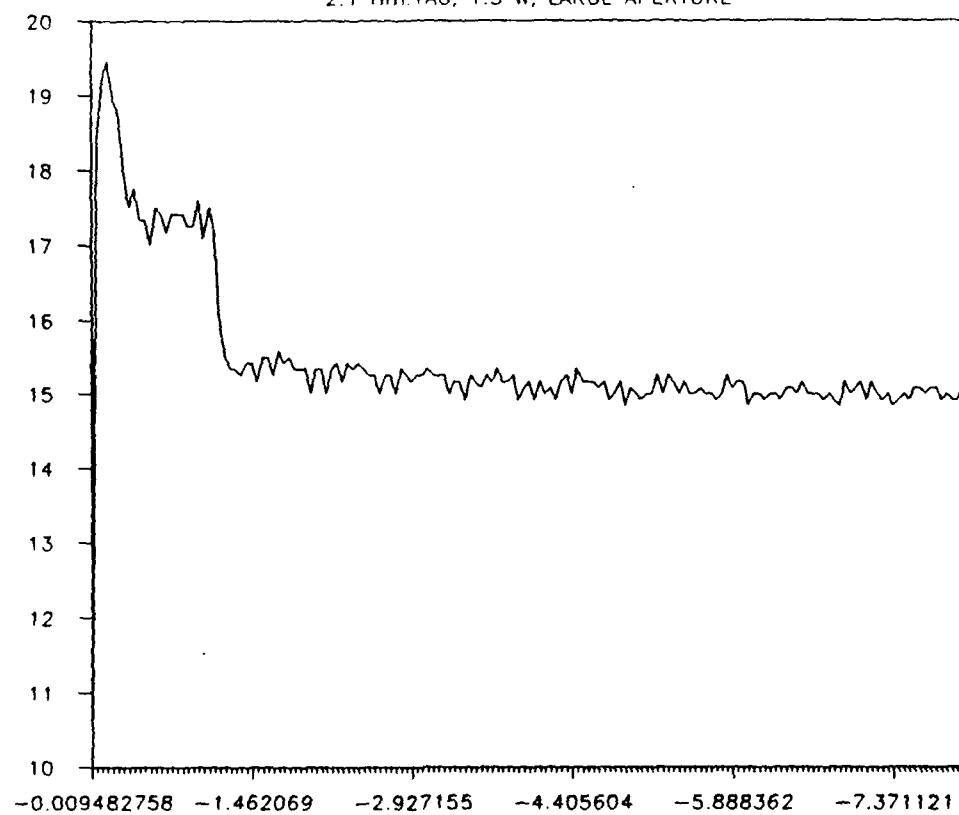
DEC21R-BLOOD CONTROLLED AT 80, 10s

2.1 Hm:YAG, 1.5 W, LARGE APERTURE



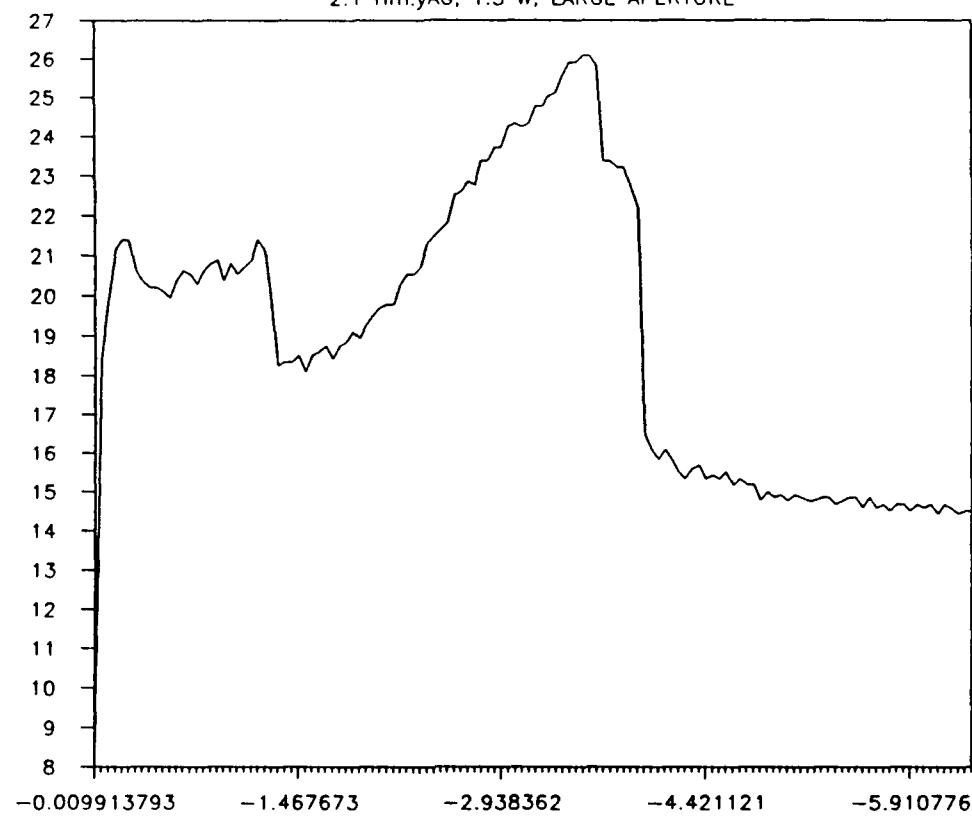
DEC21T-2X ICG CONTROLLED AT 50, 10s

2.1 Hm:YAG, 1.5 W, LARGE APERTURE



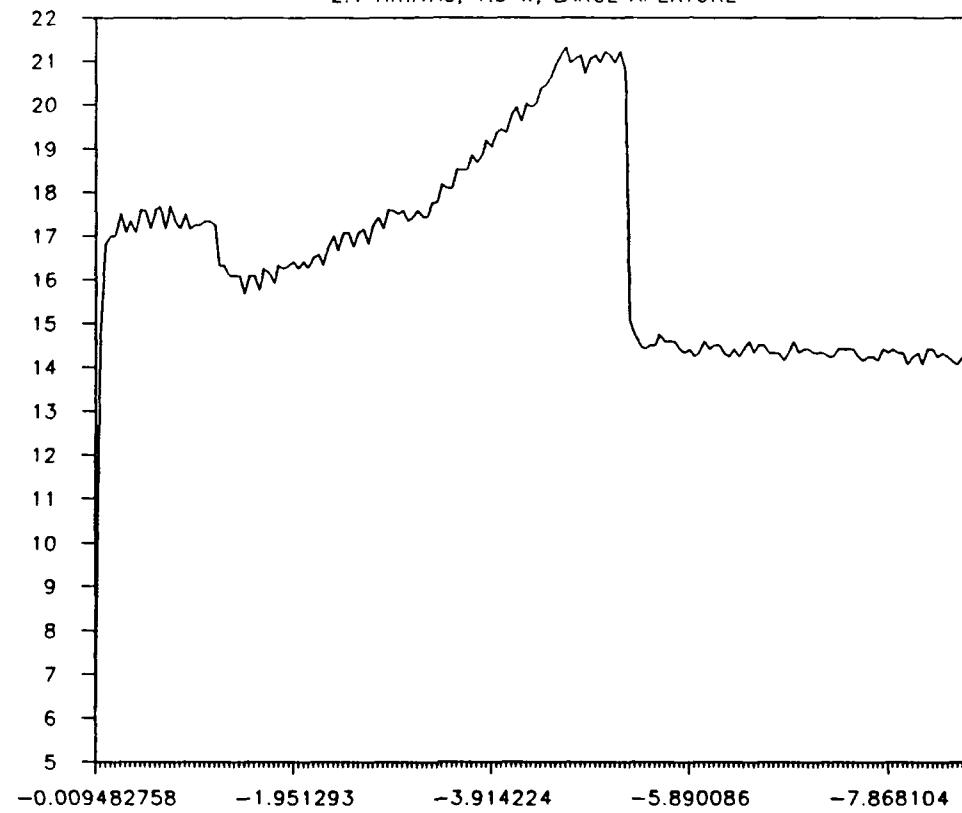
DEC21S-BLOOD CONTROLLED AT 100, 10s

2.1 Hm:yAG, 1.5 W, LARGE APERTURE



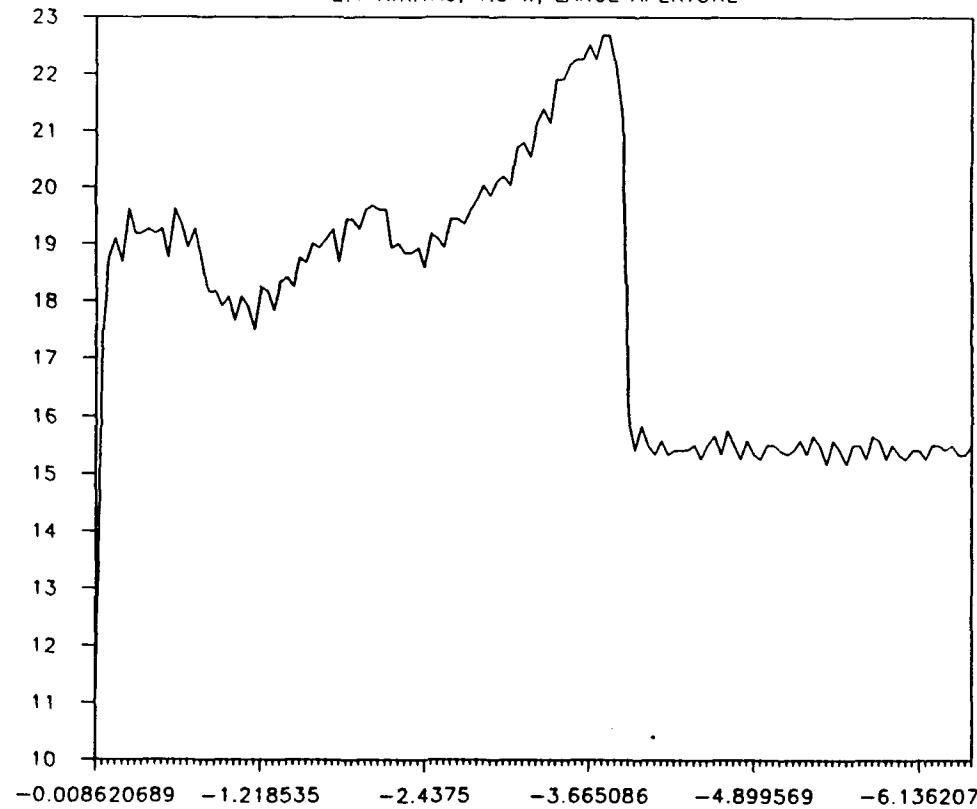
DEC21U-2X ICG CONTROLLED AT 60, 10s

2.1 Hm:YAG, 1.5 W, LARGE APERTURE



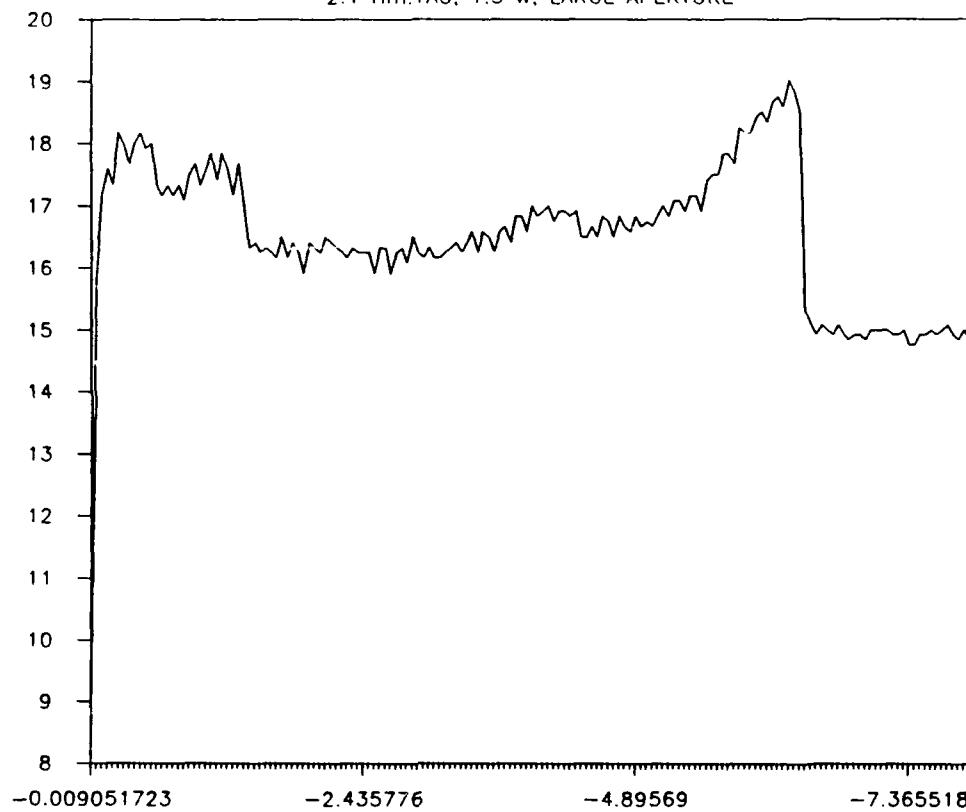
DEC21V-2X ICG CONTROLLED AT 70, 10s

2.1 Hm:YAG, 1.5 W, LARGE APERTURE



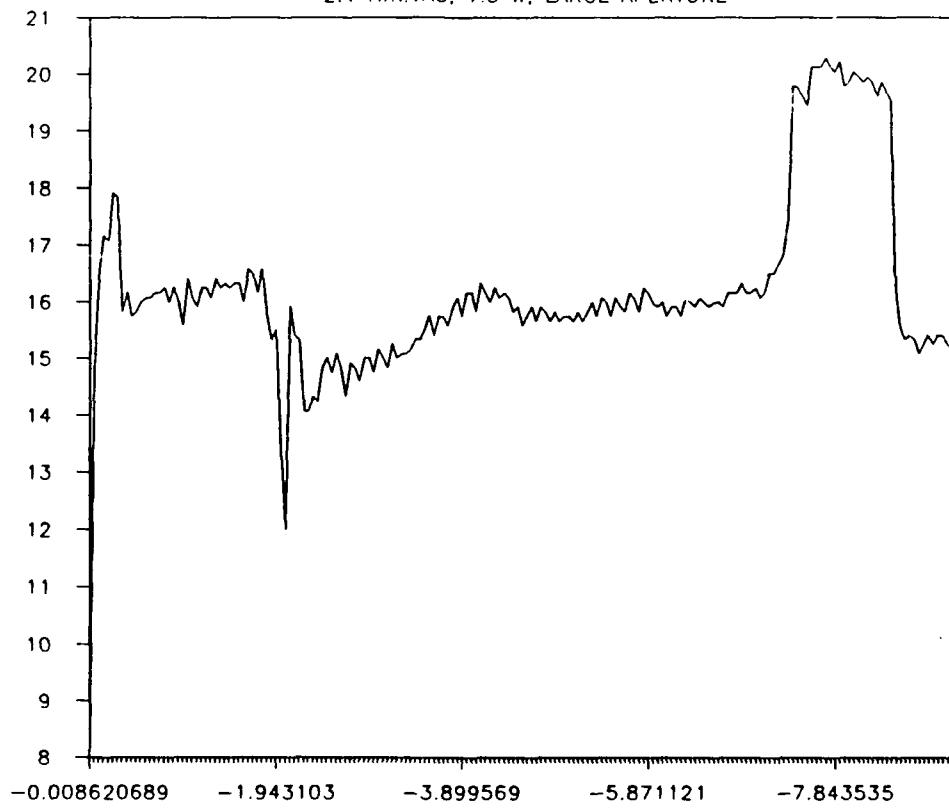
DEC21W-2X ICG CONTROLLED AT 80, 10s

2.1 Hm:YAG, 1.5 W, LARGE APERTURE



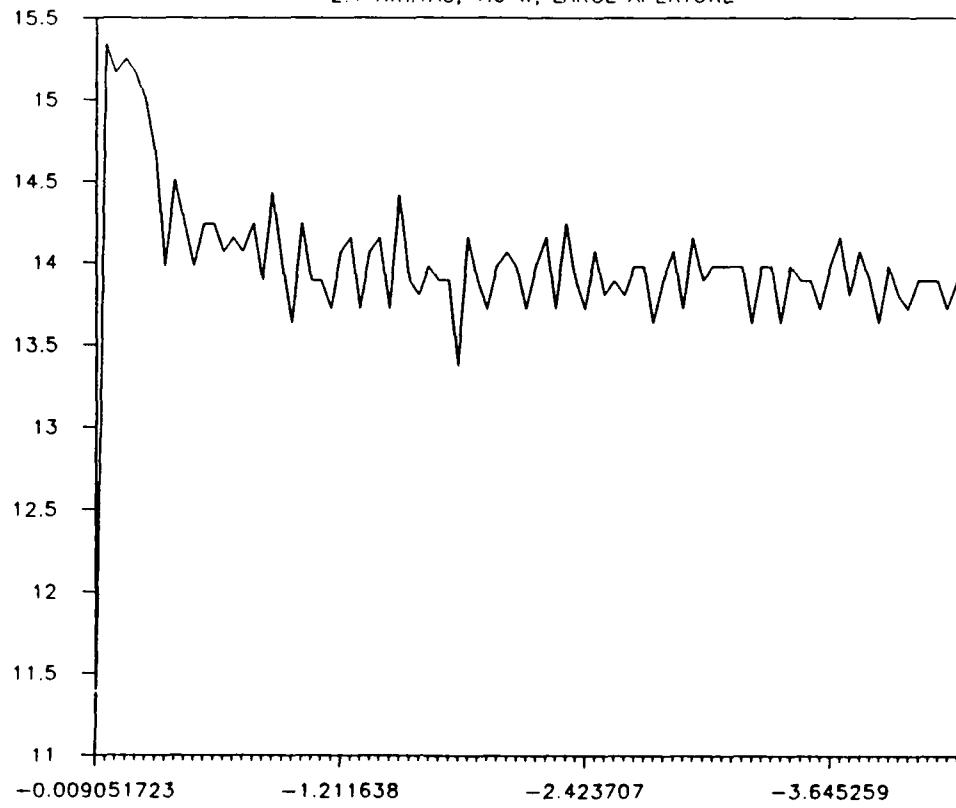
DEC21X-2X ICG CONTROLLED AT 100, 10s

2.1 Hm:YAG, 1.5 W, LARGE APERTURE

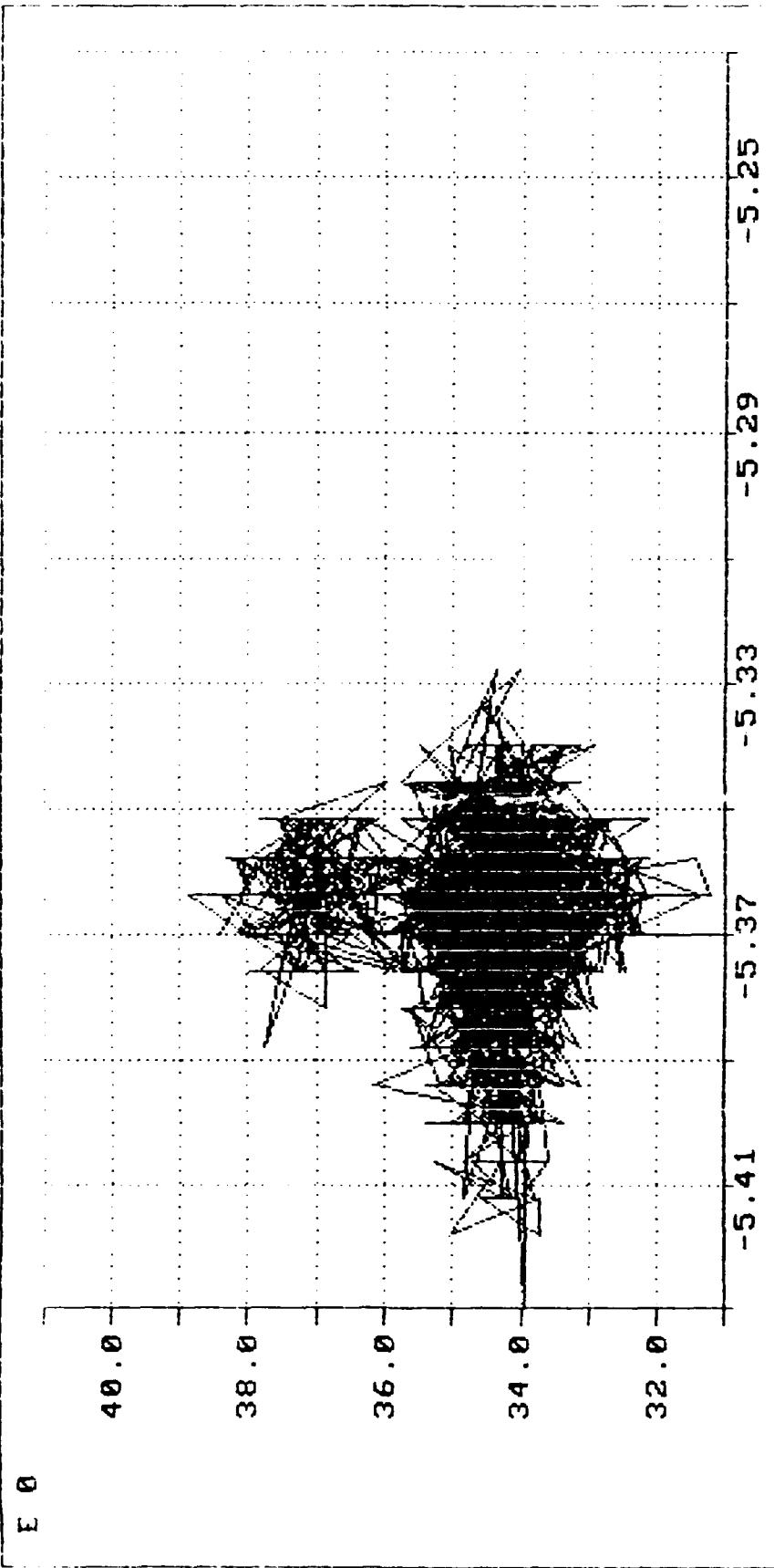


DEC21Y-NO CHROMO CONTROLLED AT 50, 10s

2.1 Hm:YAC, 1.5 W, LARGE APERTURE

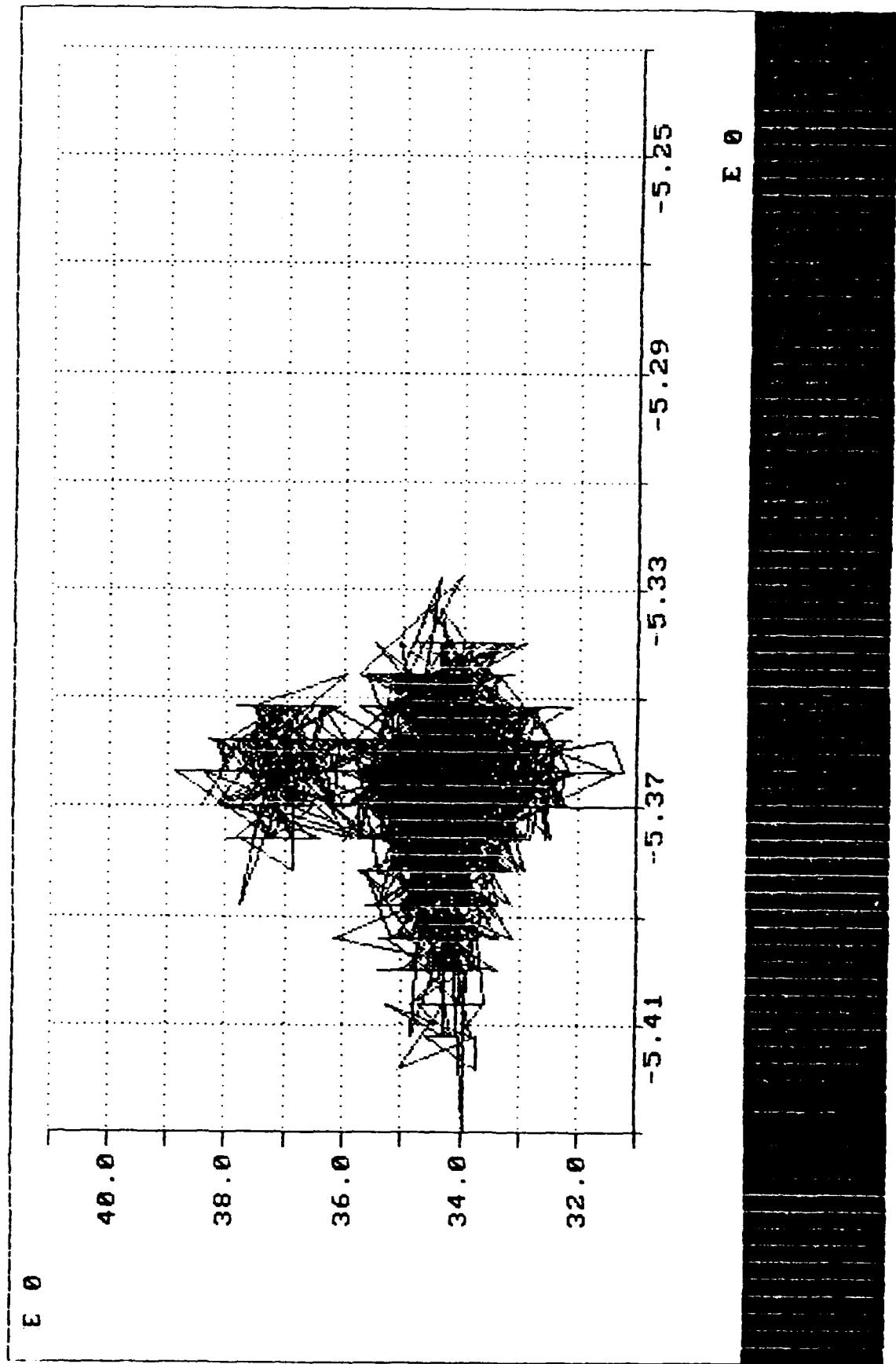


Appendix B- Excluded Data

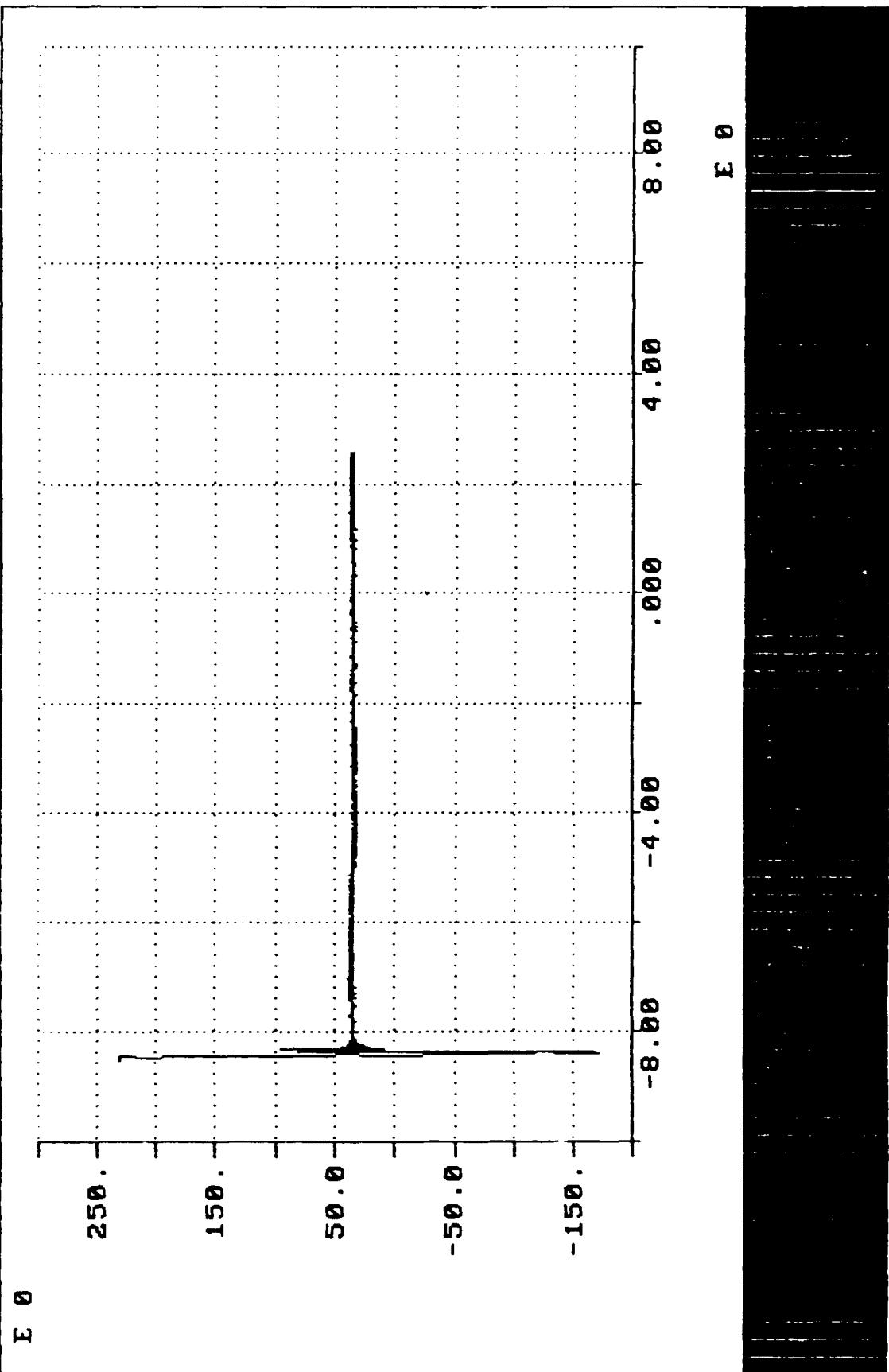


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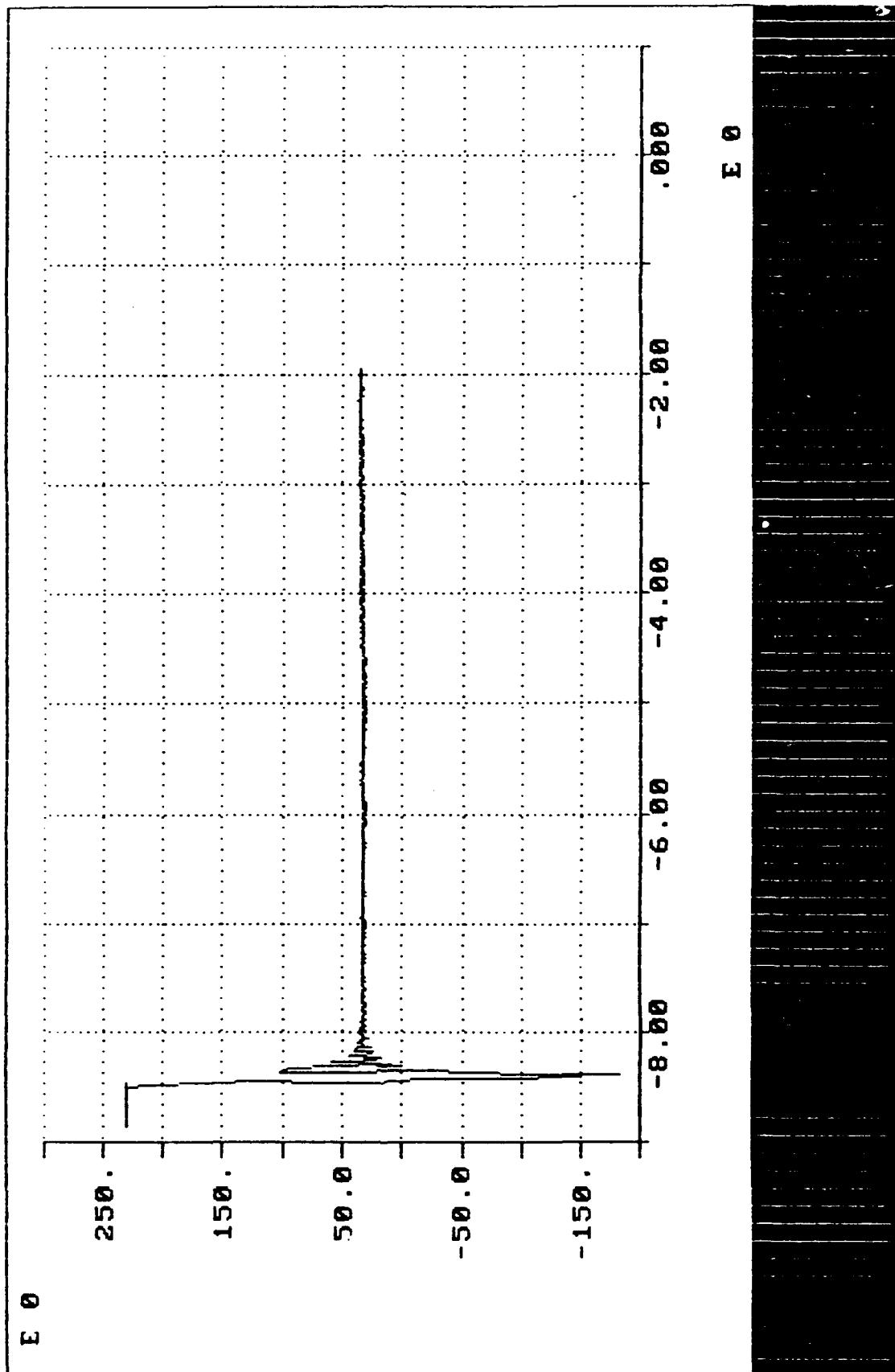




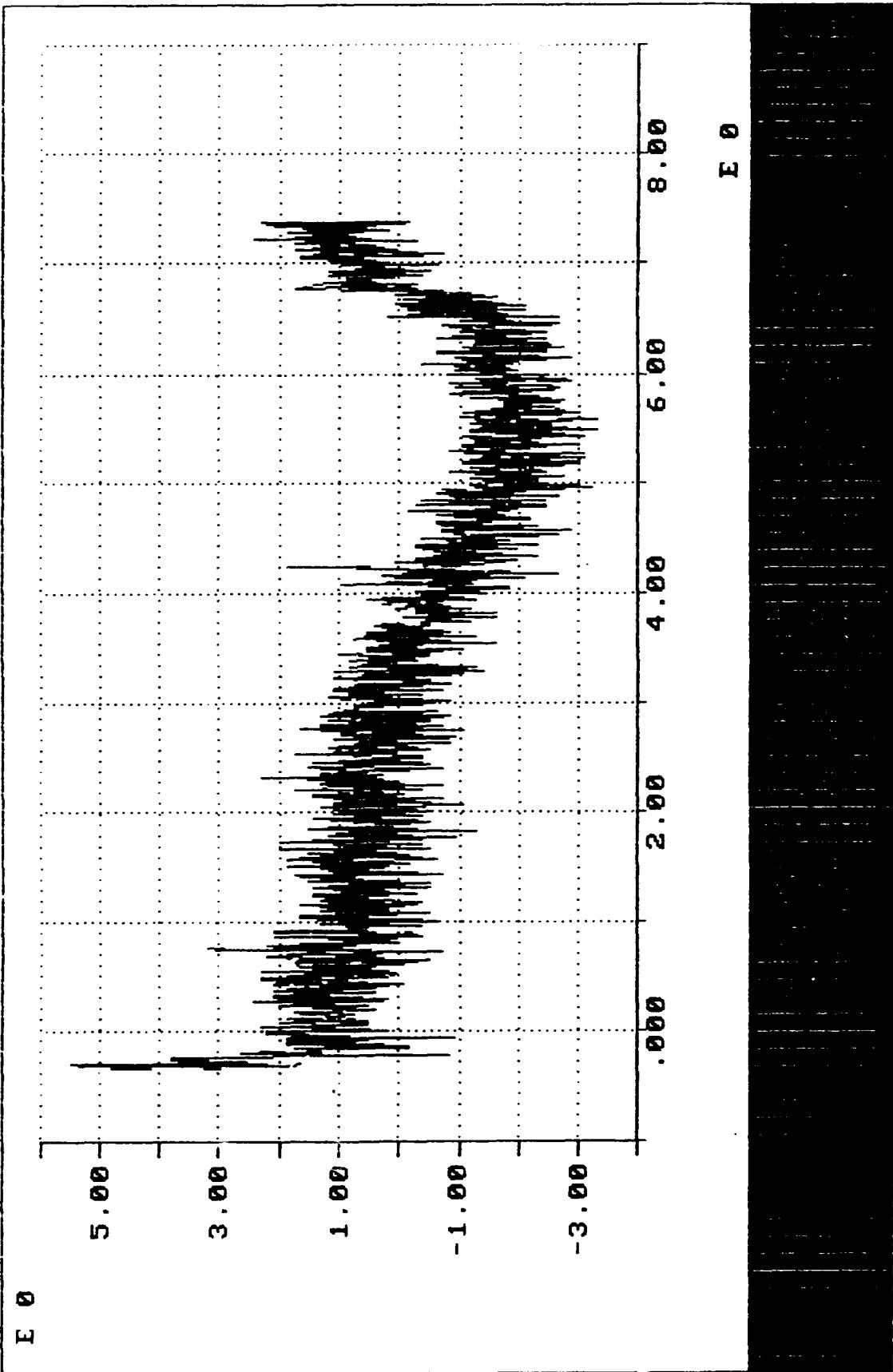
908 105



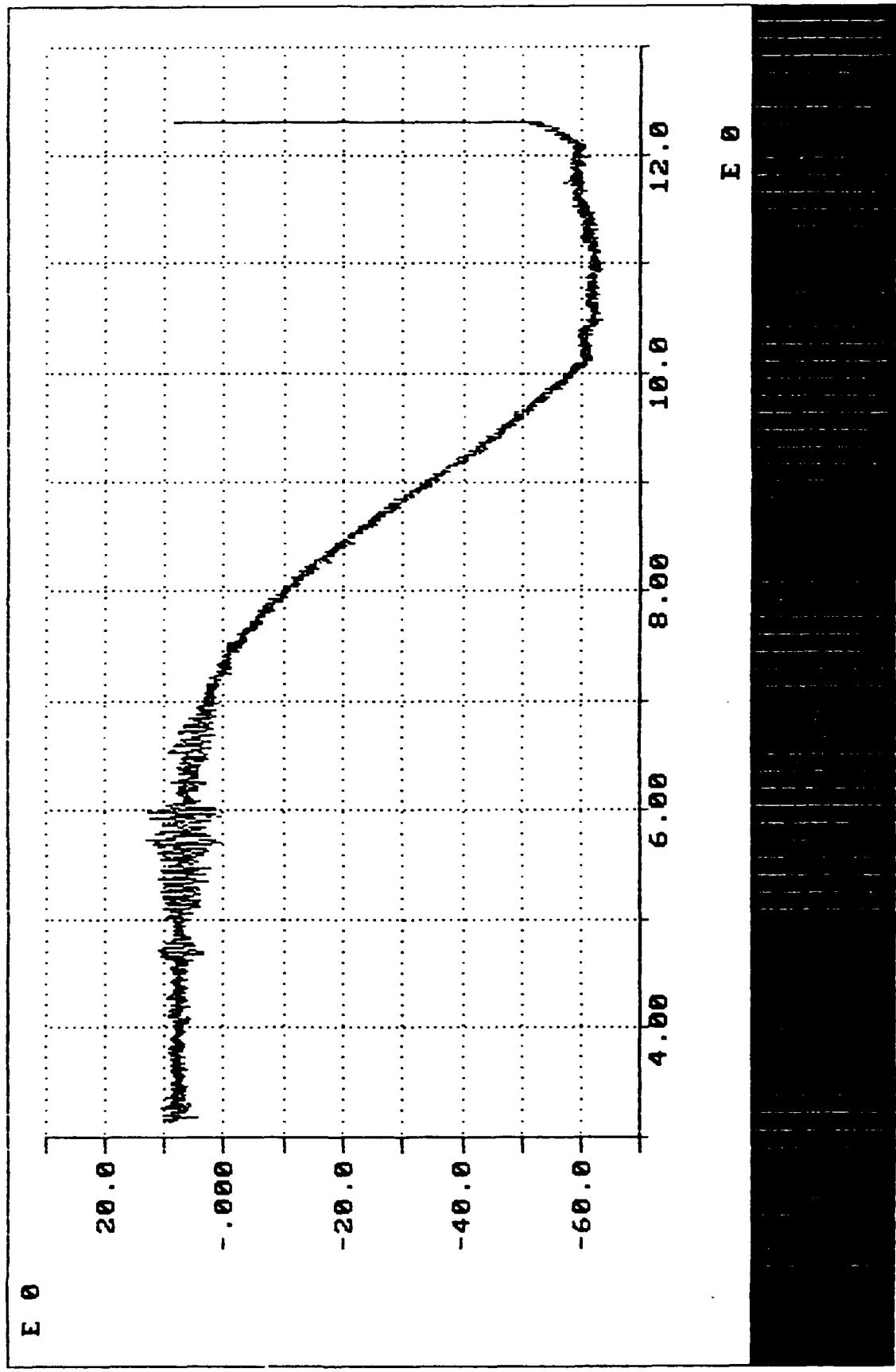
-35P 1075



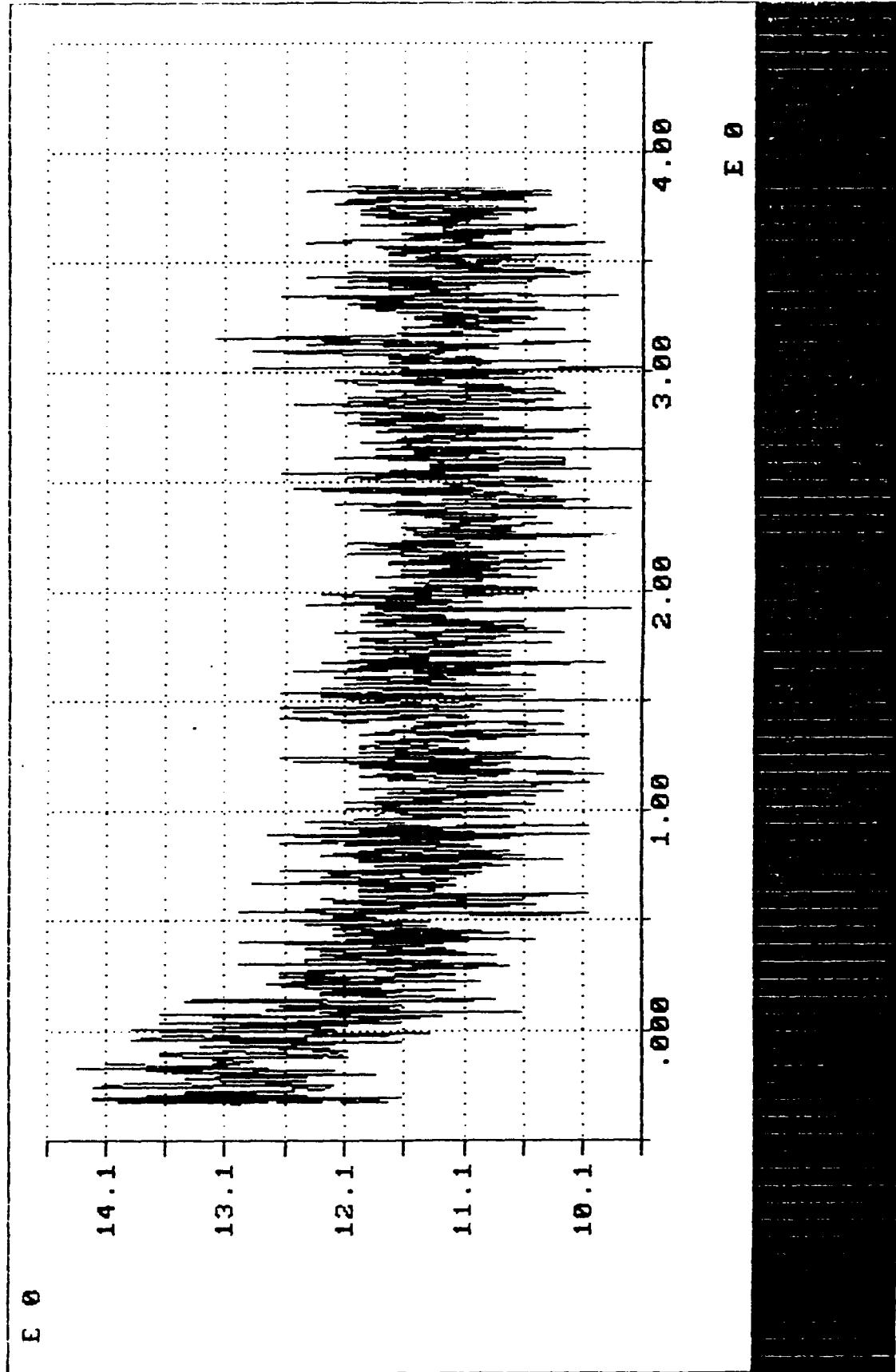
11/20/08 10:55



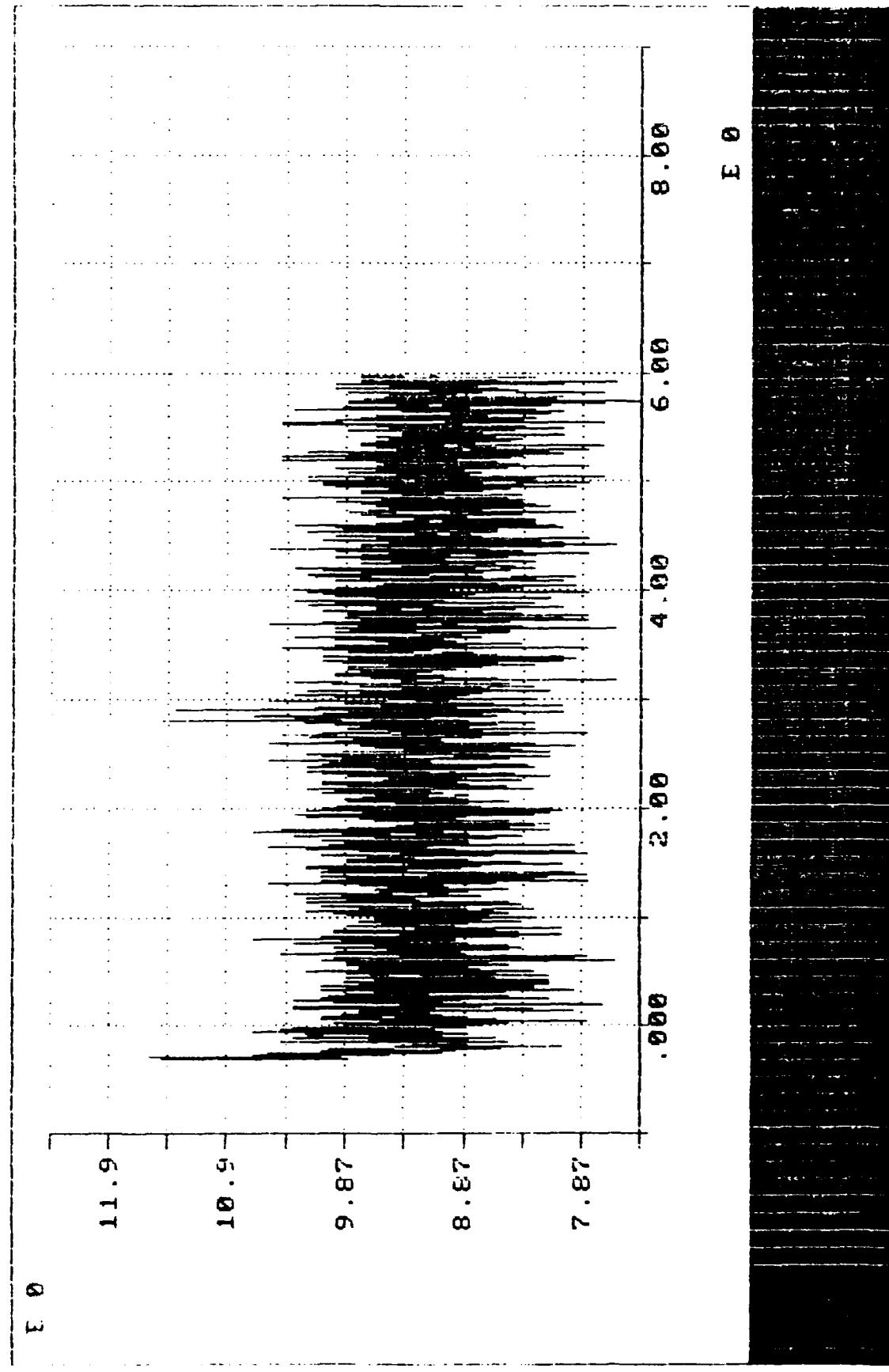
4 L 8 105



-0 1.0 105

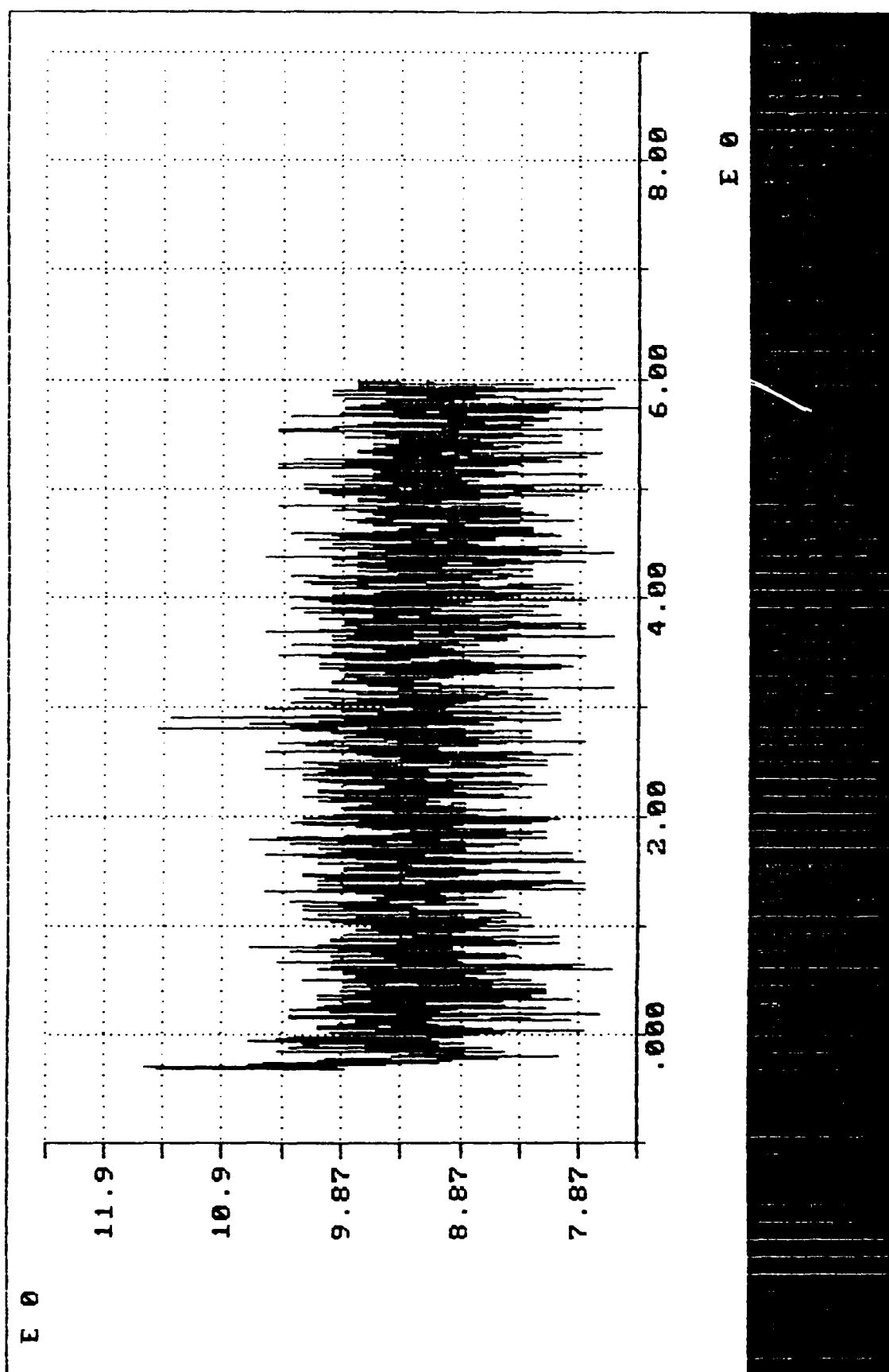


6 10 125

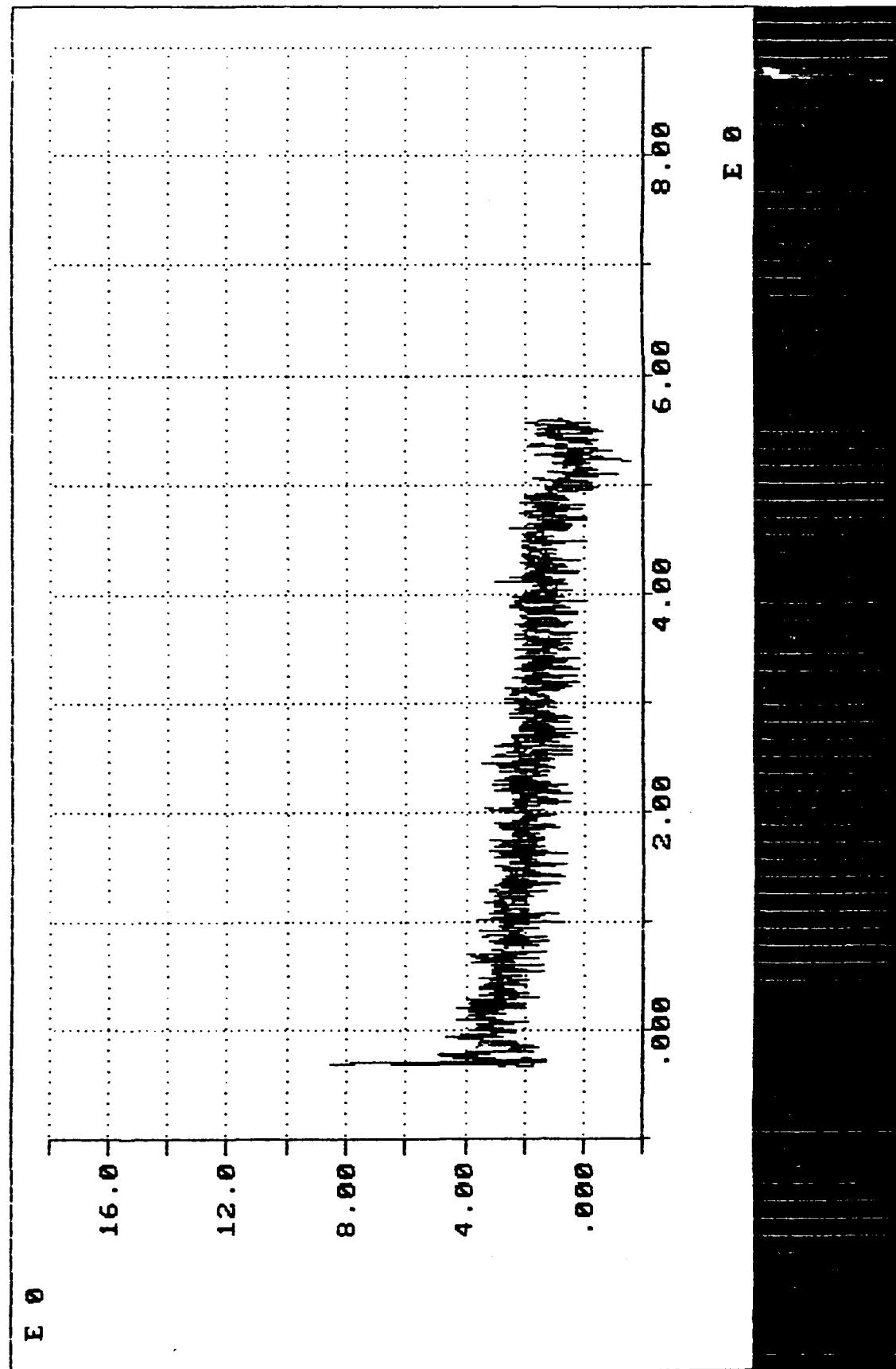


+ 18 105

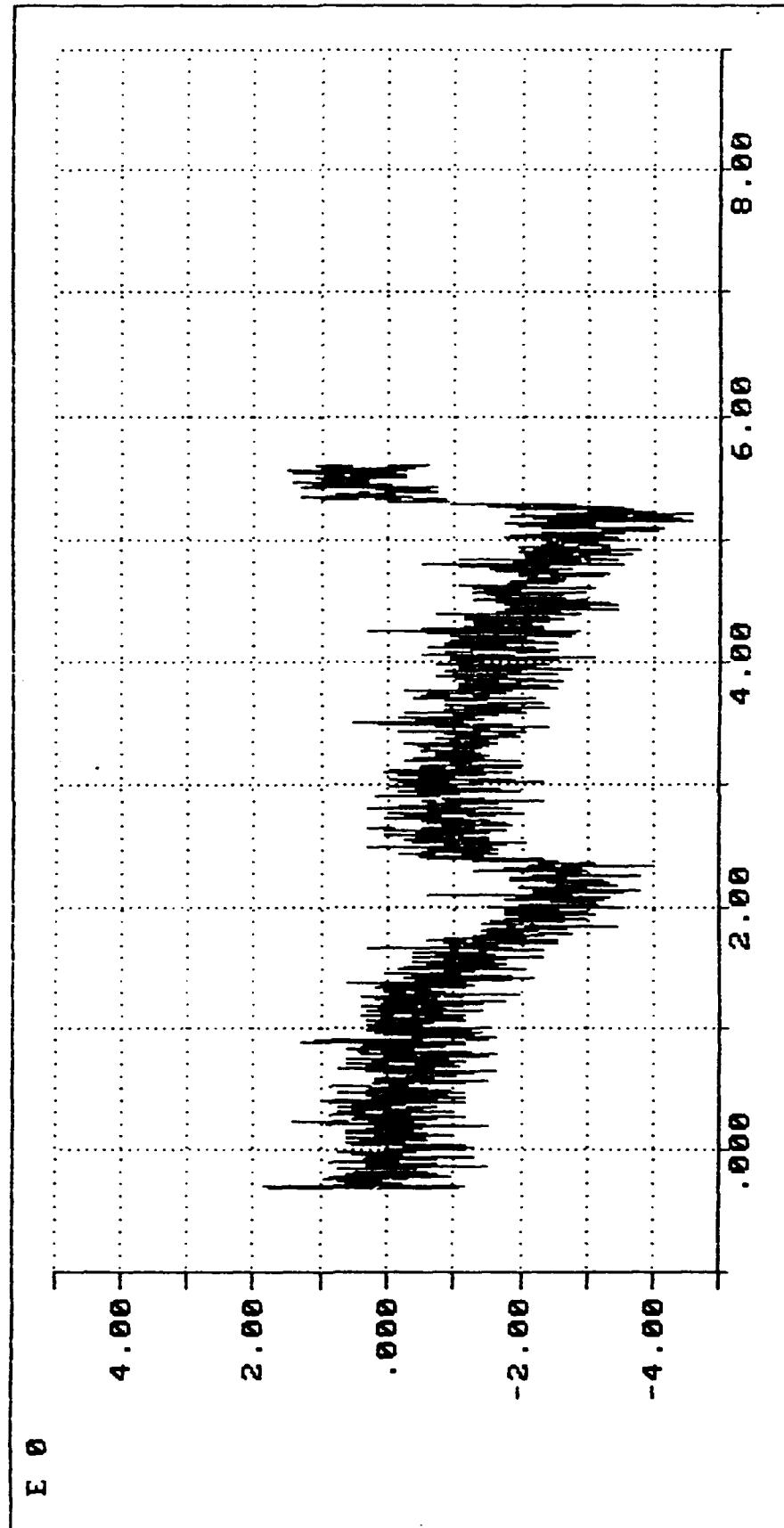
0 3



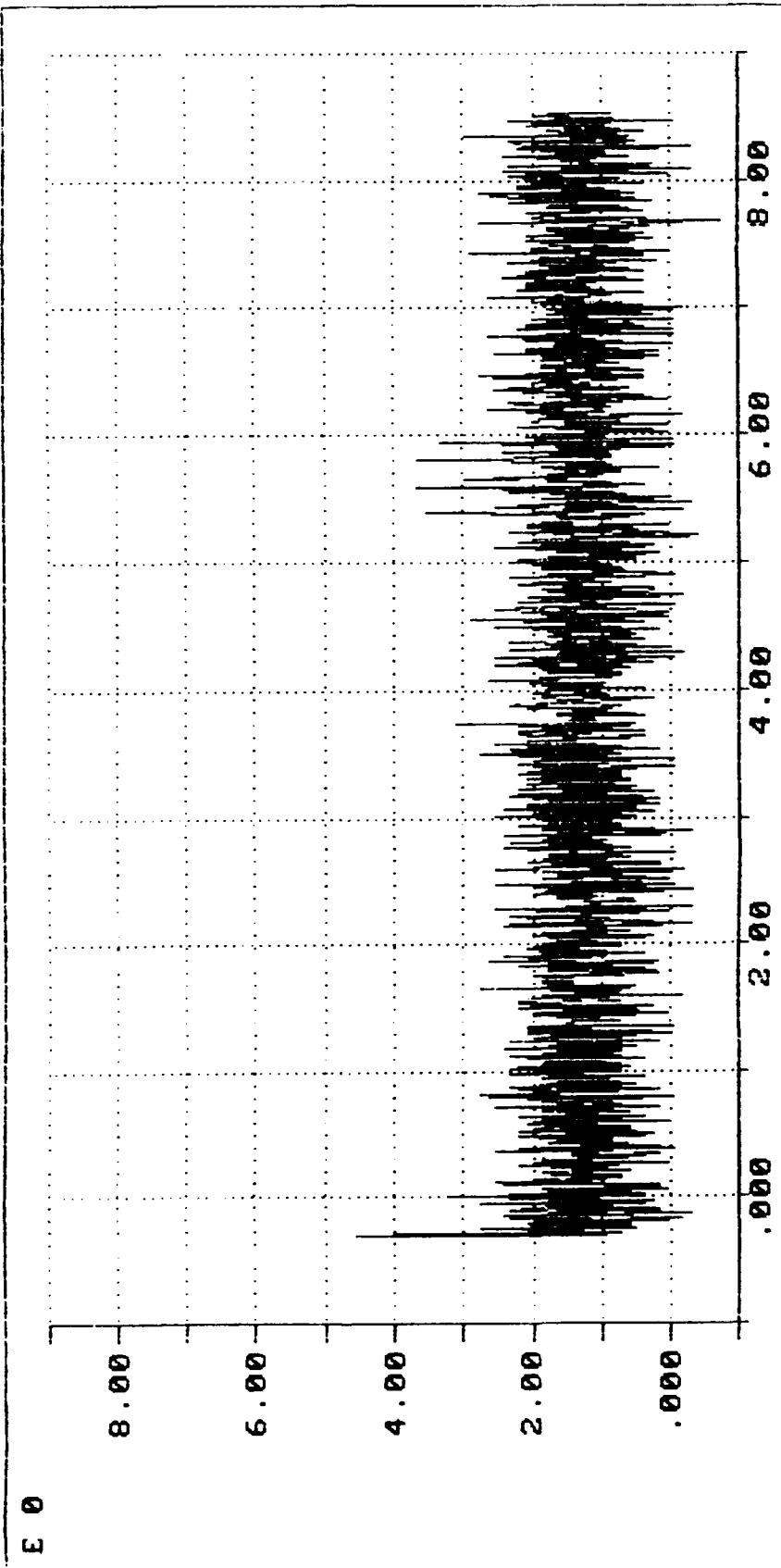
OLR 125



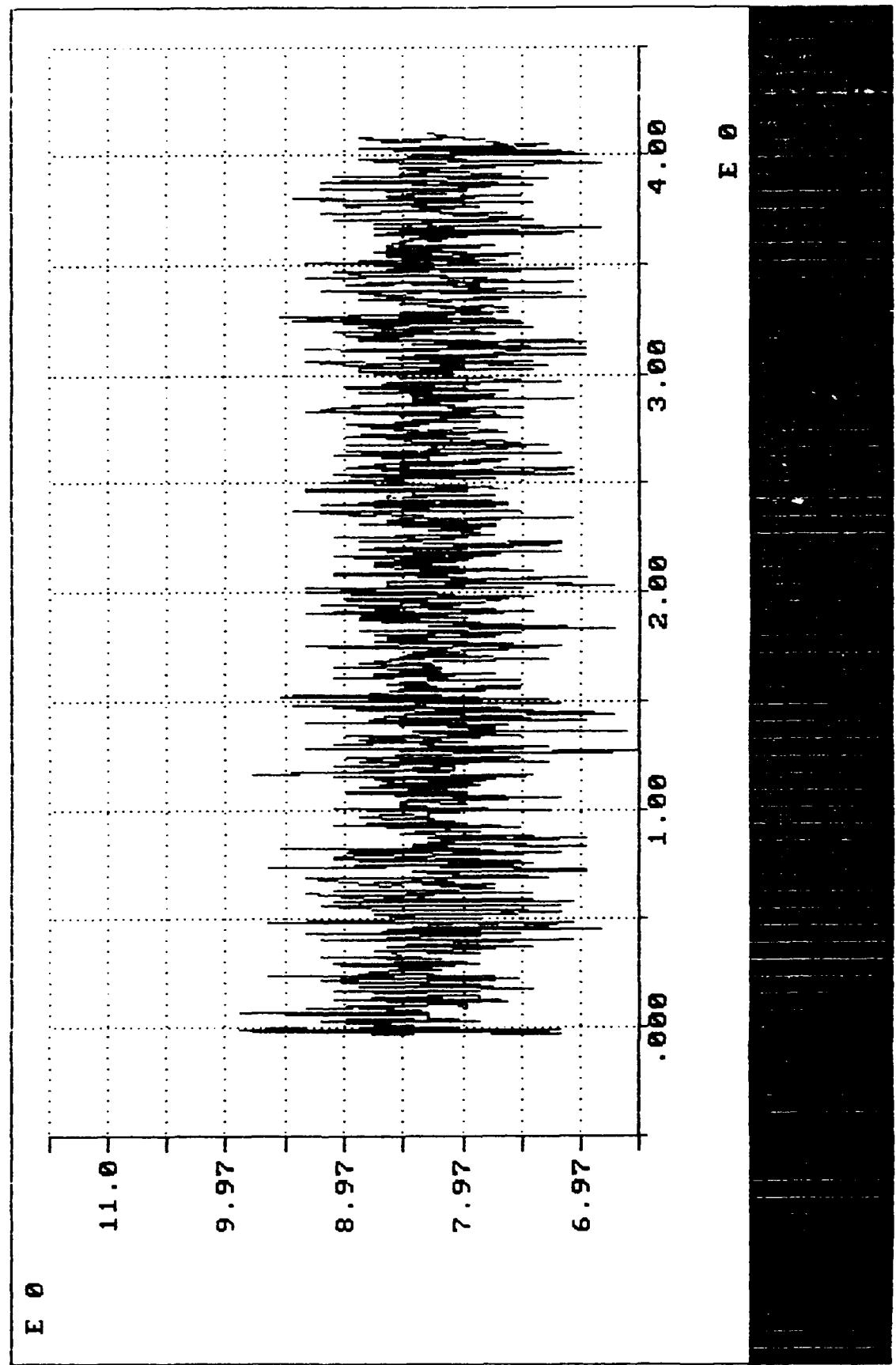
I LP 175



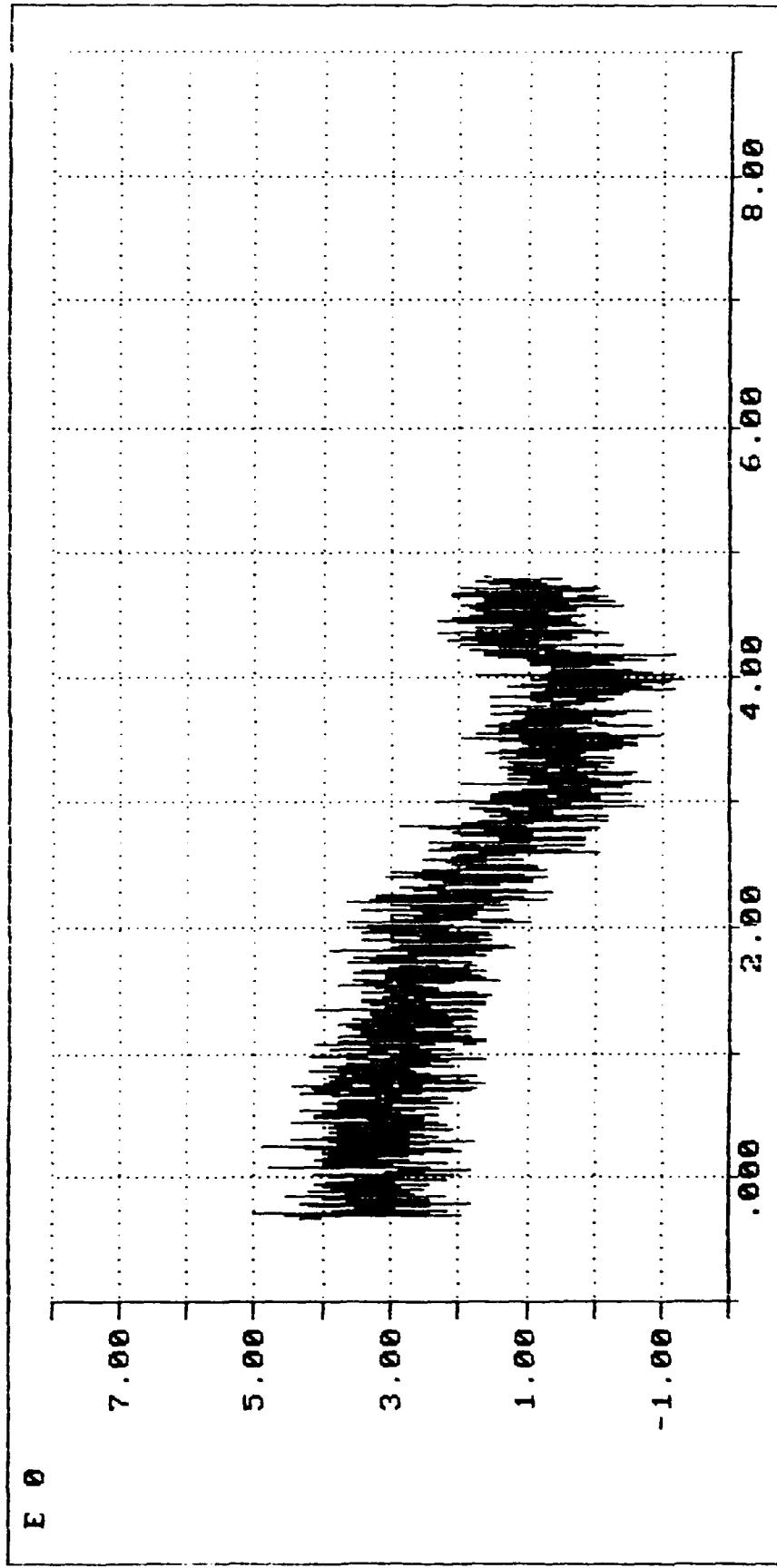
5 L 8 101



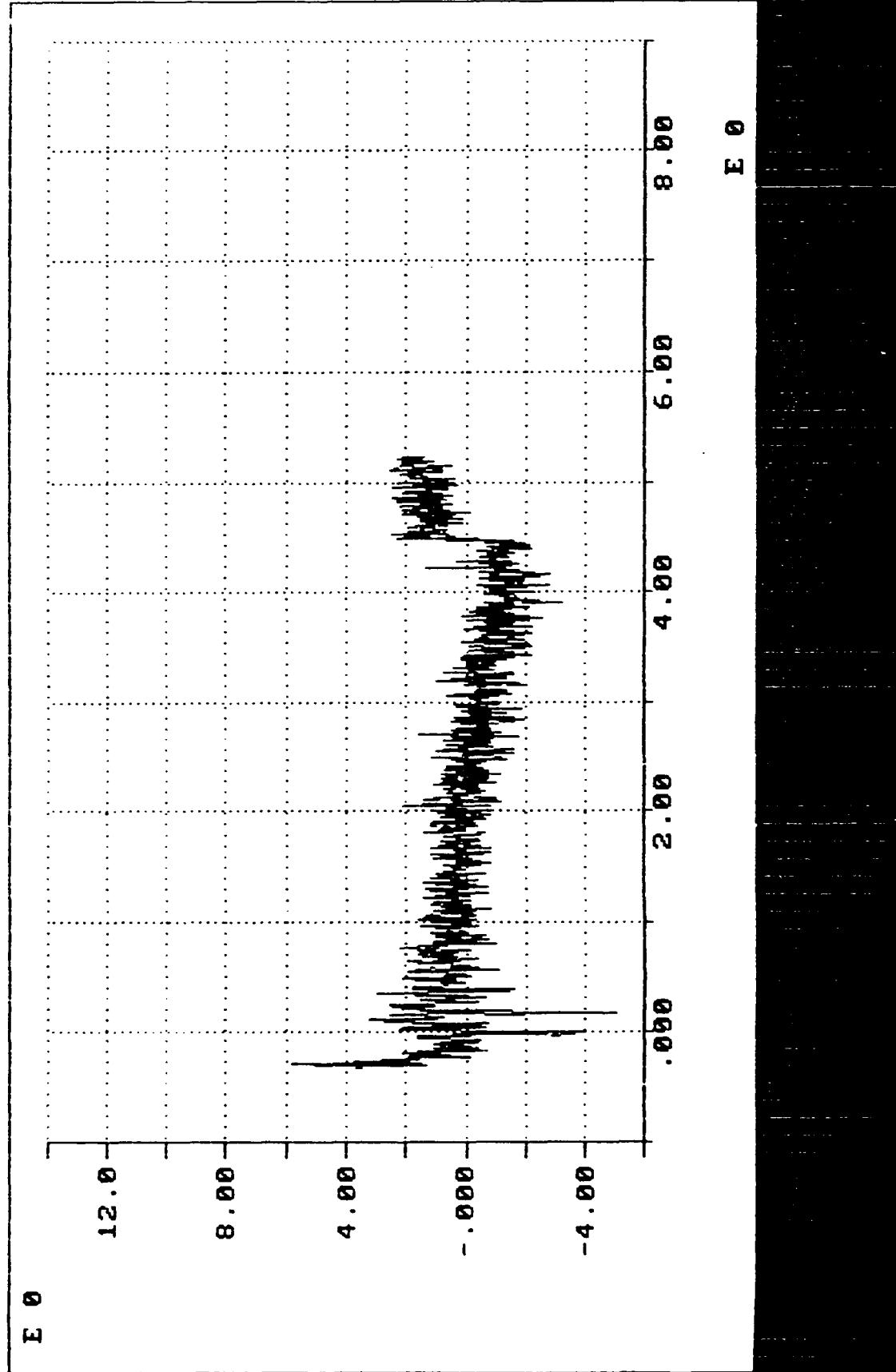
100-105



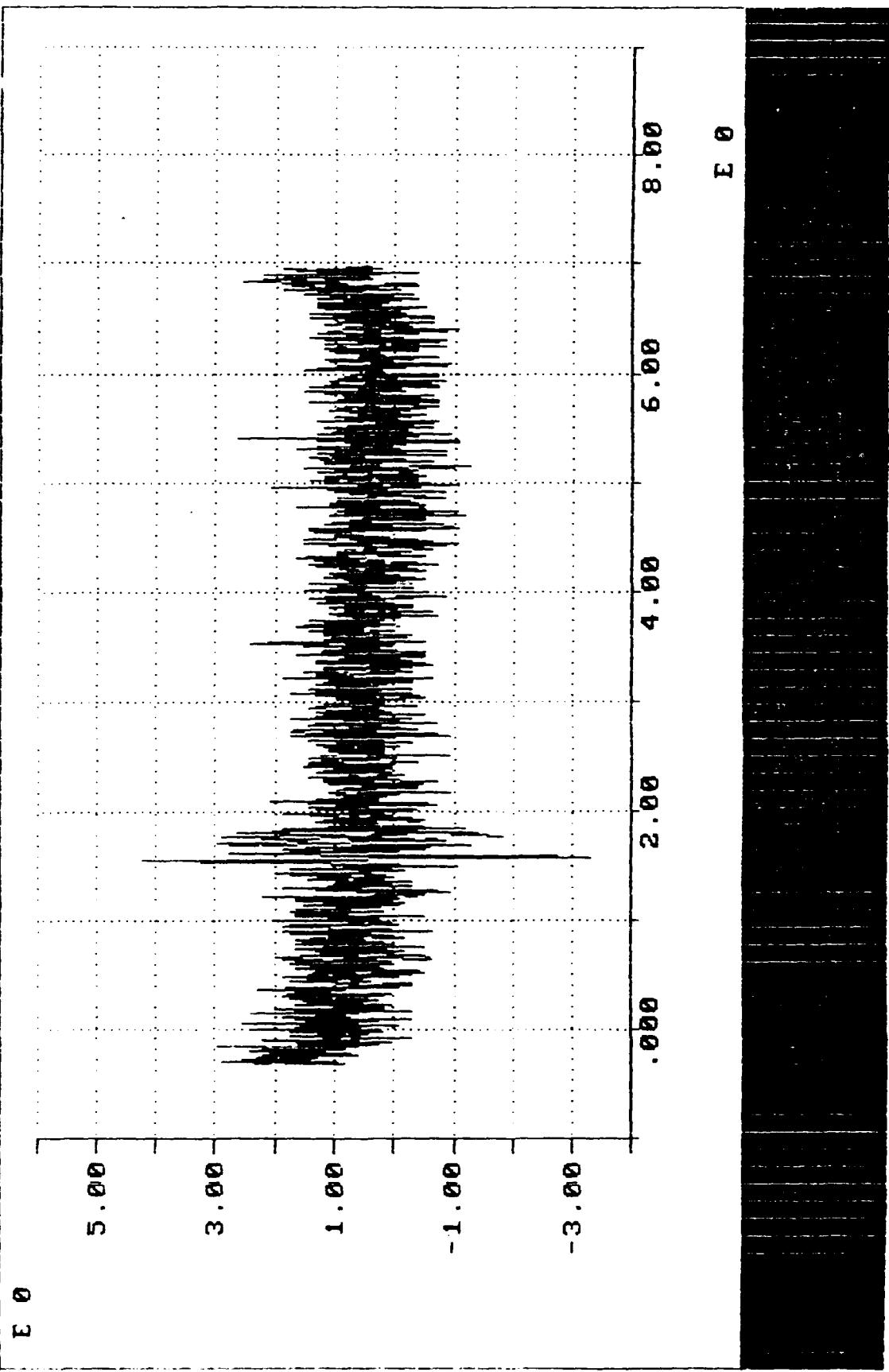
21 E 105



Y LE 175

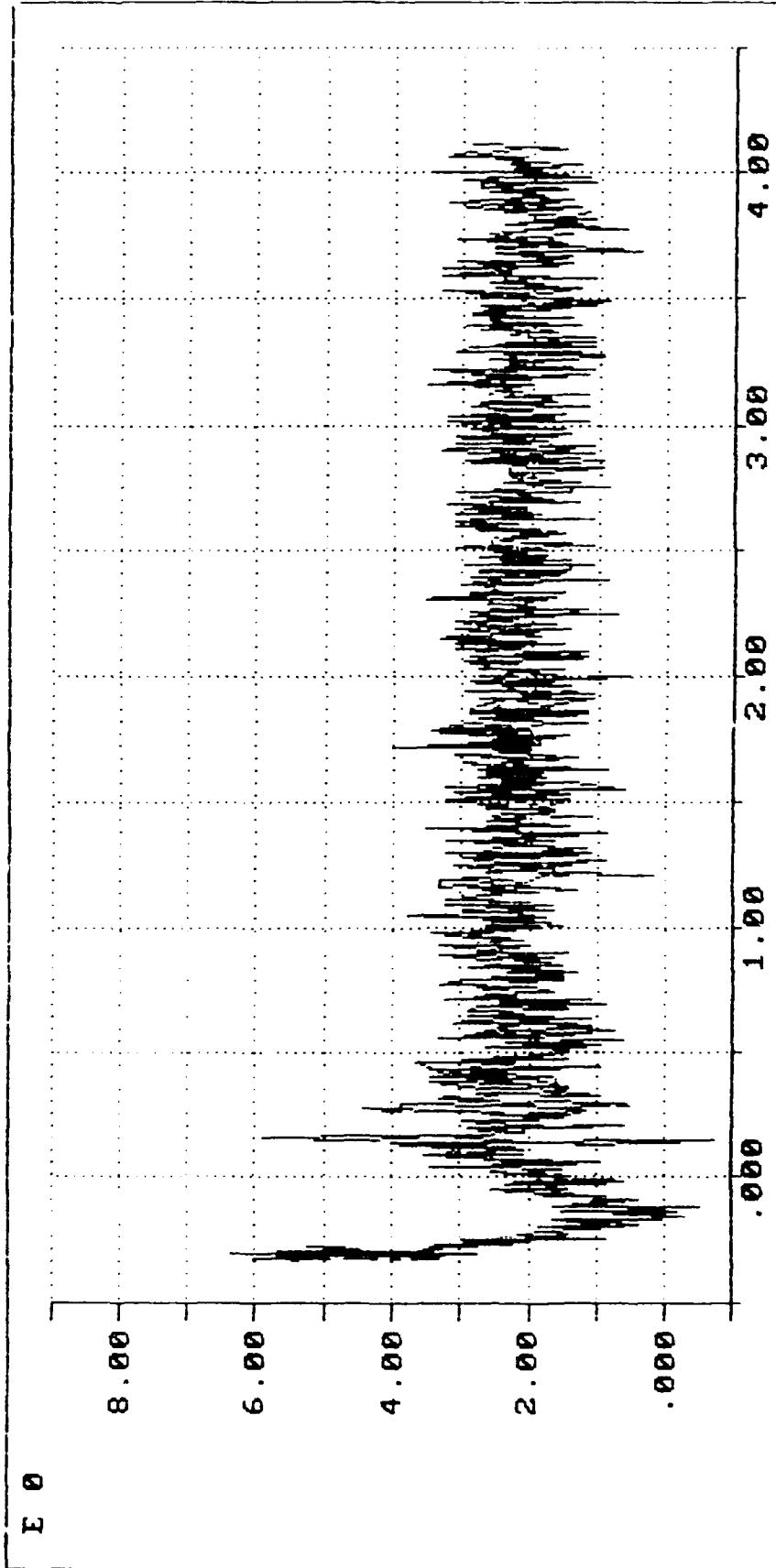


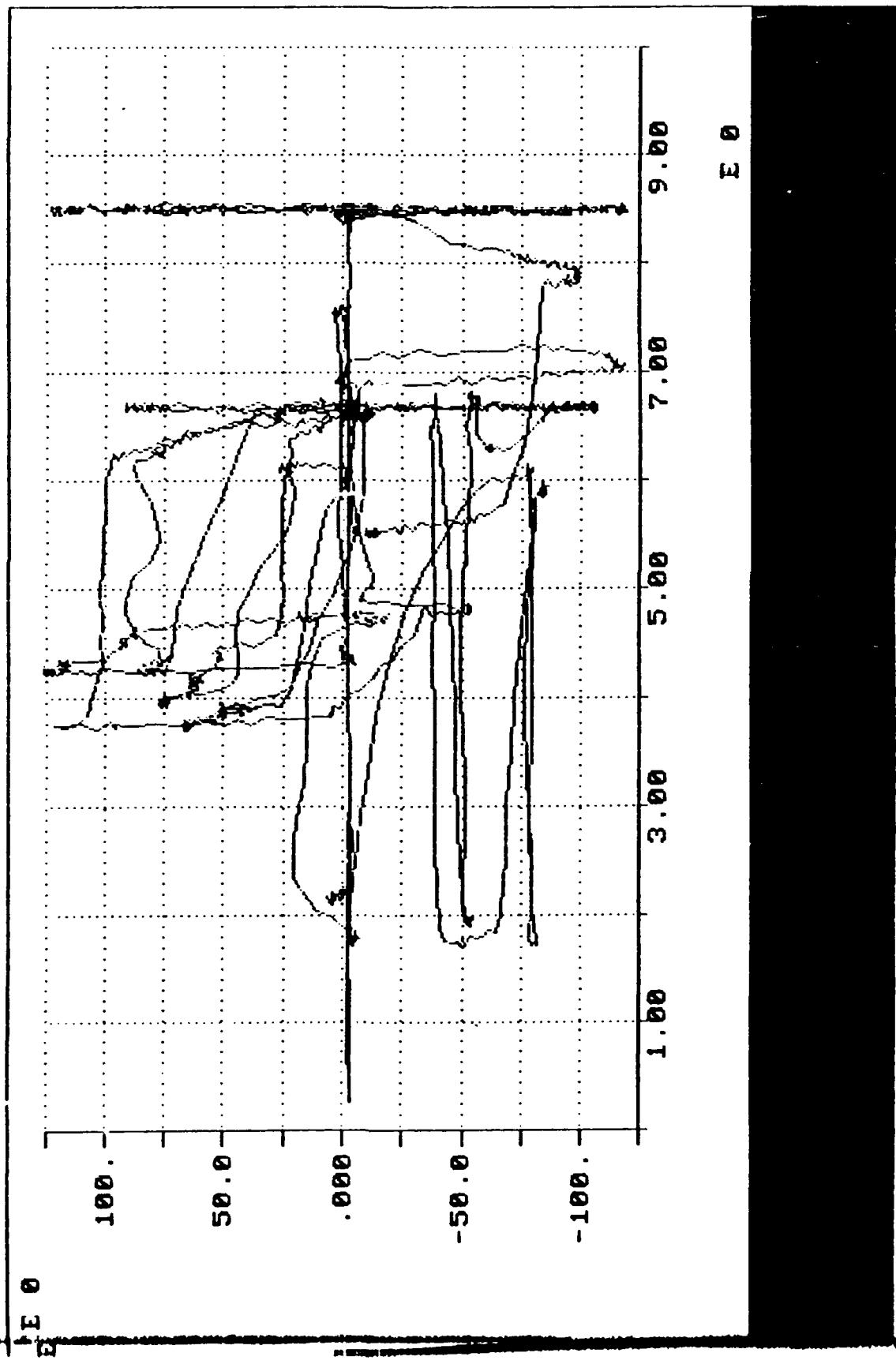
710105



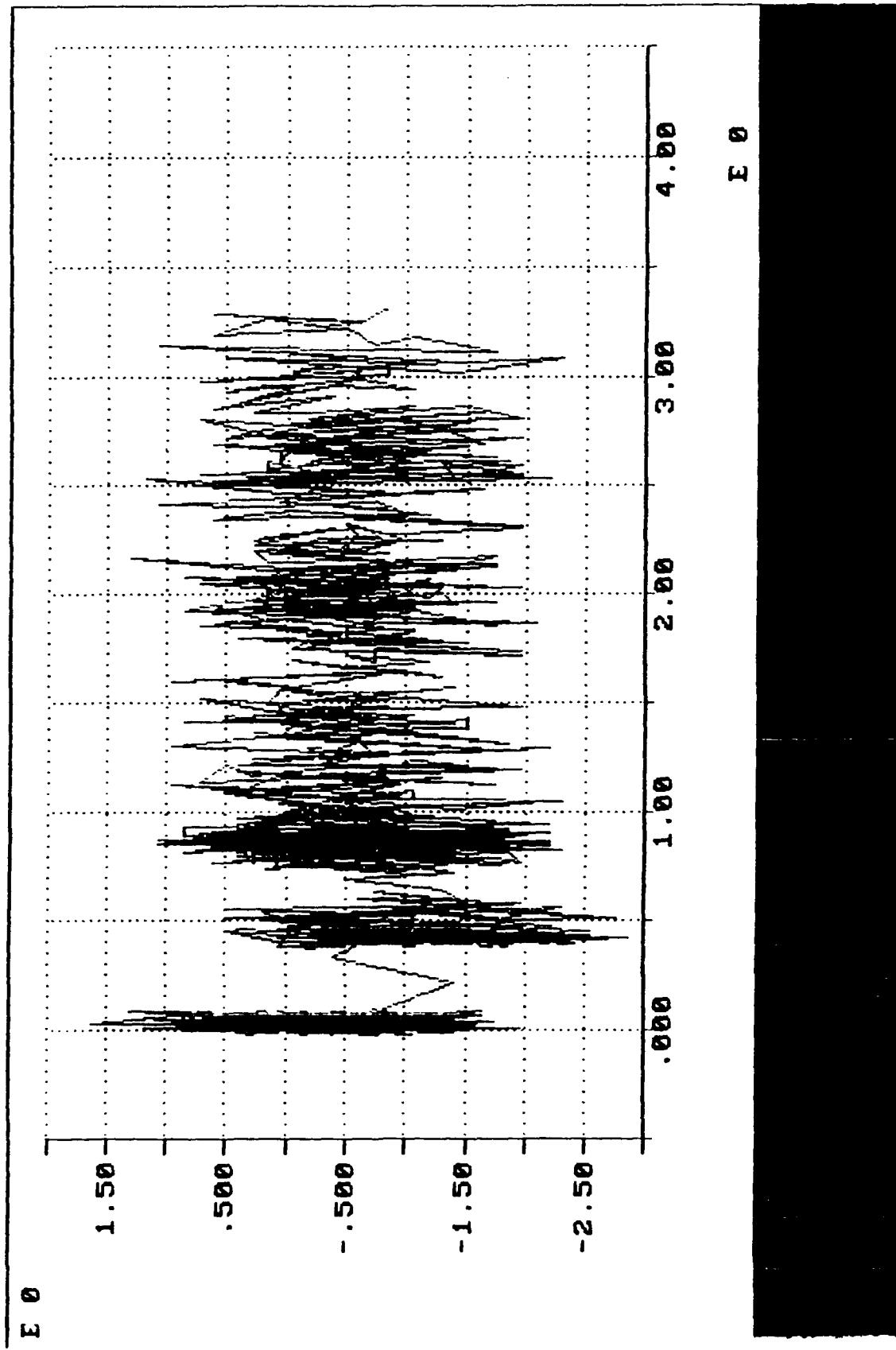
III & 105

X² {S, n}

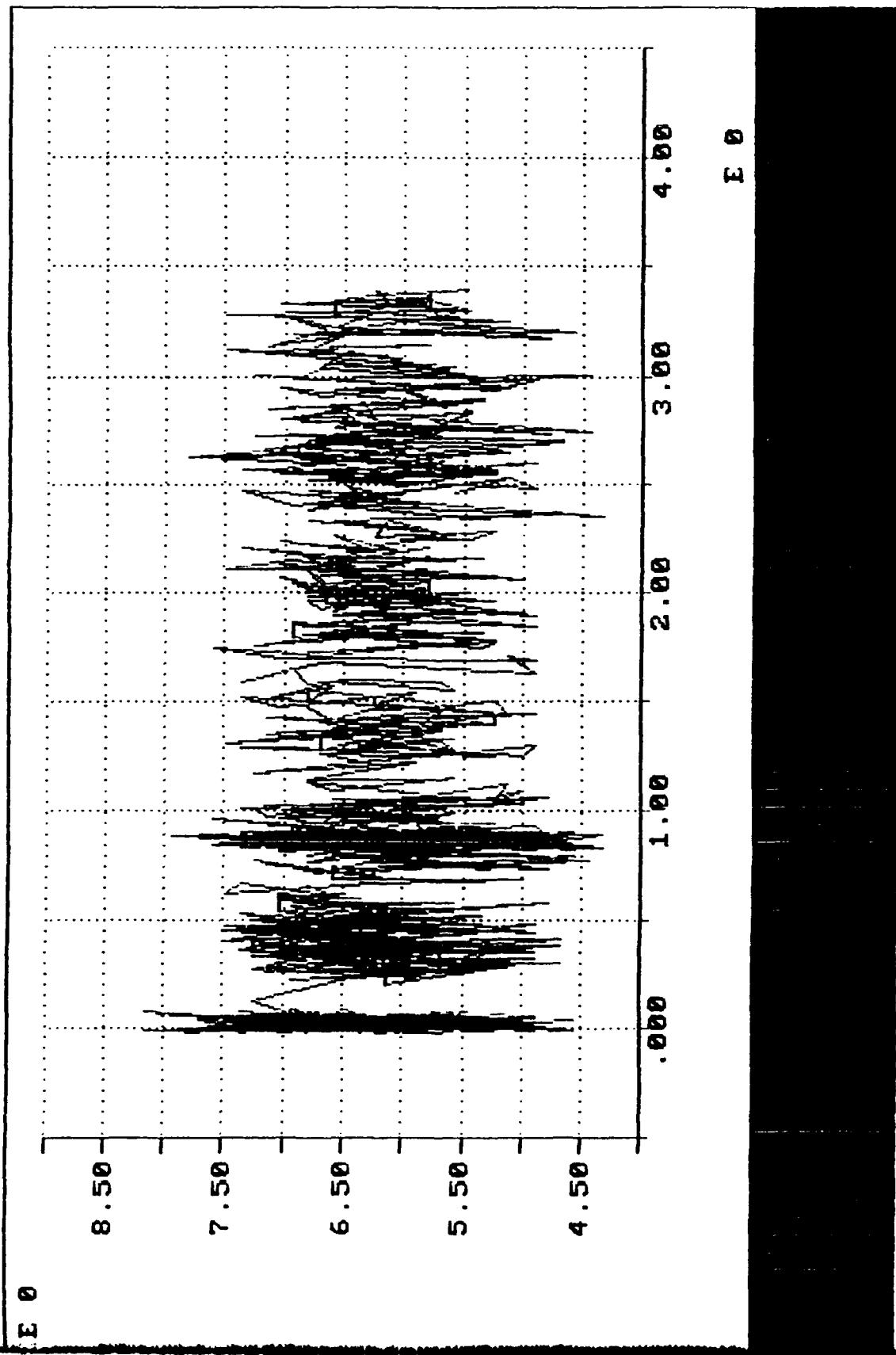




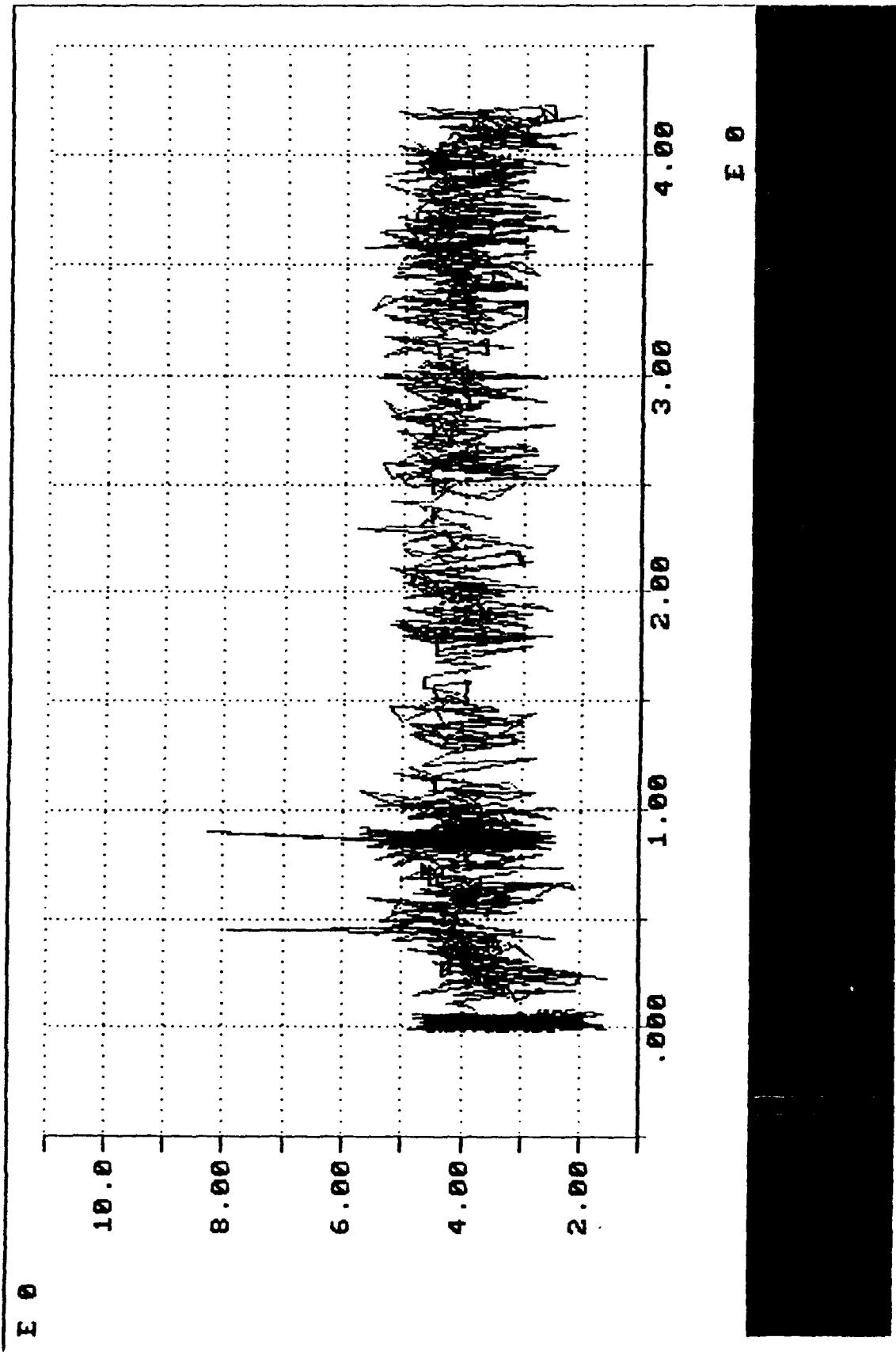
Aug 30

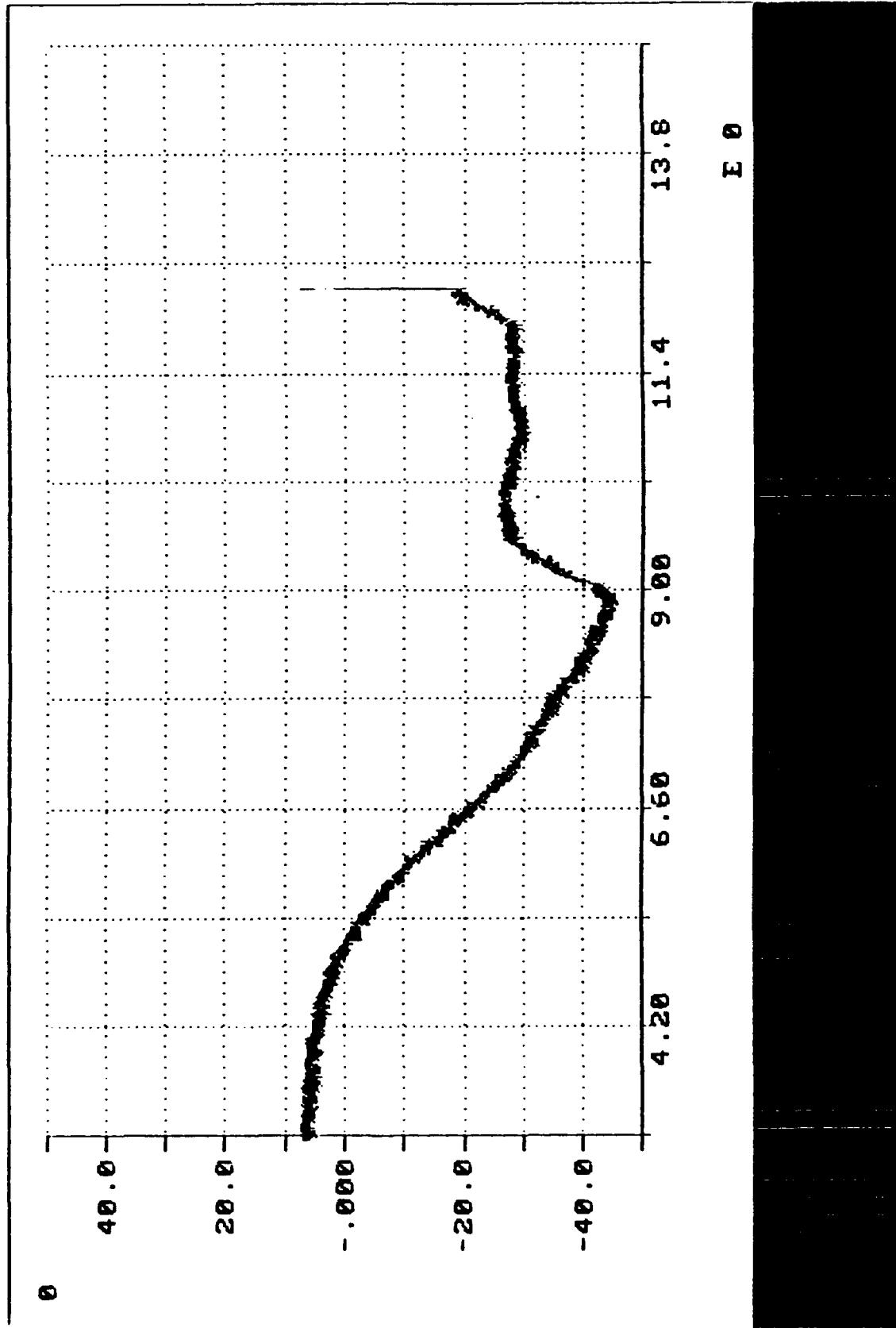


AUG 7 19

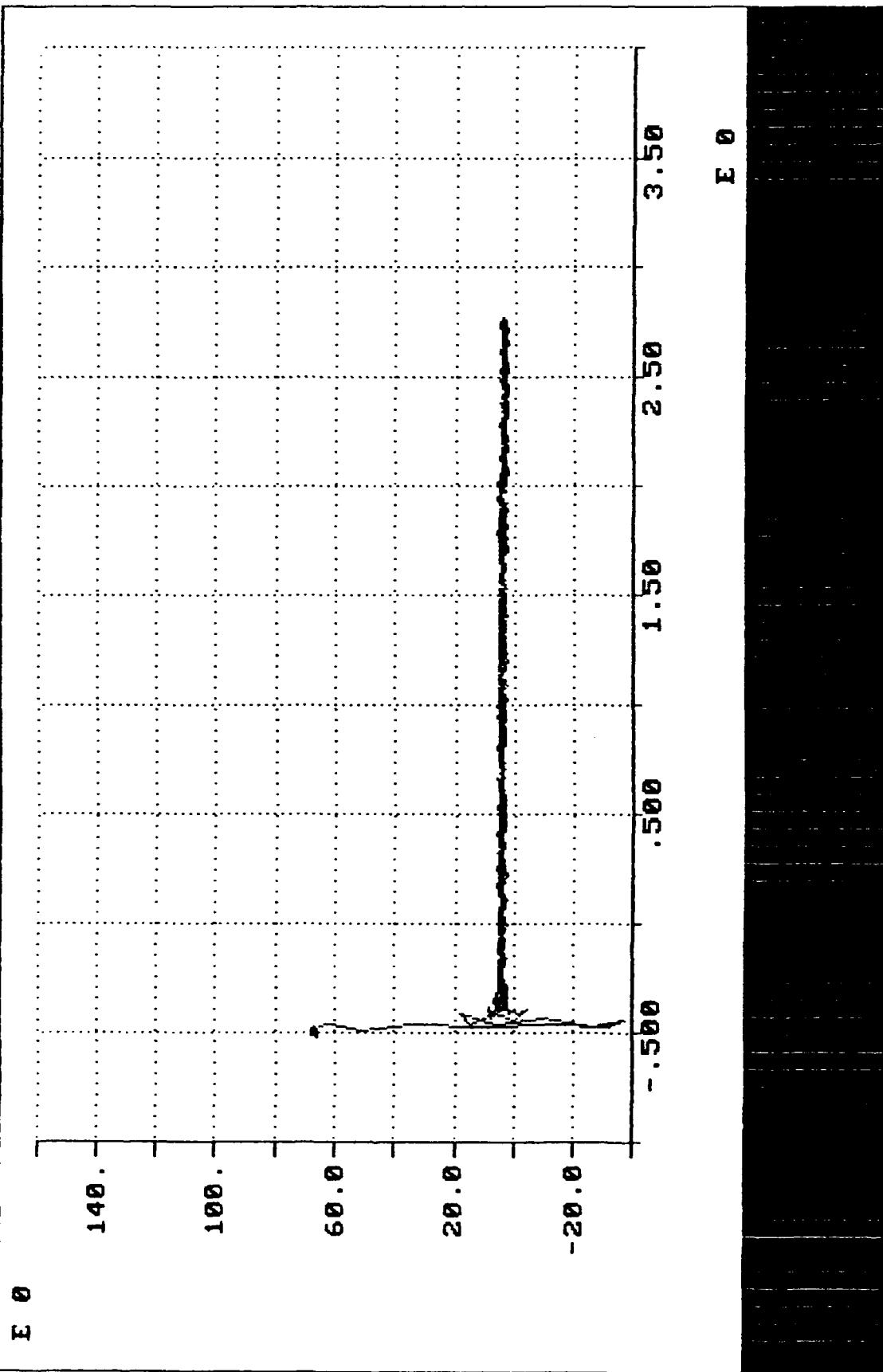


Aug 24



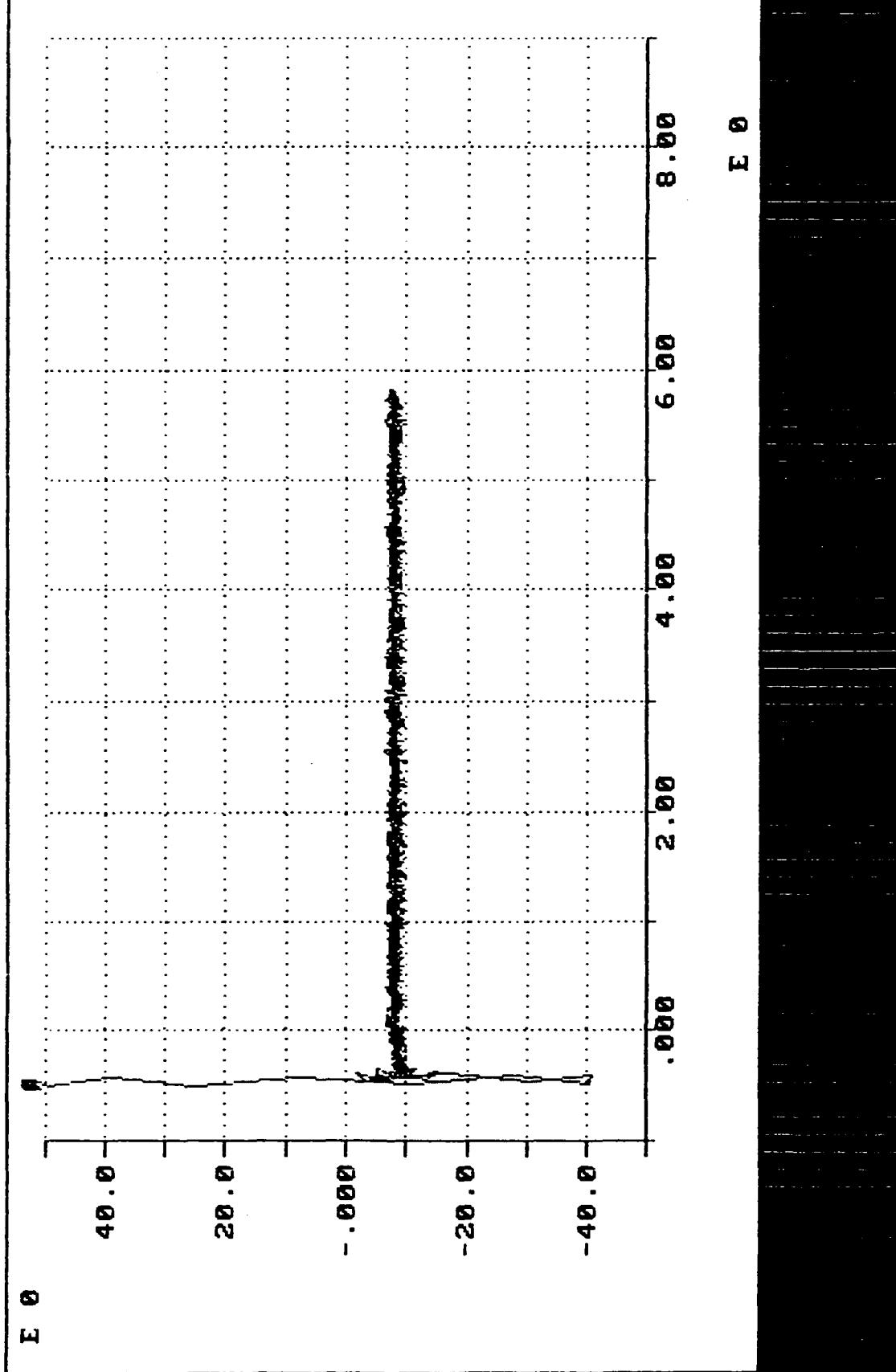


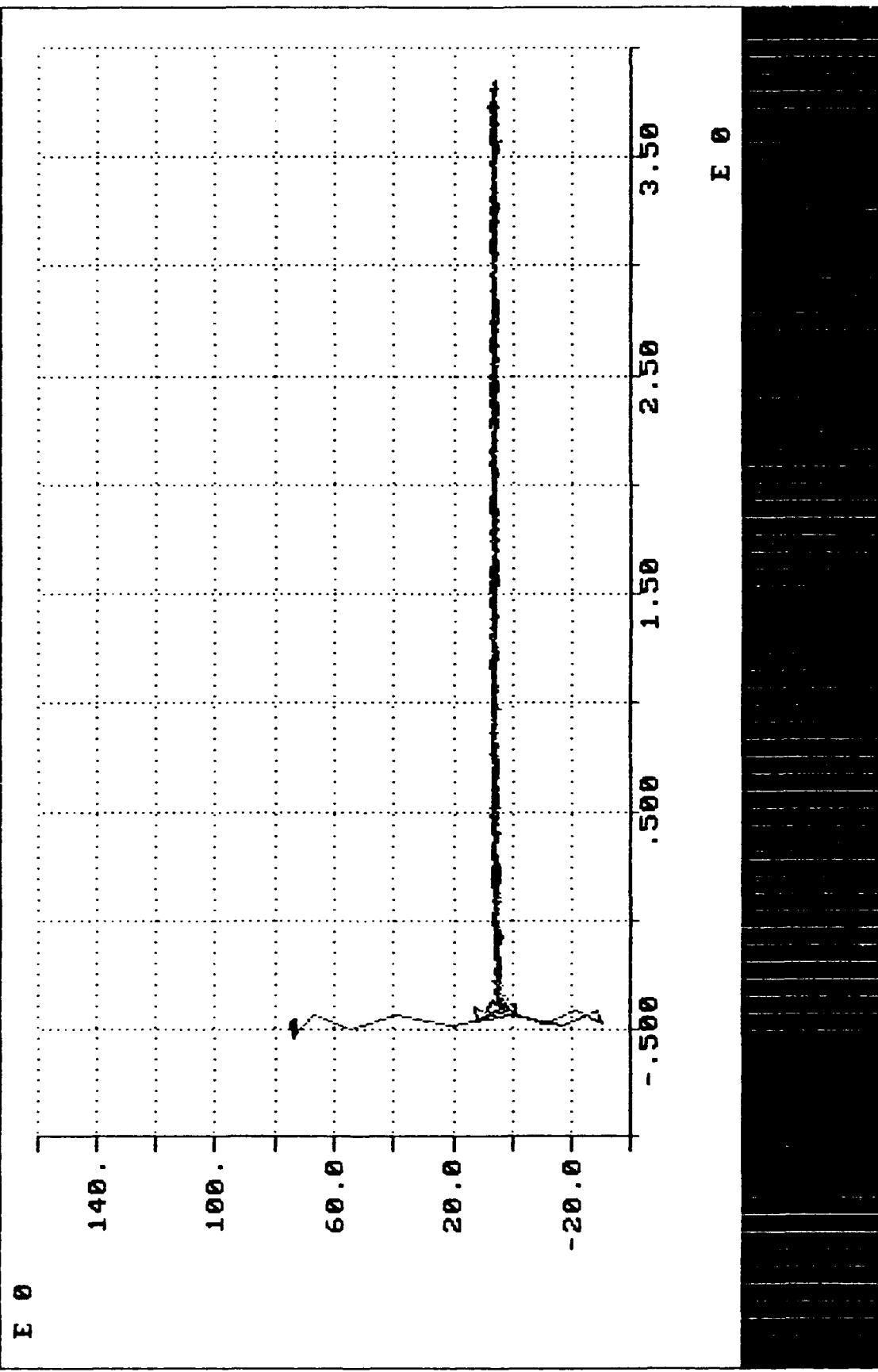
Aug 26



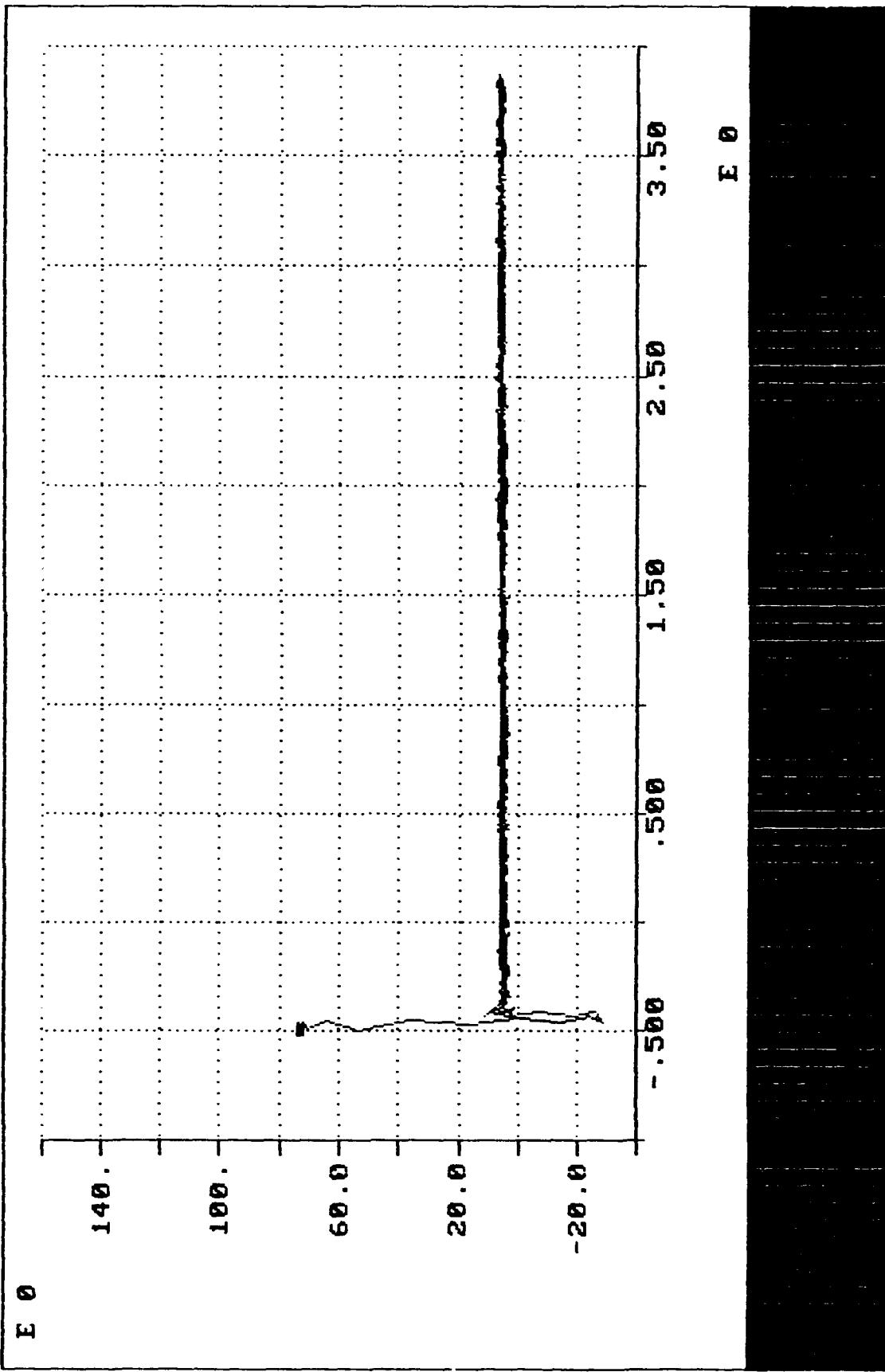
Aug 8

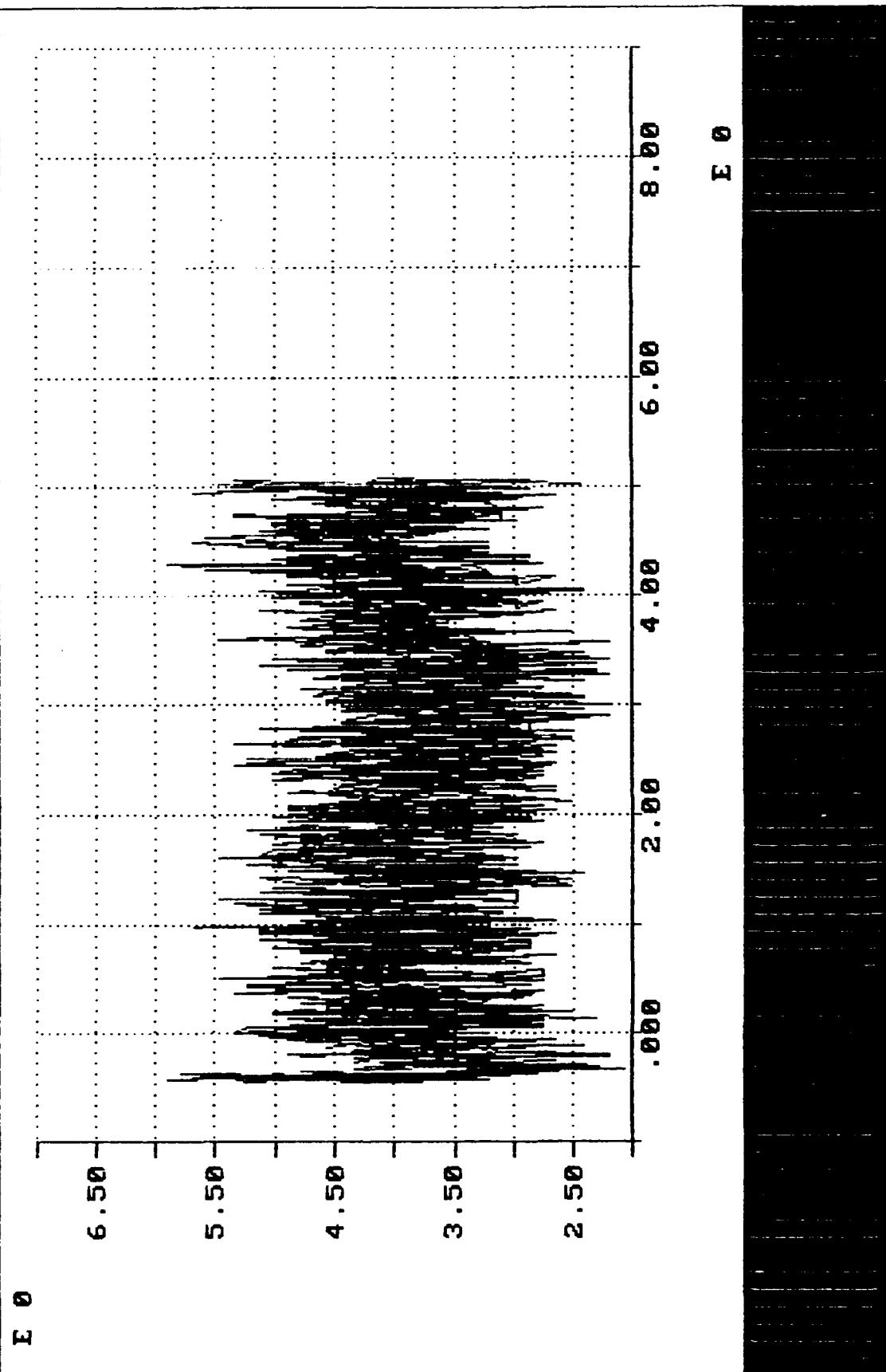
44-28





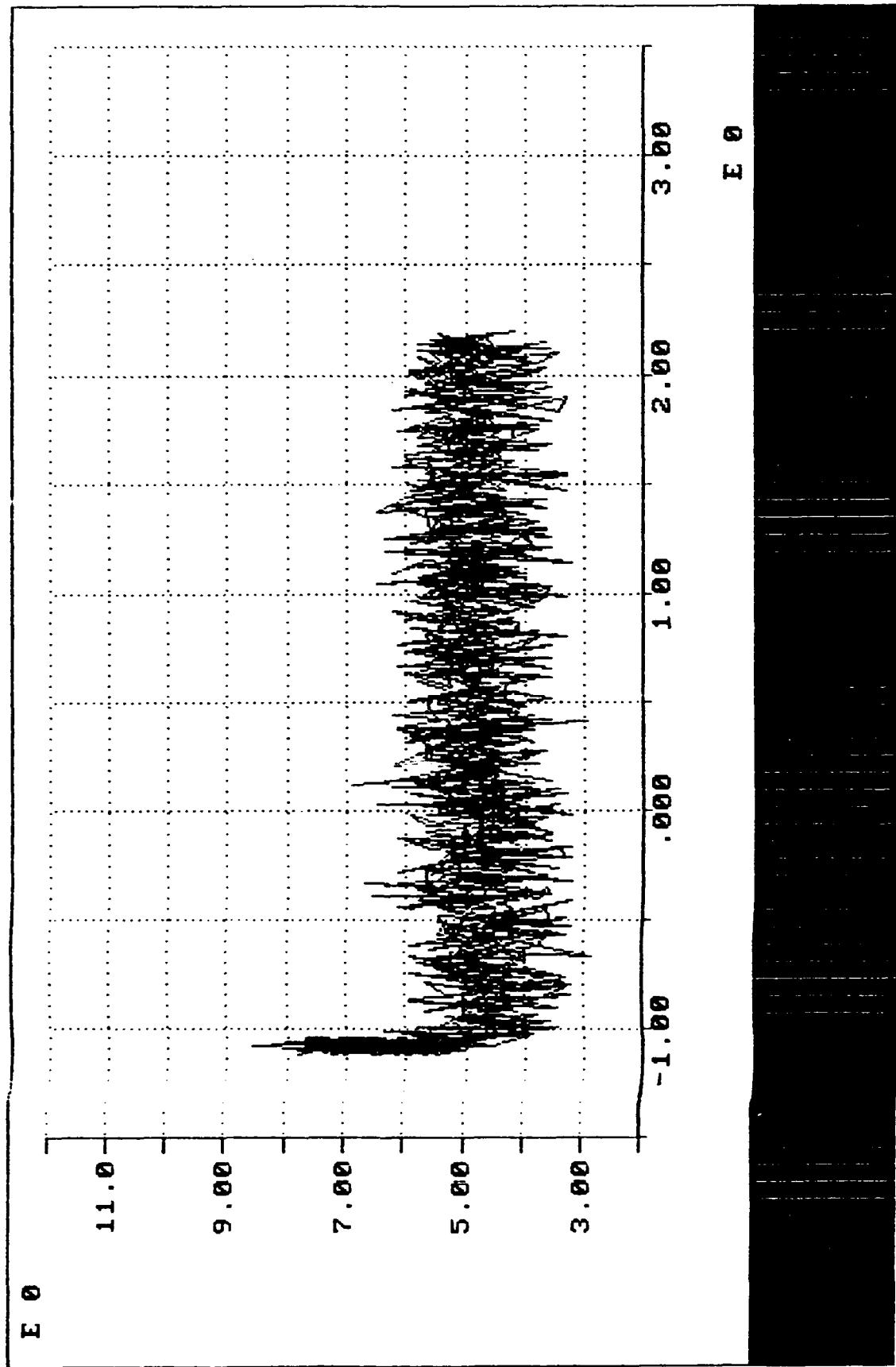
Aug 8



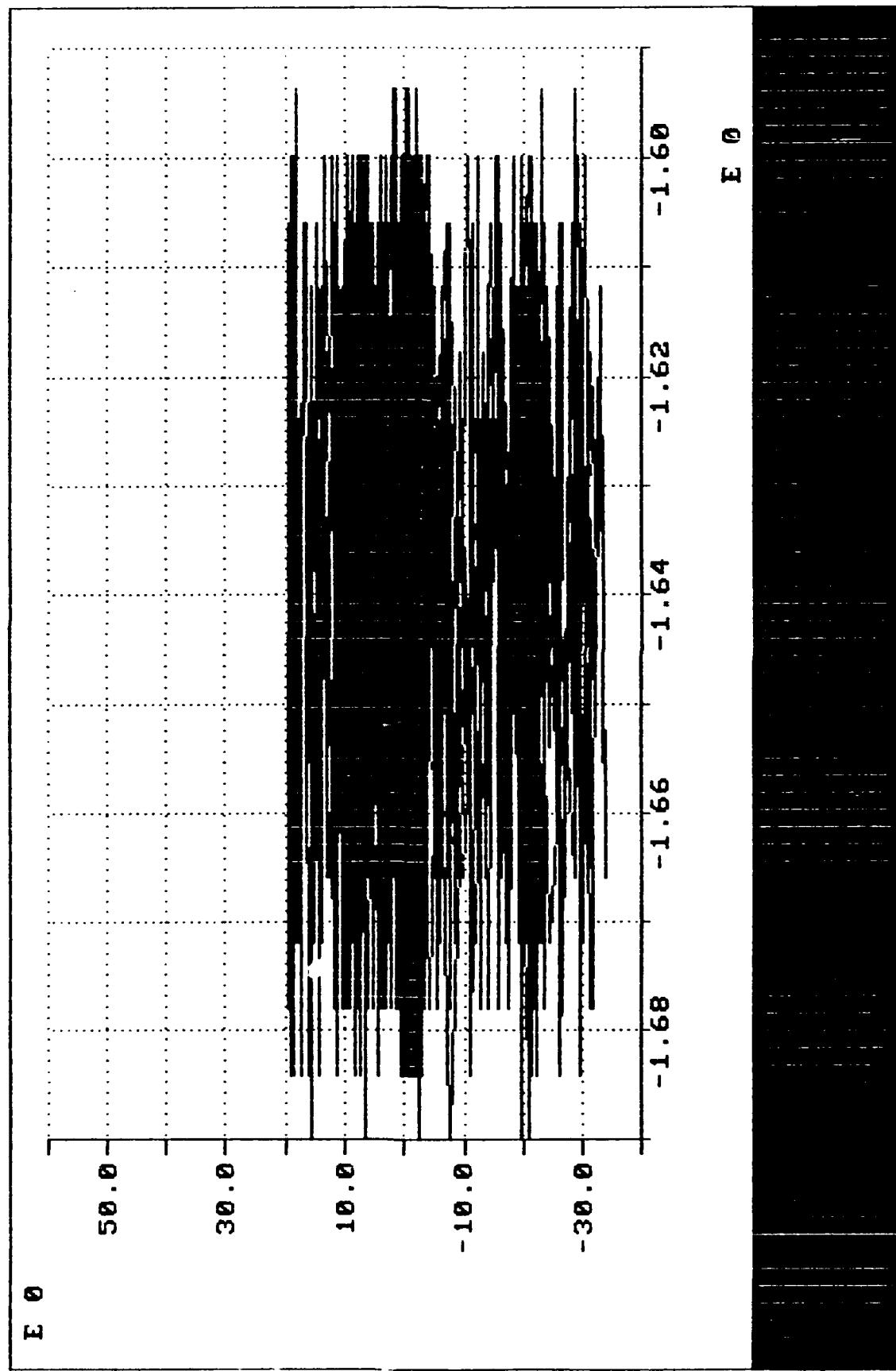


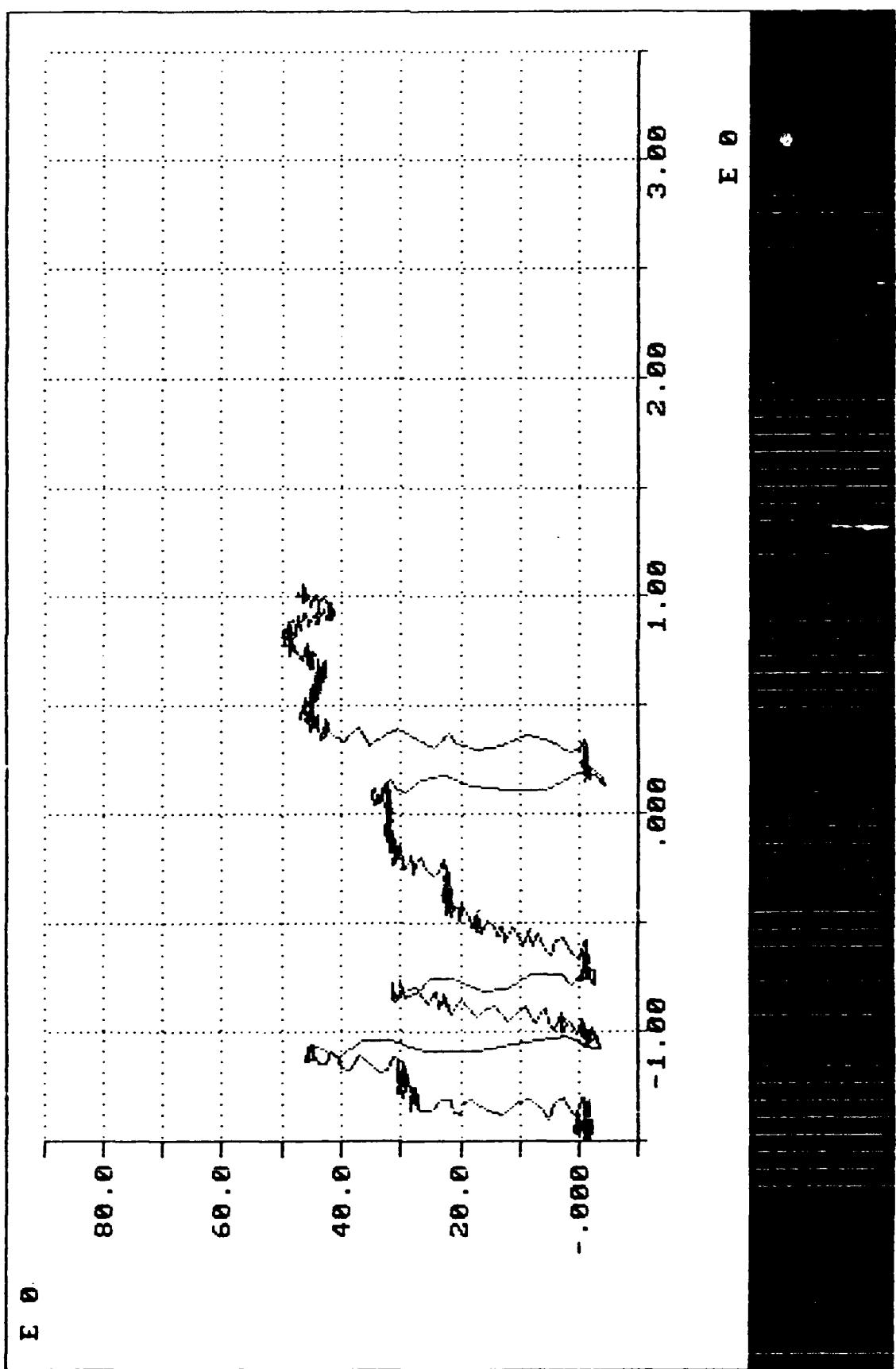
Aug 86

AUG 4



AUC 9E





JUL26

File Name: <B:AUG28V.USR>

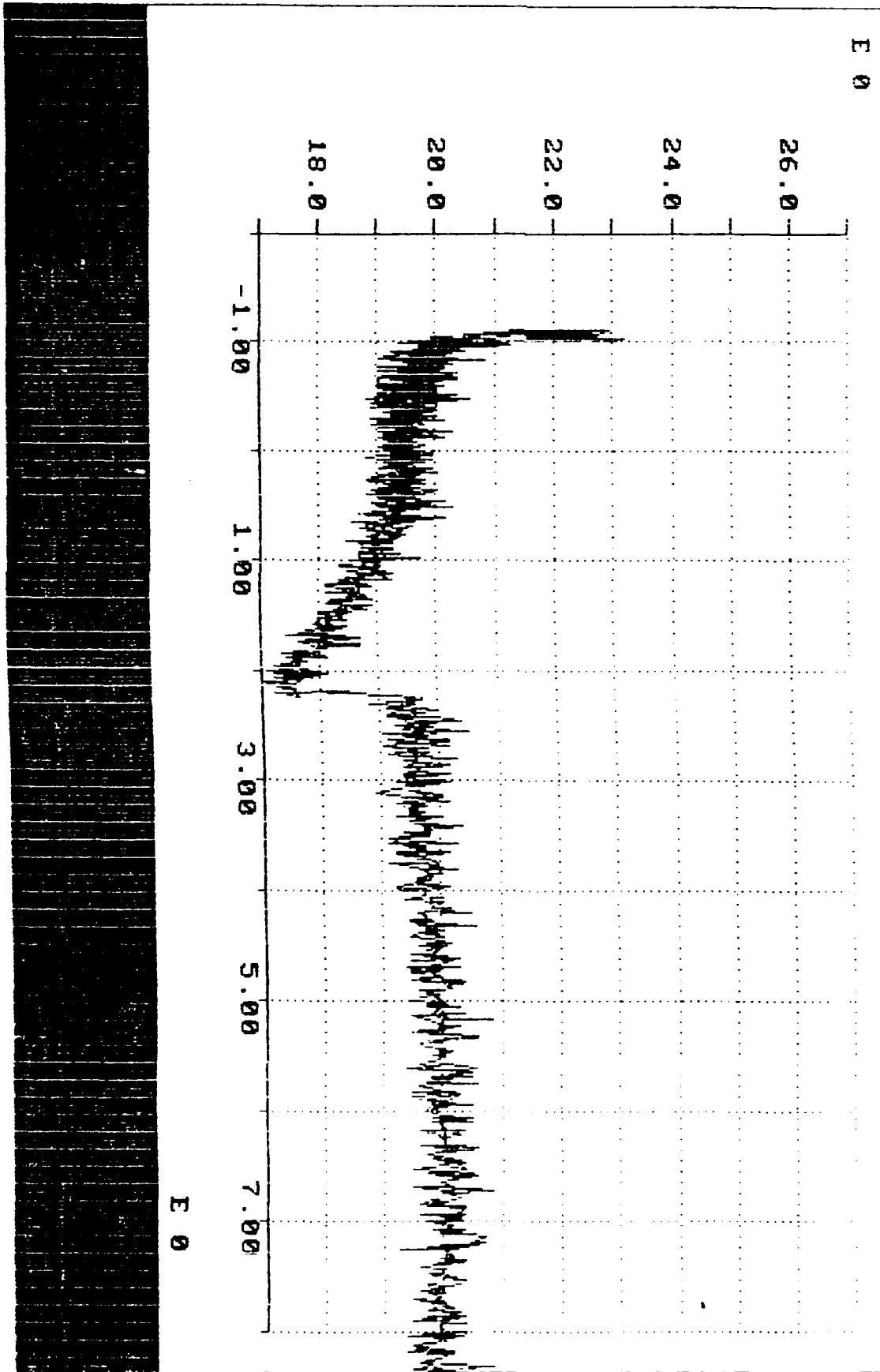
Quit/Continue: *

8 Comments

Subfiles (total #: 7)

- 1> FERRONYL, 100 DEGREES CONTROLLED
- 2> SOME BLANCHING
- 3> ONE OF THE BEST WELDS TODAY

: Start# Shape #Repts :
: 1 2 x 256 7 :
:



AUG 28 V

Ferronyl Temp = 100

Fresh Eclipses

1MMMMMMMMMMMMMMMMMMMMMM File Input/Output MMMMMMMMMMMMMMMMMMMMM

File Name: <B:AUG28W.USR>

Quit/Continue: *

8 Comments

> FERRONYL, 50 DEGREES CONTROLLED
> NO VISUAL CHANGES
> NOT A VERY GOOD WELD
> BURLEIGH STAGE DID NOT MOVE AT THE BEGINN:

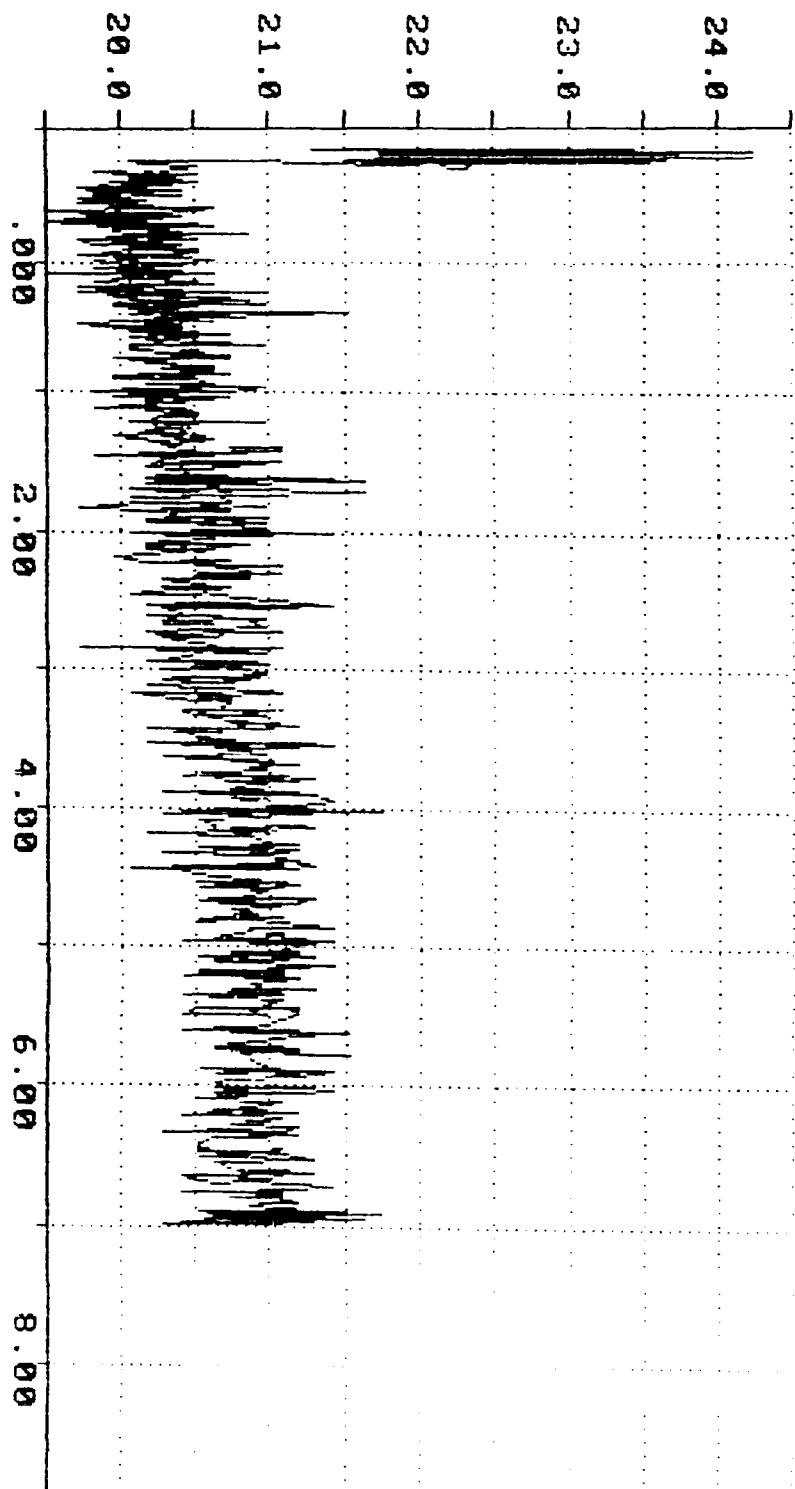
Subfiles (Total #: 9) :

: Start# Shape #Repts :

: 1 2 x 256 9 :

AUG 28W

Ferronyl Temp = 50
Fresh Bone



XXXXXXXXXXXXXXXXXXXXXXXXXXXX File Input/Output XXXXXXXXXXXXXXXXXXXXXXX;

File Name: <B:AUG28X.USR>

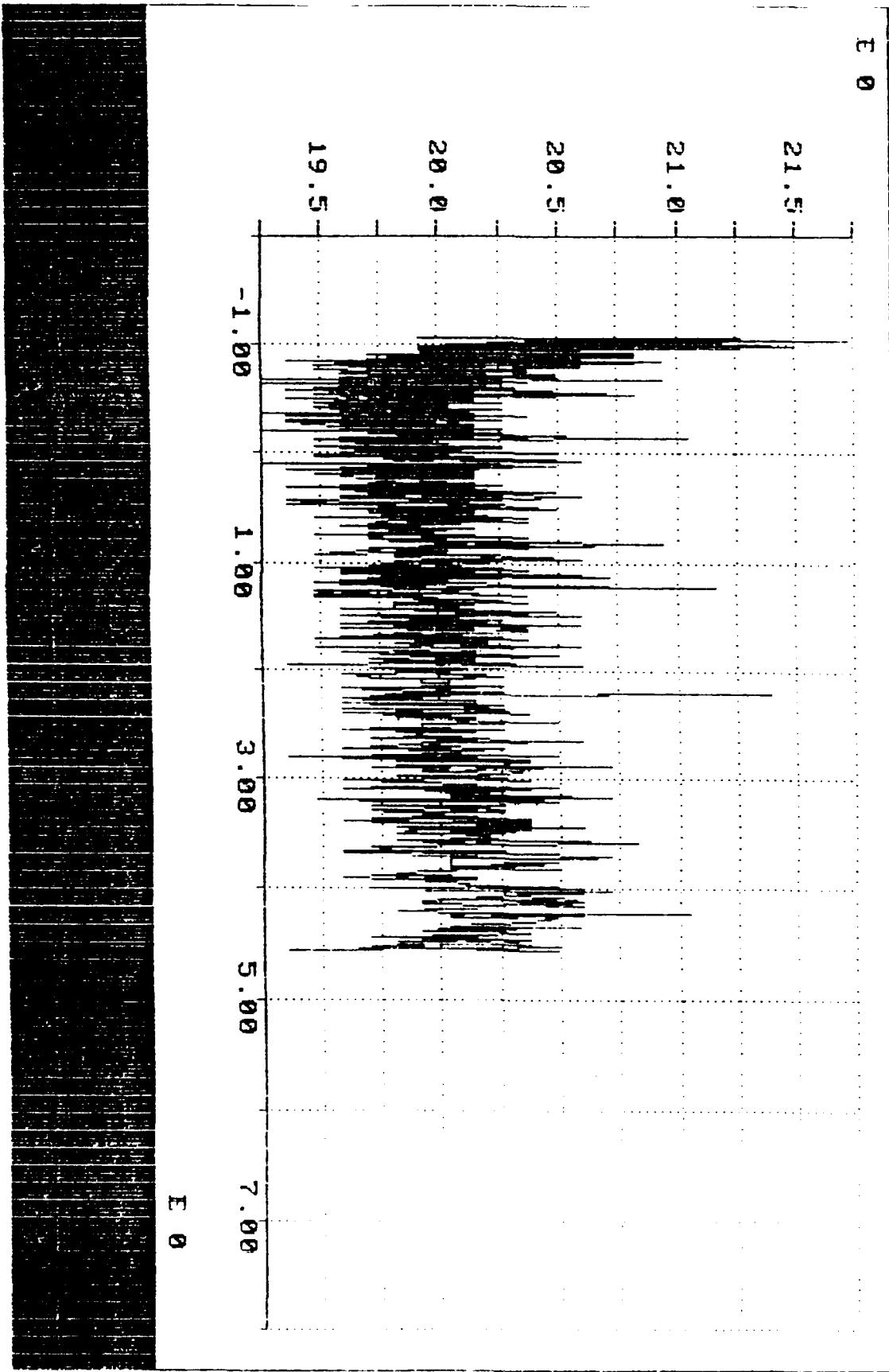
Quit/Continue: *

8 Comments

Subfiles (Total #: 6)

- 1> FERRONYL-60 DEGREES CONTROLLED
- 2> NO VISUAL CHANGES
- 3> ONE OF THE BETTER WELDS

: Start#	Shape	#Repts :
: 1	2 x 256	6 :
:		



File Name: <B:AUG28Y.USR>

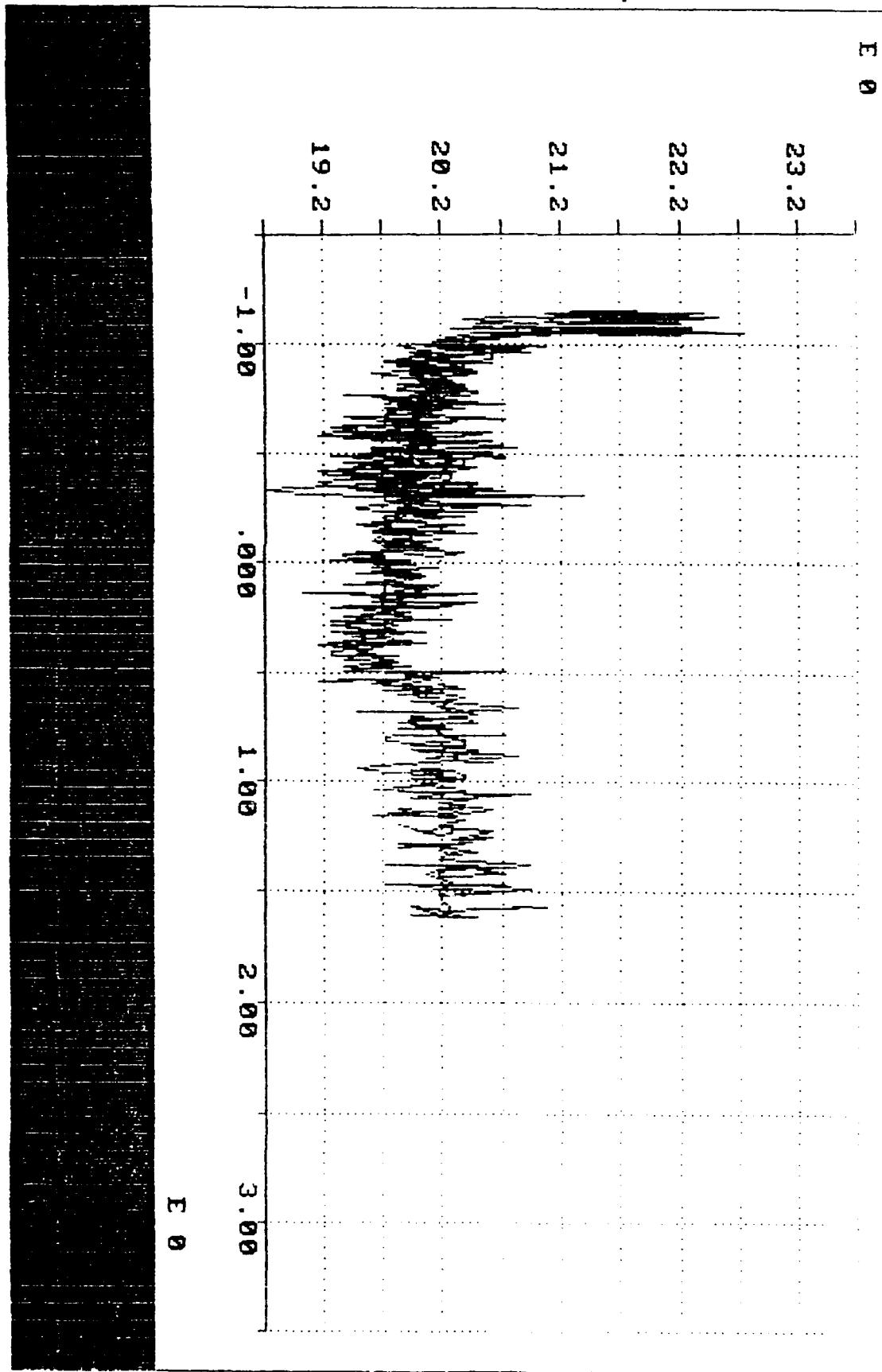
Quit/Continue: *

8 Comments

Subfiles (Total #: 4)

- : 1> INDIA INK, TEMP CONTROLLED AT 60
- : 2> SLIGHT DESSICATION
- : 3> A GOOD WELD
- : 4> LOOKS BETTER THAN FERRONYL

: Start#	Shape	#Repts :
: 1	2 x 256	4 :
:		:
:		:



AUG 28 Y

Ink Temple
Teng-chau

XXXXXXXXXXXXXXXXXXXXXXXXXXXX File Input/Output XXXXXXXXXXXXXXXXXXXXXXX

File Name: <B:AUG28AB.USR>

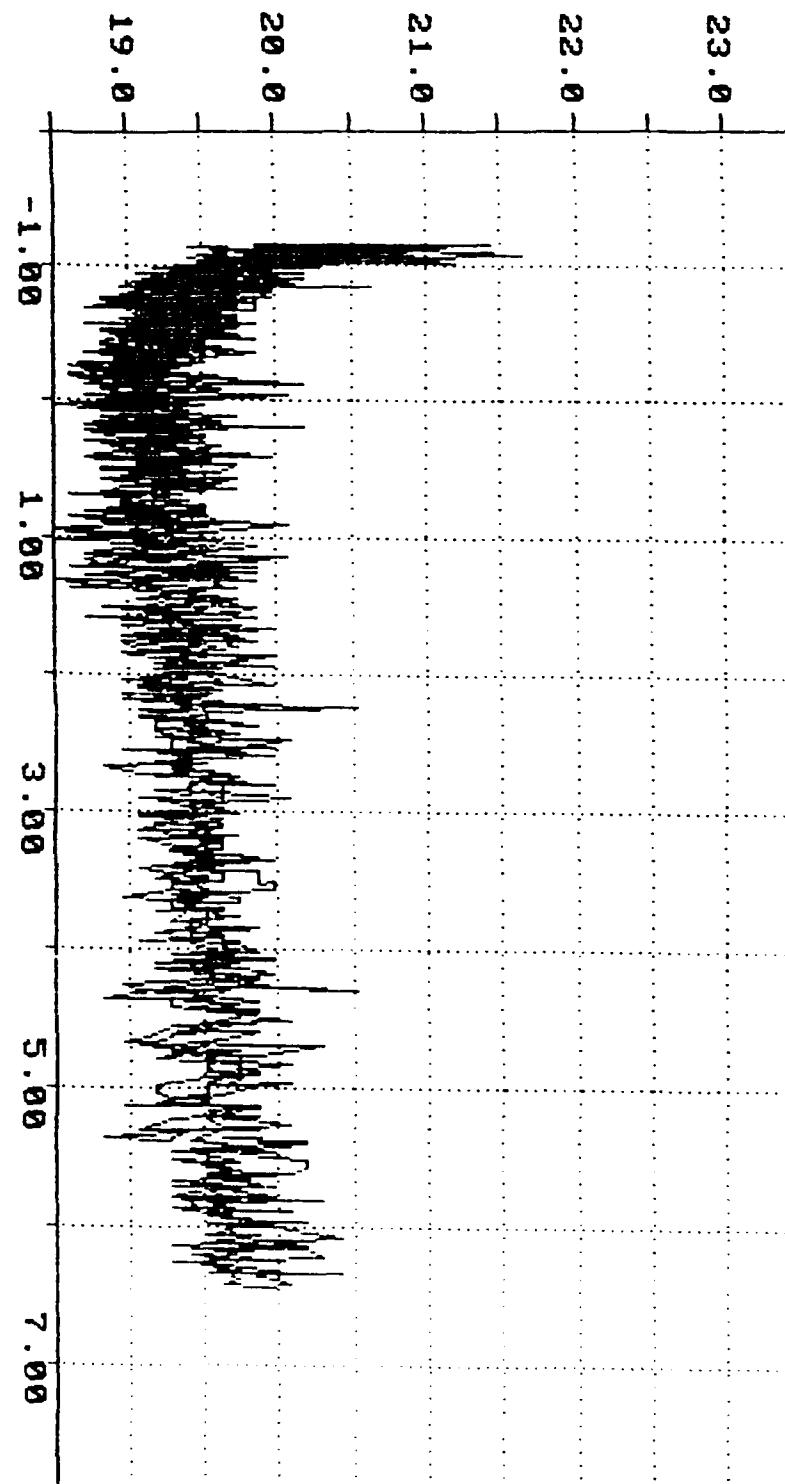
Quit/Continue: *

8 Comments

Subfiles (Total #: 7)

1> INDIA INK, 60 DEGREES, LARGE APERTURE, SL: Start# Shape #Repts :
2> WELDED. ONE OF THE BETTER WELDS : 1 2 x 256 7 :
3>
4>

AUG 28 1975



Temp = 60°

LARGE APERTURE
India Ink

XX
XX File Input/Output XXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXX

File Name: <B:AUG28AA.USR>

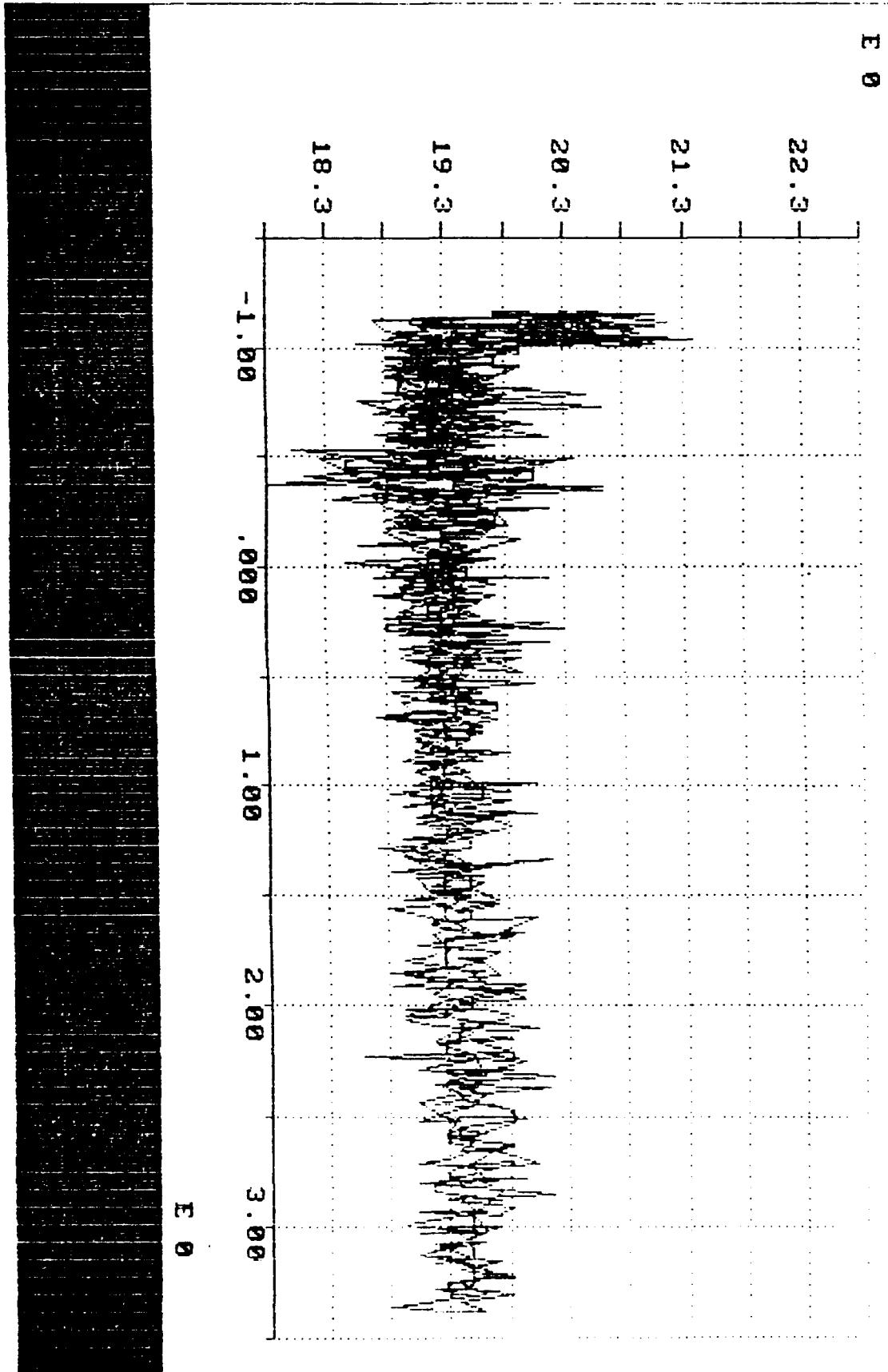
Quit/Continue: *

8 Comments

Subfiles (Total #: 5) :

1> LARGE APERTURE, INDIA INK, 50 DEGREES
2> WELDED

: Start# Shape #Repts :
: 1 2 x 256 5 :



AUG 28 AA

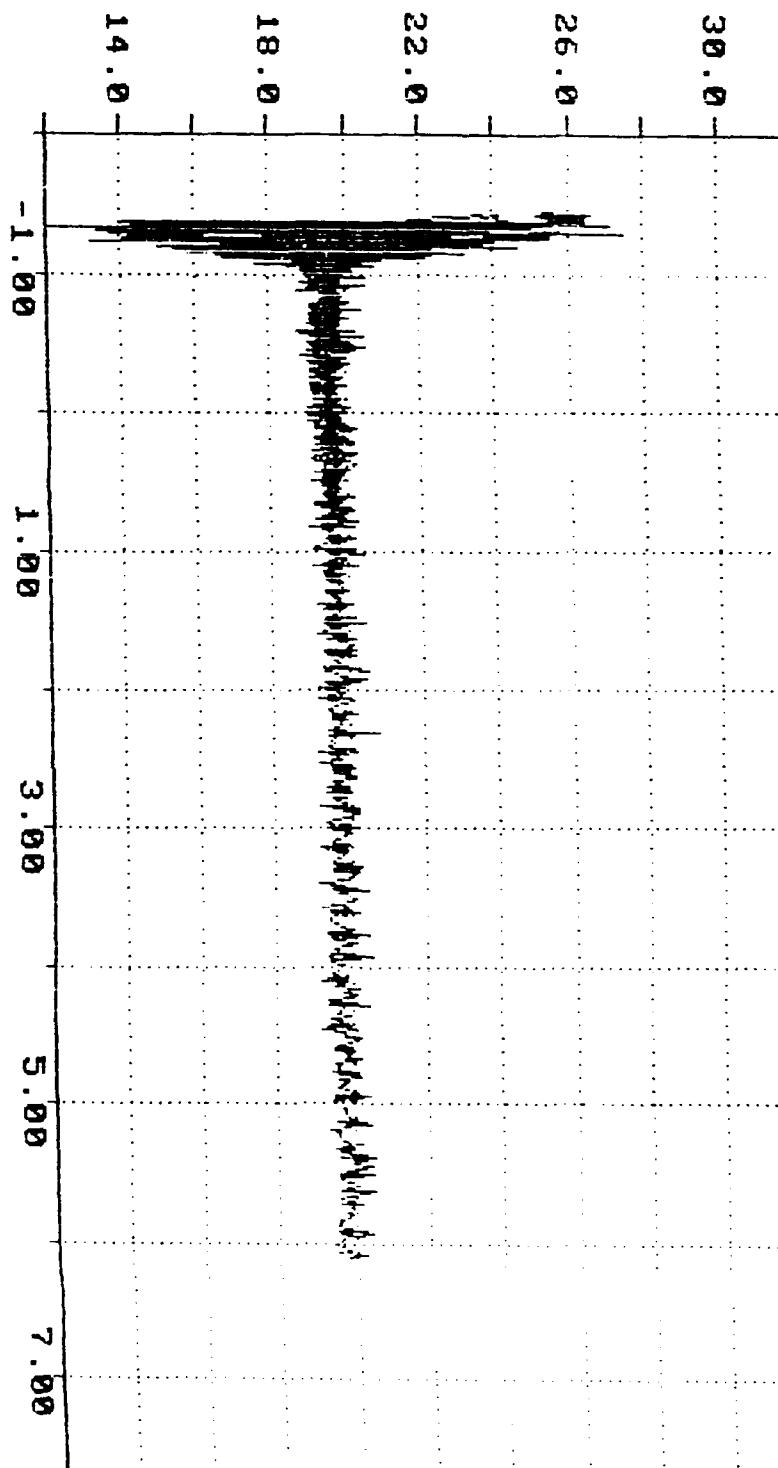
Large Aperture
India Ink
Temp = 50

File Name: <B:AUG28Q.USR>

Quit/Continue: *

8 Comments

Subfiles (Total #: 7)



AUG 28 Q

Fernow Temp 80°
TC too deep

XXXXXXXXXXXXXXXXXXXXXXXXXXXX File Input/Output .XXXXXXXXXXXXXXXXXXXXXXXXXXXX

File Name: <B:AUG28R.USR>

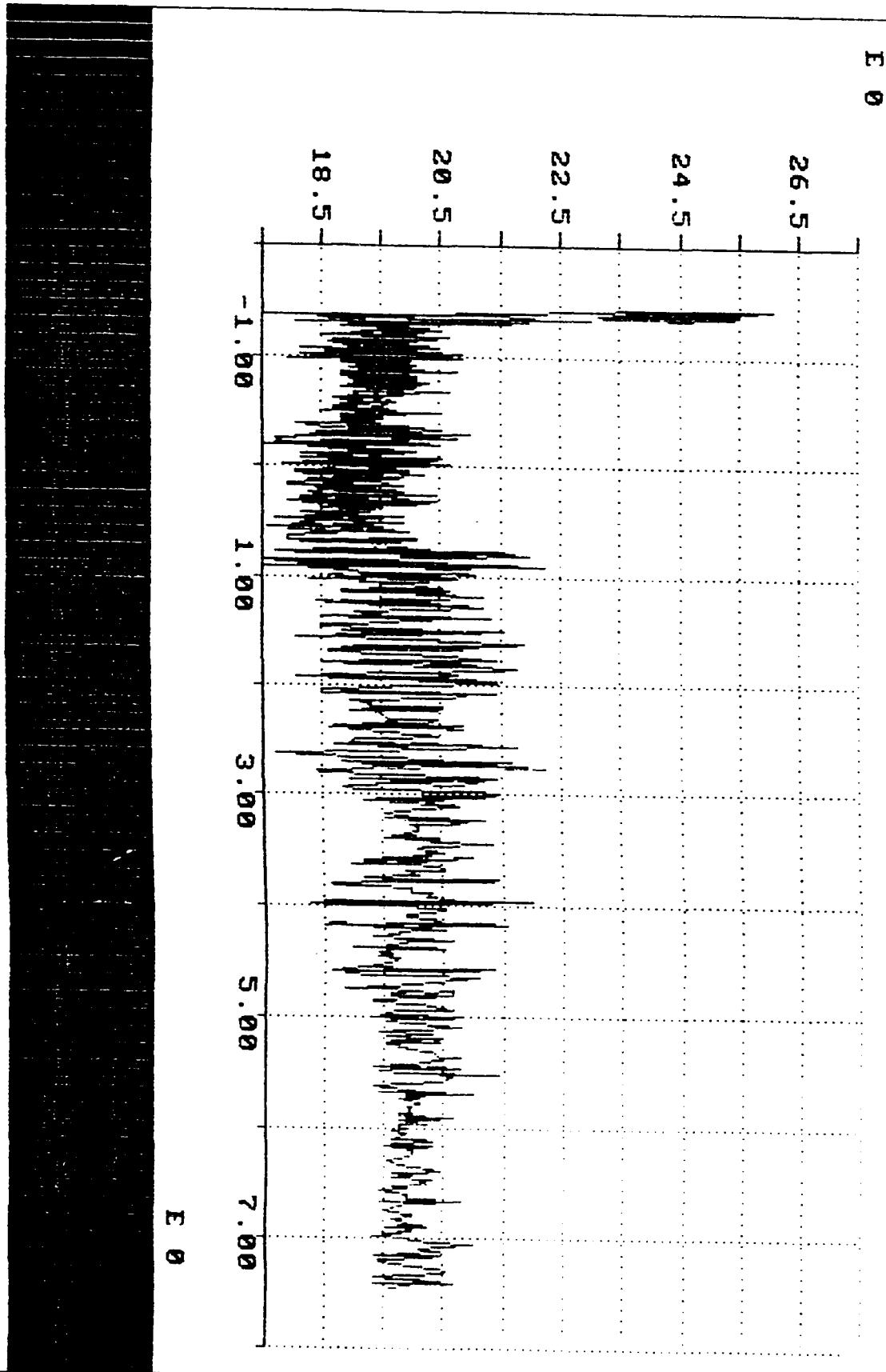
Quit/Continue: *

8 Comments

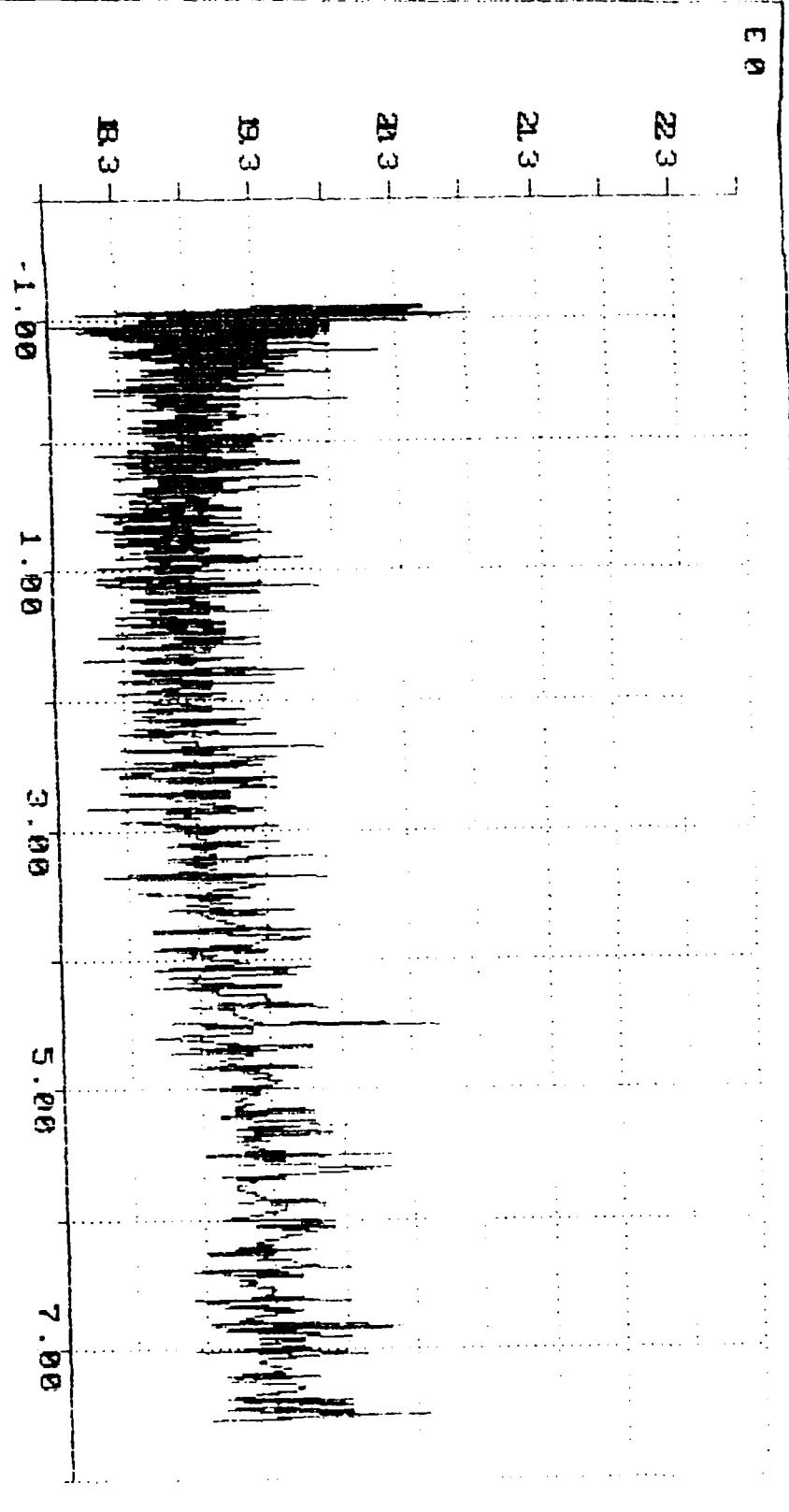
Subfiles (Total #: 7)

- 1> REPEAT FERRONYL-TEMP = 80
2> BETTER TC PLACEMENT
3> SOME BLANCHING
4> WELDED

: Start# Shape #Reots :
: 1 2 x 256 :
: : :
: : :



Ferronyl Temp 80°



Aug 28

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File Name: <B:AUG28S.USR> Quit/Continue: *
8 Comments Subfiles (Total #: 8) . . :
1> FERRONYL TEMPERATURE AT 100 DEGREES : Start# Shape #Repts :
2> SOME BLANCHING AND DESSICATION : 1 2 x 256 8 :
3> WELD WAS NOT VERY STRONG :
MMMM File Input/Output MMMMM File Input/Output MMMMM File Input/Output MMMMM File Input/Output

LMMMMMMMMMMMMMMMMMMMMMMMM File Input/Output LMMMMMMMMMMMMMMMMMMMMMM

File Name: <B:AUG28T.USR>

Quit/Continue: *

8 Comments

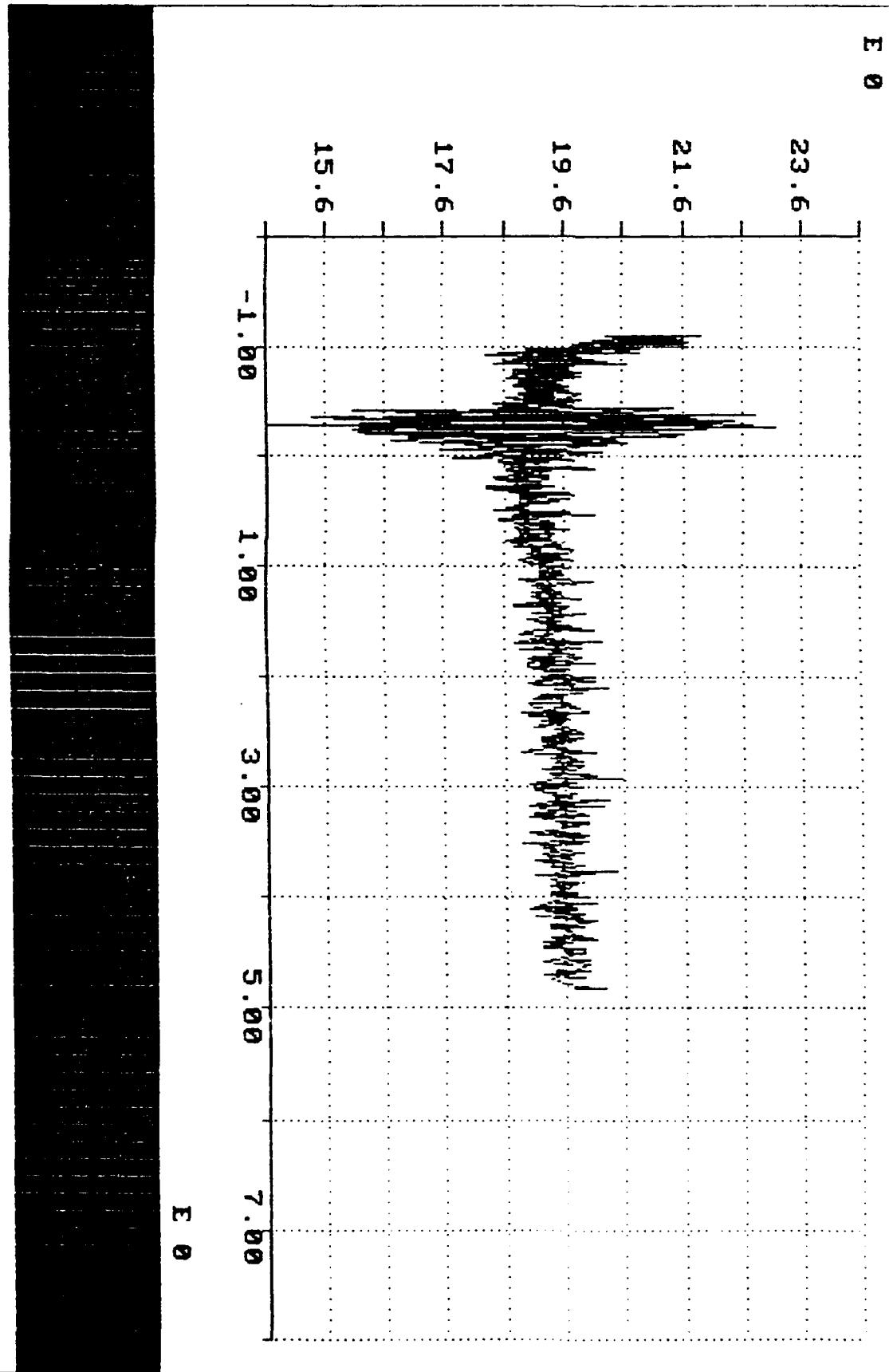
Subfiles (Total #: 5)

: 1> FERRONYL, 70 DEGREES, TC MAX=55, TC TOO D: Start# Shape #Repts :

: 2> WE HAVE A WELD

: 1 2 x 256

: 5



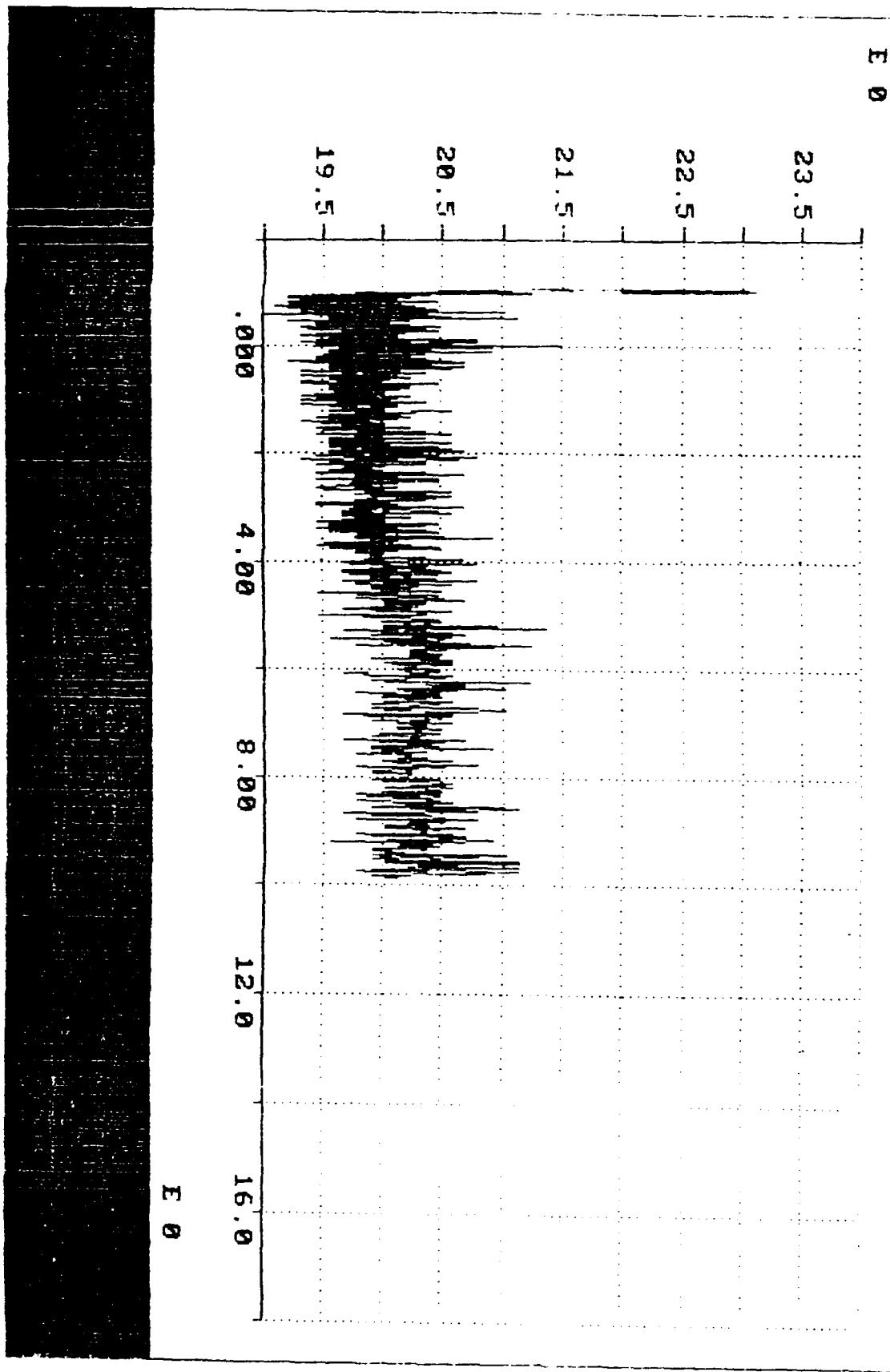
Fresh Bonded
Ferronyl Temp = 70

File Name: <B:AUG26U.JSR>

Quit/Continue: *

8 Comments

- Subfiles (Total #: 7) :
1> FERRONYL, TEMP SET AND CONTROLLED AT 80 D: Start# Shape #Repts :
2> GES 1 2 x 256 7 :
3> LOOKED STRONGER THAN THE 70 DEGREE AND ST:
4> WELDS FROM THIS MORNING.



Fresh Probe
Ferronyl Temp = 80°

IMMMMMMMMMMMMMMMMMMM File Input/Output MMMMMMMMMMMMMMMMMMMMMMMMMMMMMMMMM

File Name: <B:AUG28K.USR>

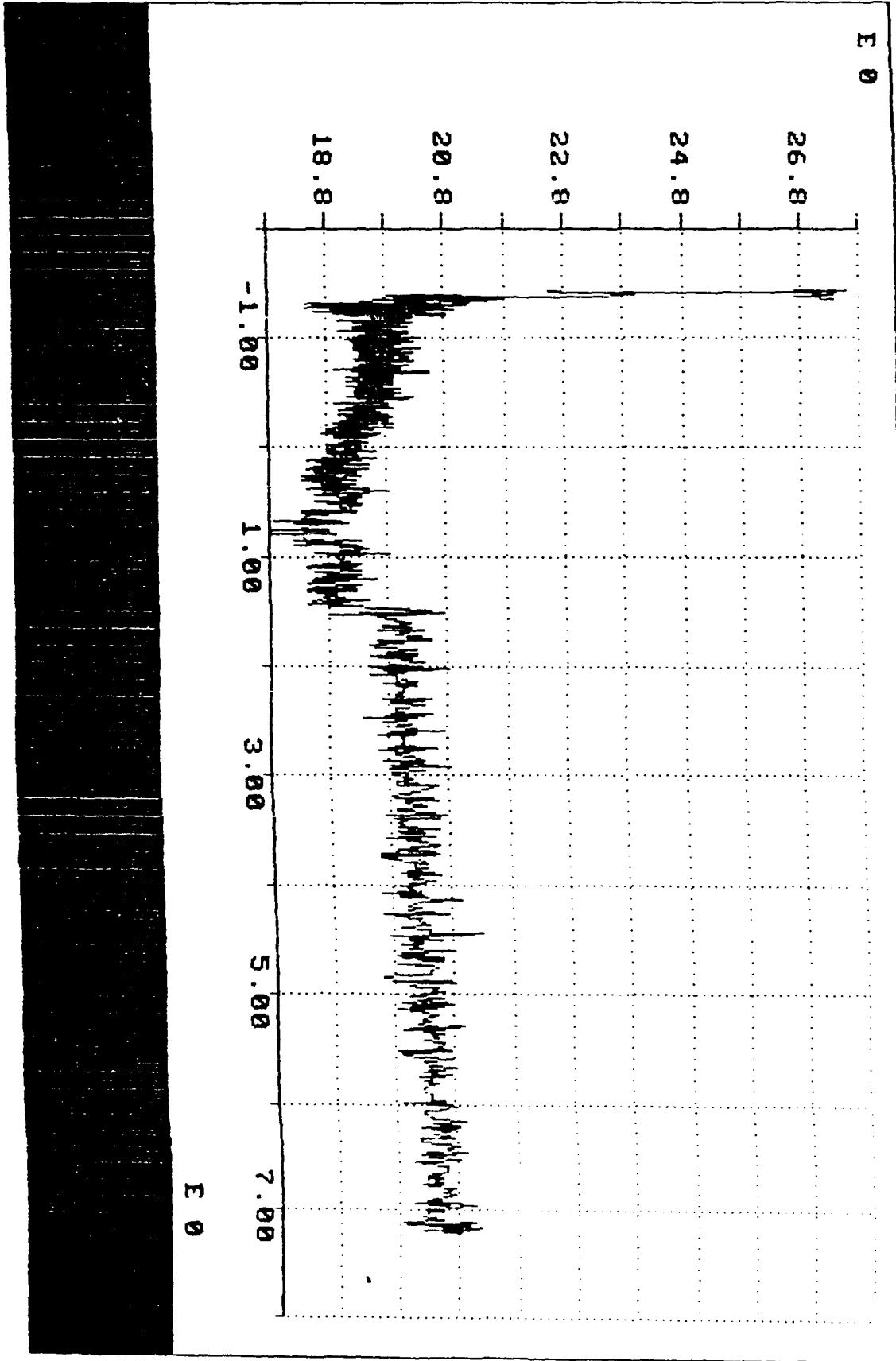
Quit/Continue: *

8 Comments

Subfiles (Total #: 7) :

1> INDIA INK, 70 DEGREES (NOT REACHED) SOME : Start# Shape #Repts :
2> WELDED, BROKE APART AT SEROSAL SURFACE. : 1 2 x 256 7 :
3> WELD SEEMED TO EXTEND THROUGHOUT TISSUE :

AUG 28K



India ink temp - 70

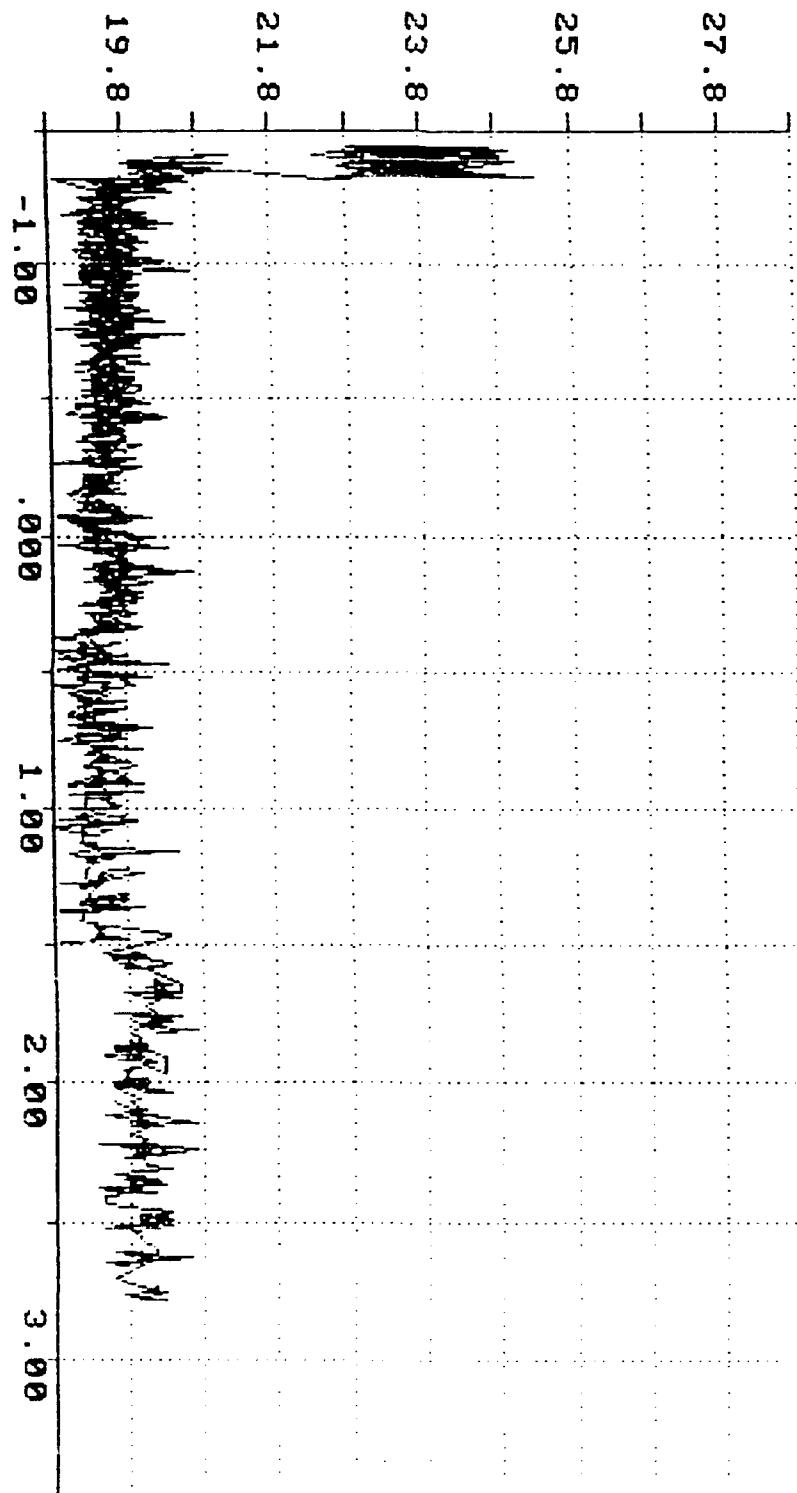
File Name: <B:AUG28L.USR>

Quit/Continue: *

: 8 Comments

Subfiles (Total #: 6

: 1> INDIA INK, TEMP = 80 DEGREES. TC MAX = 38: Start# Shape #Repts :
: 2> TISSUE BLANCHING AND SLIGHT SMOKE : 1 2 x 256 6
: 3> OK WELD :



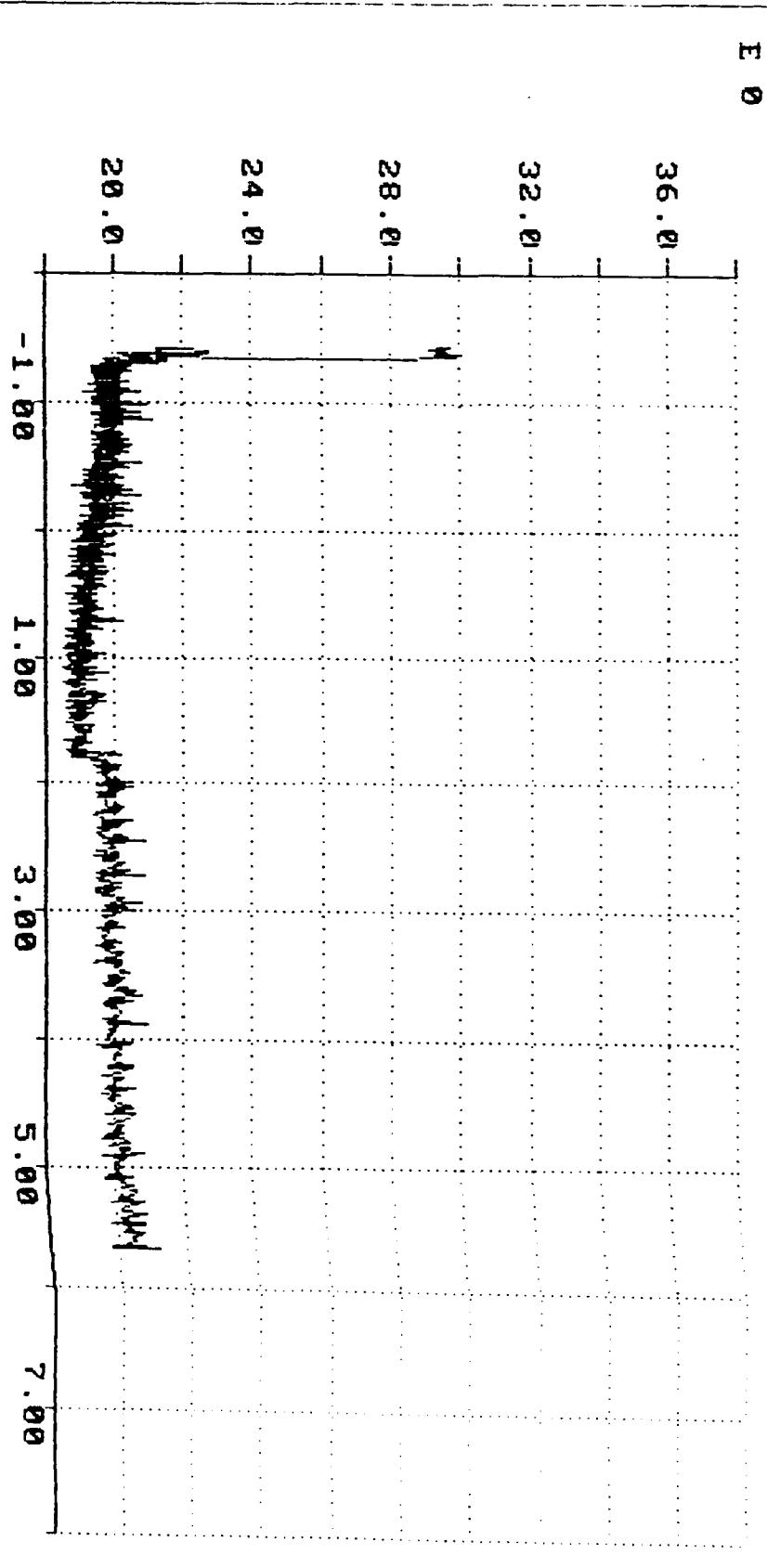
India ink Temp = 8°C

File Name: <B:AUG28M.USR>

Quit/Continue: *

8 Comments

Subfiles (Total #: 7)



AUG 28 M

India sub Temp = 100

File Name: <B:AUG28N.USR>

Quit/Continue: *

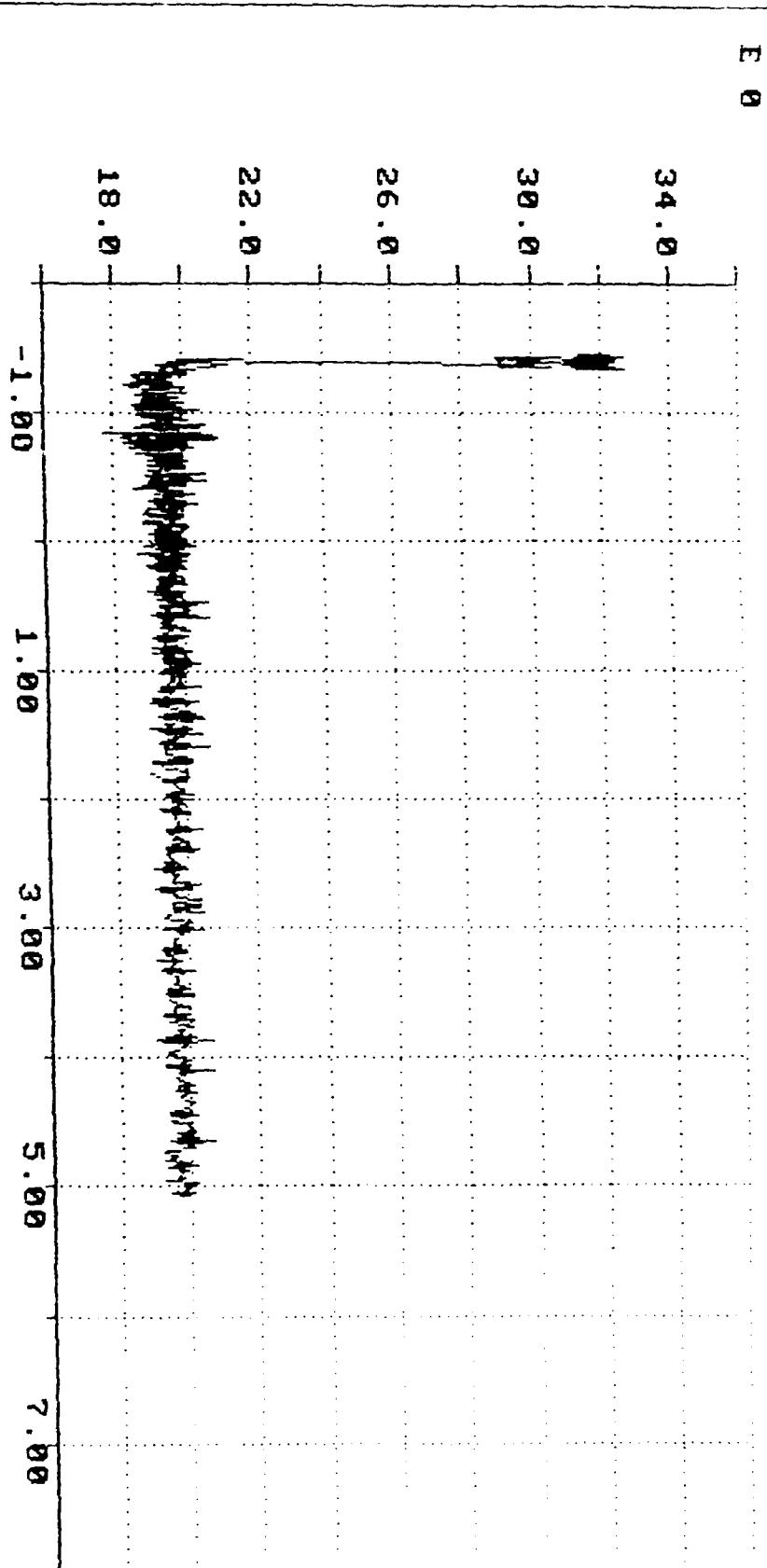
Subfiles (Total #: 7) :

8 Comments

FERRONYL TEMP = 50 CONTROLLED AT 41 DEGRE: Start#
SOME BLANCHING LOOKS LIKE A WELD : 1

Shape
 2×256

#Repts :
7 :



terrouist

$$\text{Temp} = 50^\circ$$

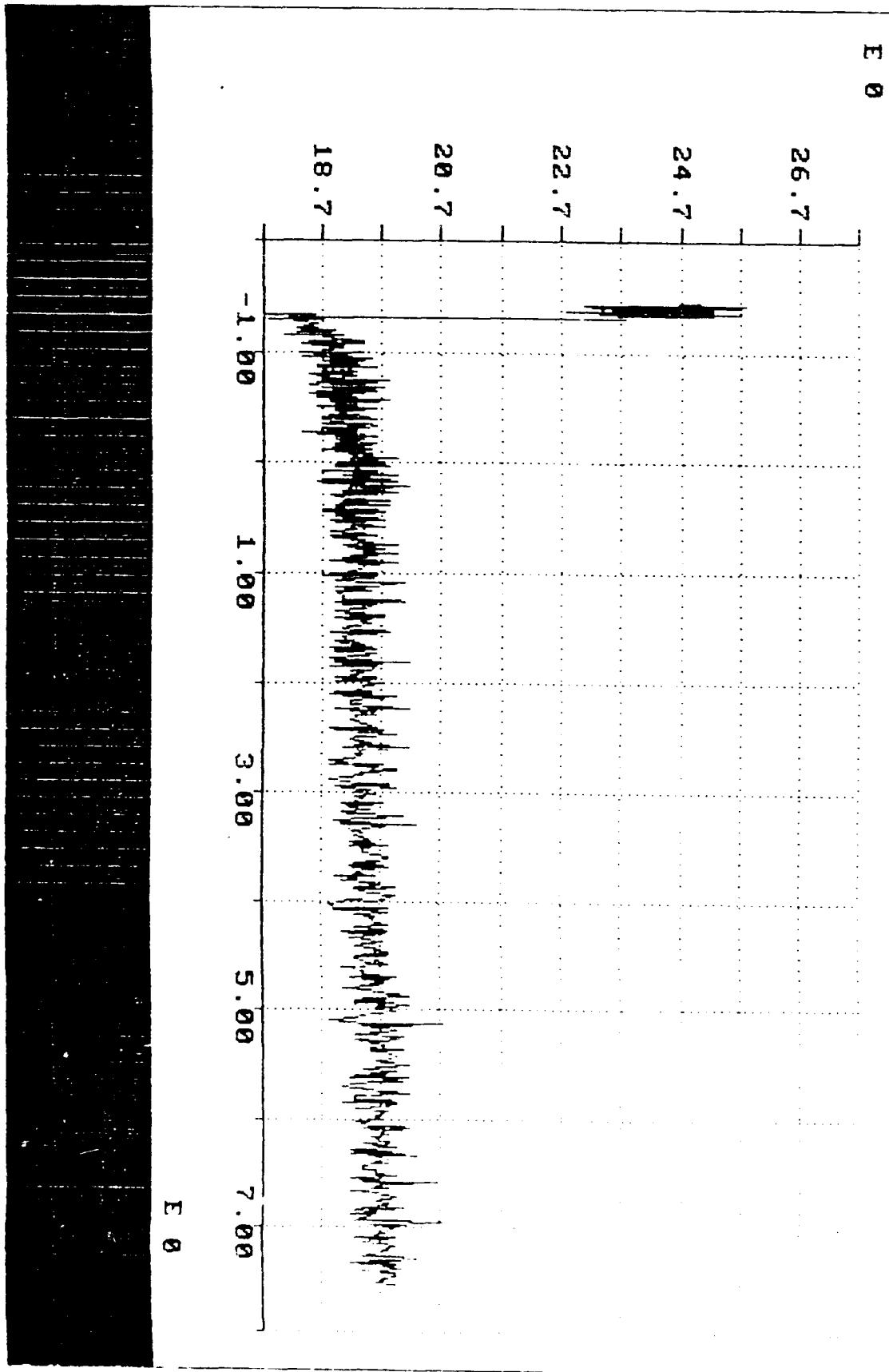
File Name: <B:AUG280.USR>

Quit/Continue: *

8 Comments

Subfiles (Total #: 8)

- 1> FERRONYL, TEMP CONTROLLED AT 60 DEGREES : Start# Shape #Repts :
2> BURLEIGH STAGE MAY NOT AHVE MOVED AT FIRS: 1 2 x 256 8 :
3> WELD DID NOT LOOK VERY STRONG :
4>



AUG 280

Ferronyl temp = 60

File Input/Output

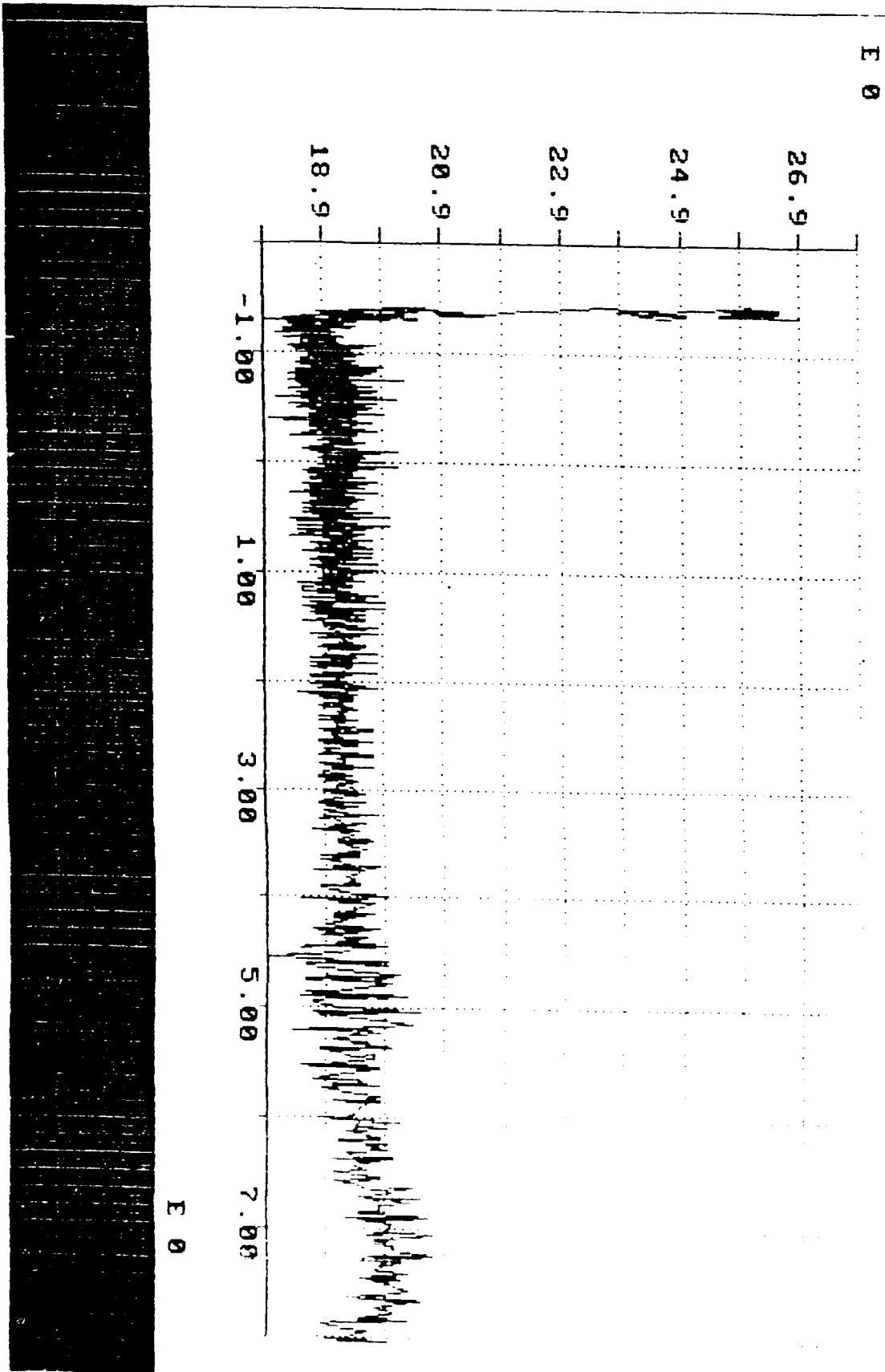
-File Name: <B:AUG28P.USR>

Quit/Continue: *

8 Comments

Subfiles (Total #: 8)

1> FERRONYL 70 DEGREES LITTLE DESSICATION AN: Start# Shape #Repts :
2> A WELD, STRONGER THAN 60 DEGRES, BUT NOT : 1 2 x 256 8 :
3> FERRONYL BARELY VISIBLE : : :



enough

$$\text{Temp} = 70^\circ$$

File Name: <B:AUG28E.USR>

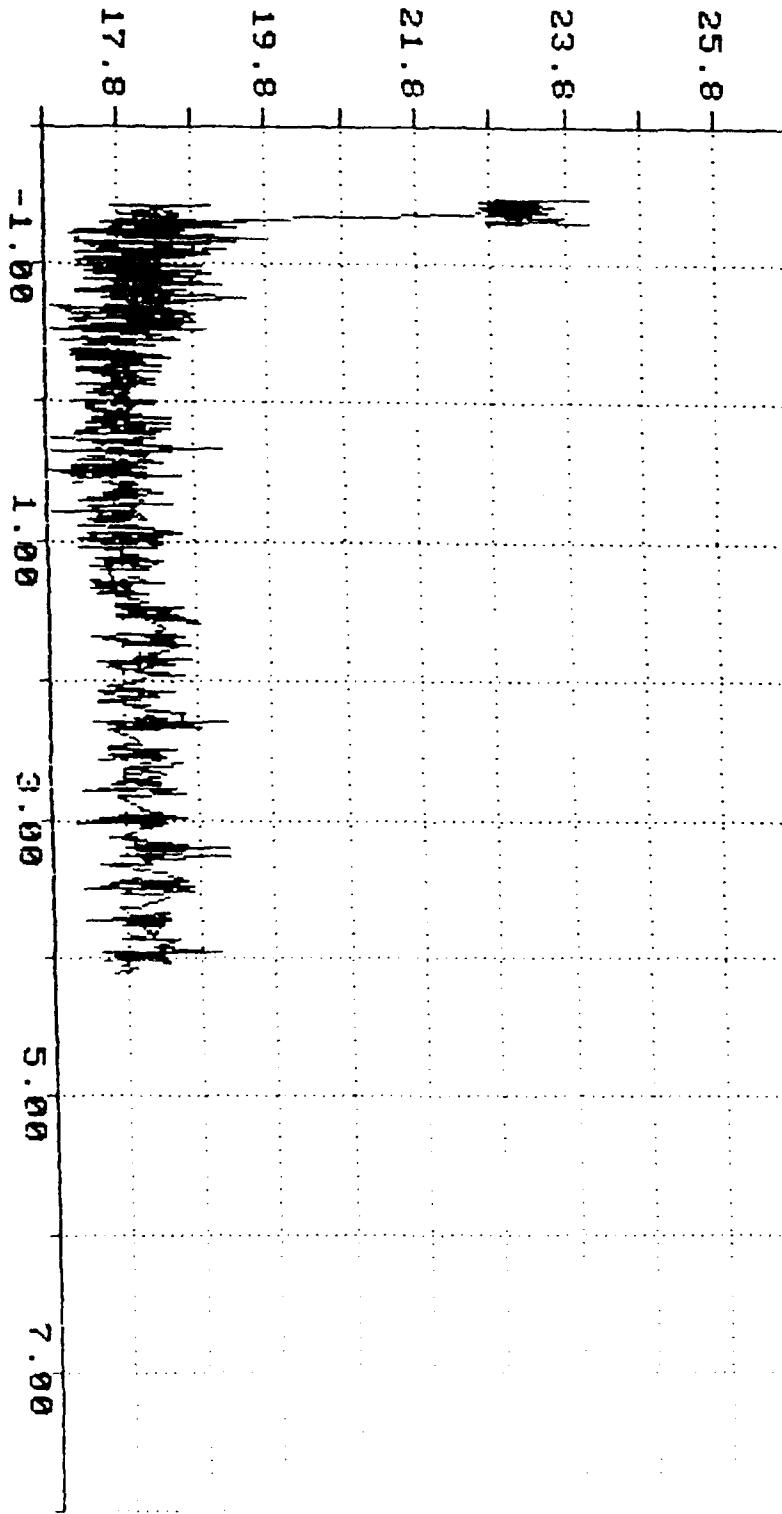
Quit/Continue: * : _____

B Comments

- 1> FERRONYL COMPOUND
- 2> 18 SECONDS
- 3> SMOKED
- 4> WELDED OR STUCK TOGETHER

Subfiles (Total #: 5) :

: Start#	Shape	#Repts :
: 1	2 x 256	5



AUG 28 E

Fervor

XXXXXXXXXXXXXXXXXXXXXXXXXXXX File Input/Output XXXXXXXXXXXXXXXXXXXXXXX

File Name: <B:AUG28F.USR>

Quit/Continue: *

8 Comments

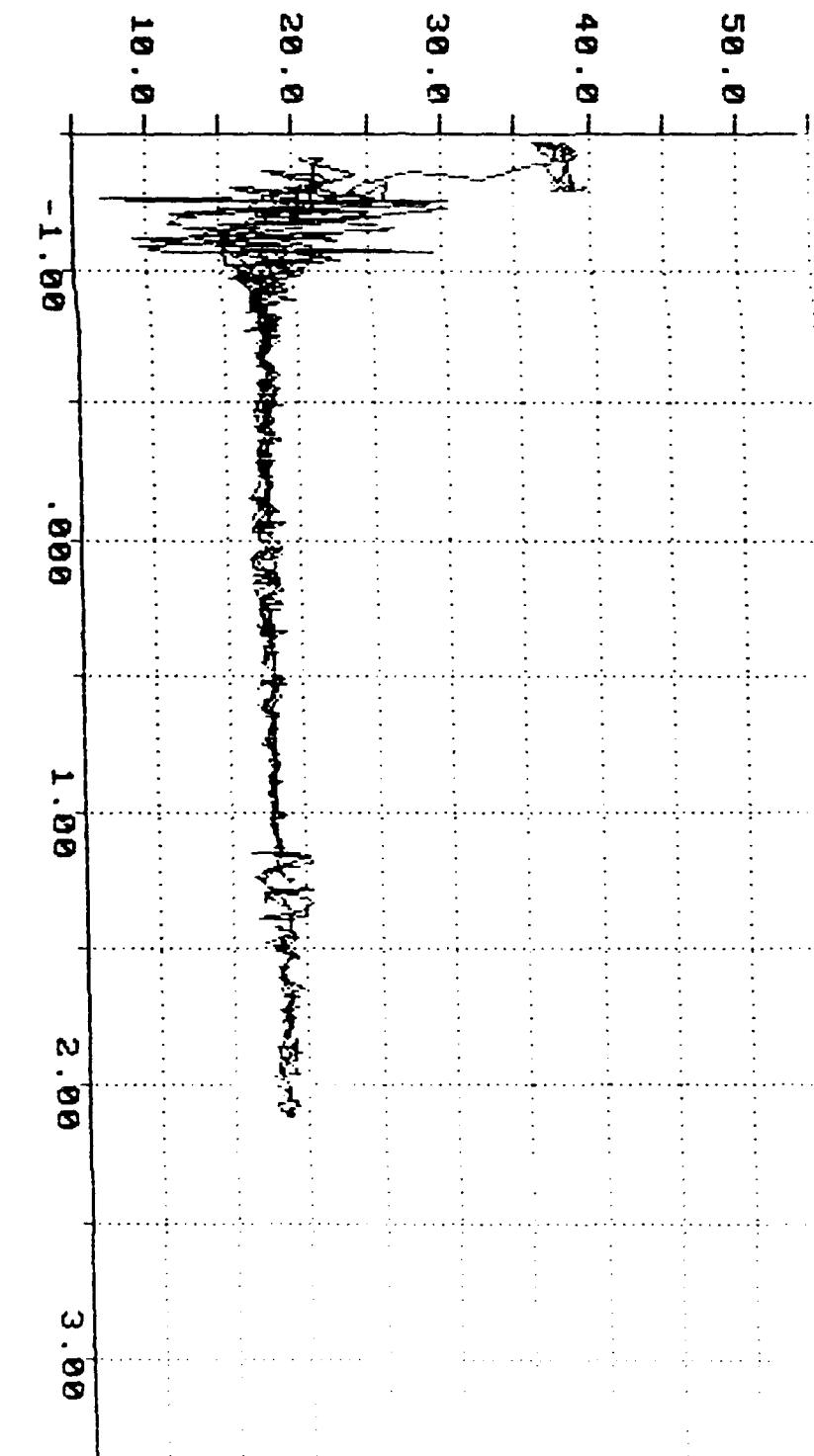
Subfiles (Total #: 4)

1> LIGHT INDIA INK, DRIED TISSUE, CONTROLLED: Start# Shape #Repts :
2> NOT MUCH OF A WELD : 1 2 x 256 4 :

E 0

AUG 28 F

Light India Ink
dried tissue



File Name: <B:AUG28G.USR>

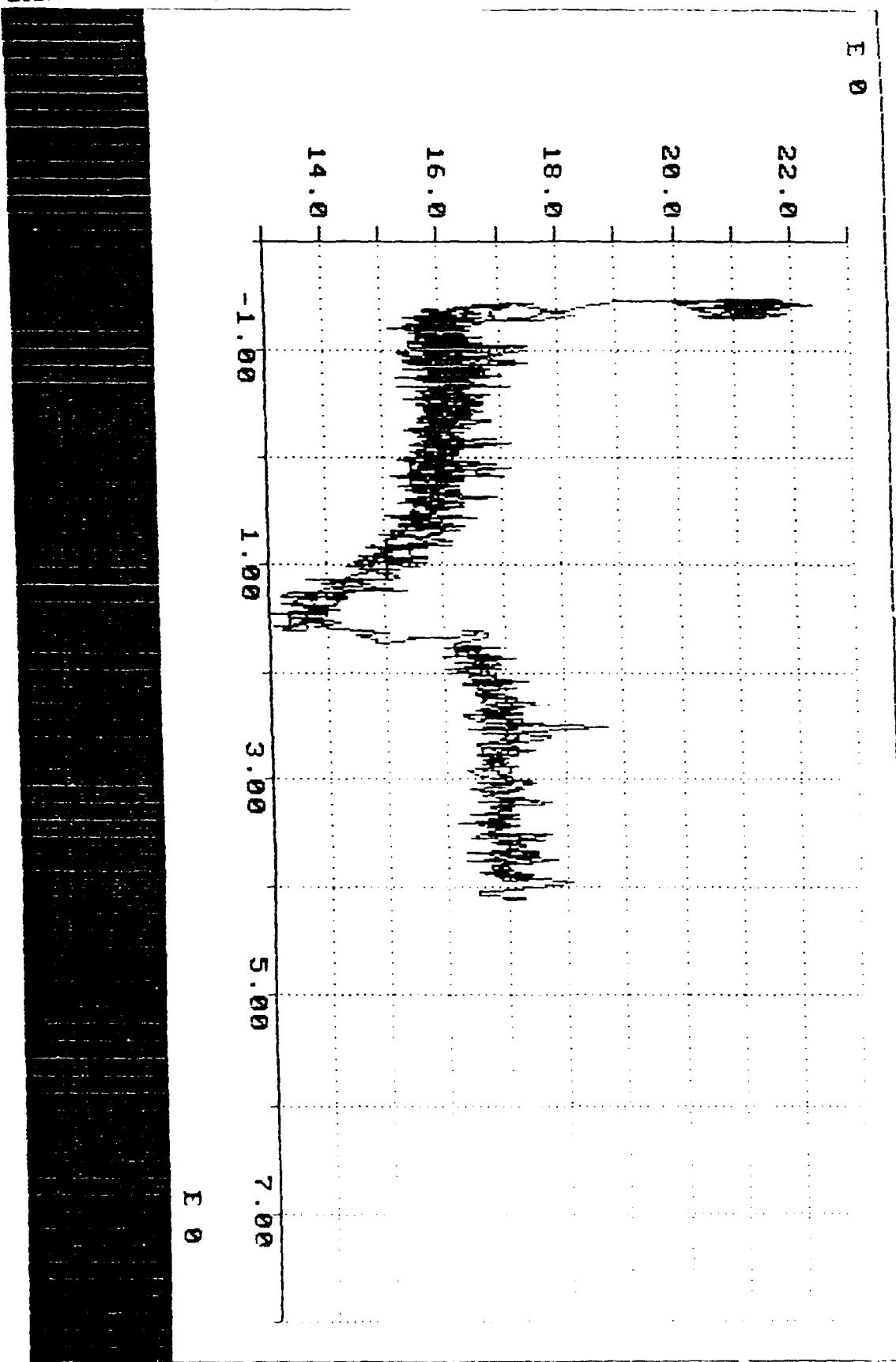
Quit/Continue: *

8 Comments

Subfiles (Total #: 6)

#Repts ::

: 1> LIGHT INDIA INK, DRIED WITH Q TIP, NEW BO: Start# Shape #Repts :



AUG 28G

high
medium
low

new pine
no camp

File Input/Output

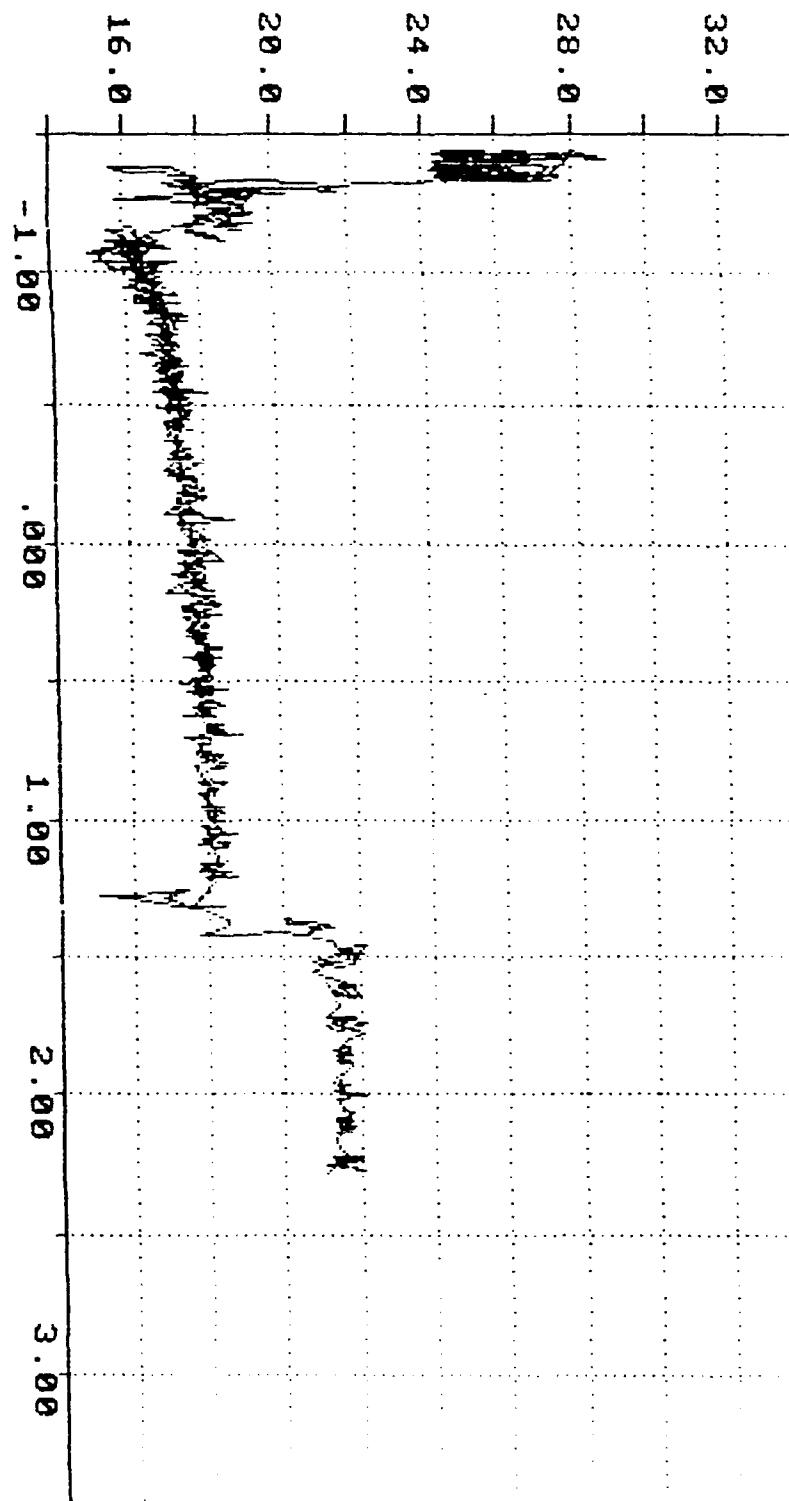
File Name: <B:AUG28H.USR>

Quit/Continue: *

8 Comments

Subfiles (Total #: 5)

- 1> NEW BOWEL WITH CLAMPS, LIGHT INDIA INK, D: Start# Shape #Repts :
2> THERMOCOUPLE PLACEMENT NOT CORRECT-ALIGNE: 1 2 x 256 5
3> CLAMPS ARE INTERFERING WITH EXPT. :



AUG 28 11

نیز

New bowed
with clamp

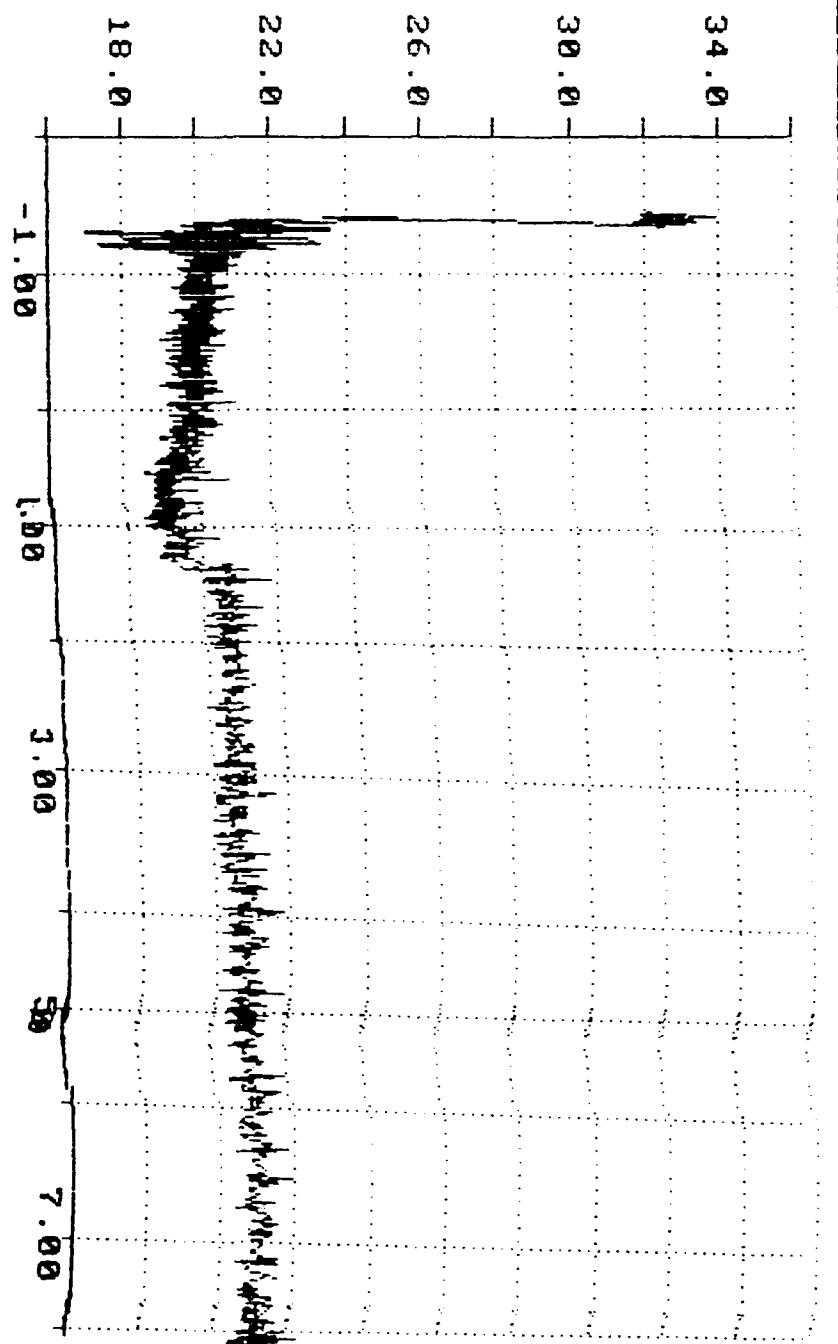
File Name: <B:AUG28I.USR>

Quit/Continue: *

8 Comments

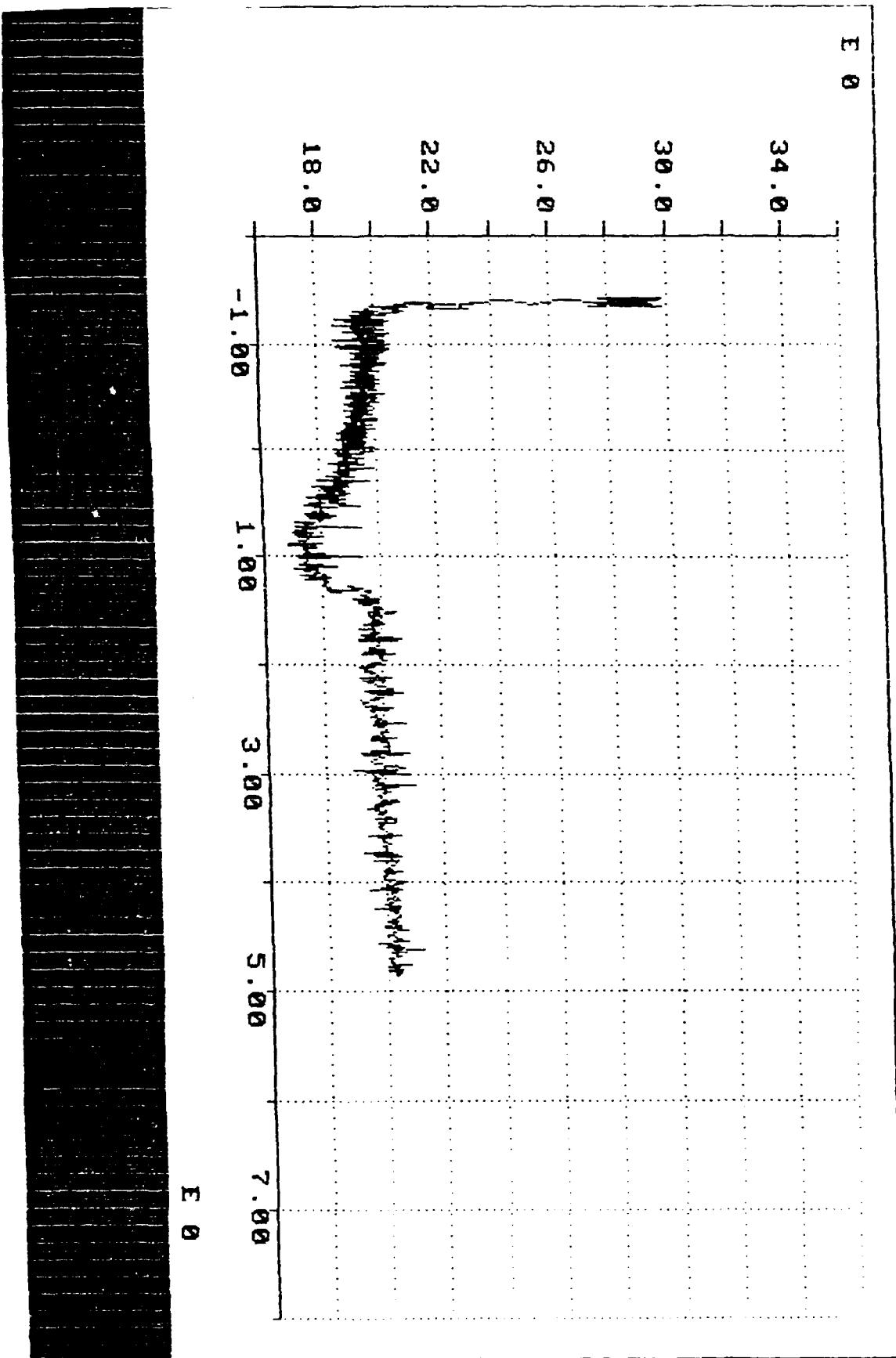
Subfiles (Total #: 8) :

: 1> TEMP AT 50 DEGREES, LIGHT INDIA INK, NO C: Start# Shape #Repts :
: 2> WELDED : 1 2 x 256 8



Dried in
Temp = 50

IMMMMMMMMMMMMMMMMMMMMMMM File Input/Output :
File Name: <B:AUG28J.USR> Quit/Continue: *
8 Comments Subfiles (Total #: 6) :
LMMMKMMMMMMMMMMMMMMMMMMMMMMMMMMMMMMMMMM
1> INDIA INK, TEMP = 60 DEGREES, SLIGHT DESS: Start# Shape #Repts :
2> LOOKS LIKE A WELD, TORE AT SEROSAL SURFCE: 1 2 x 256 6 :
AUC 285



India Ink

Temp=60

1oooooooooooooo File Input/Output oooooooo1ooooooo1ooooooo;

File Name: <B:AUG28B.USR>

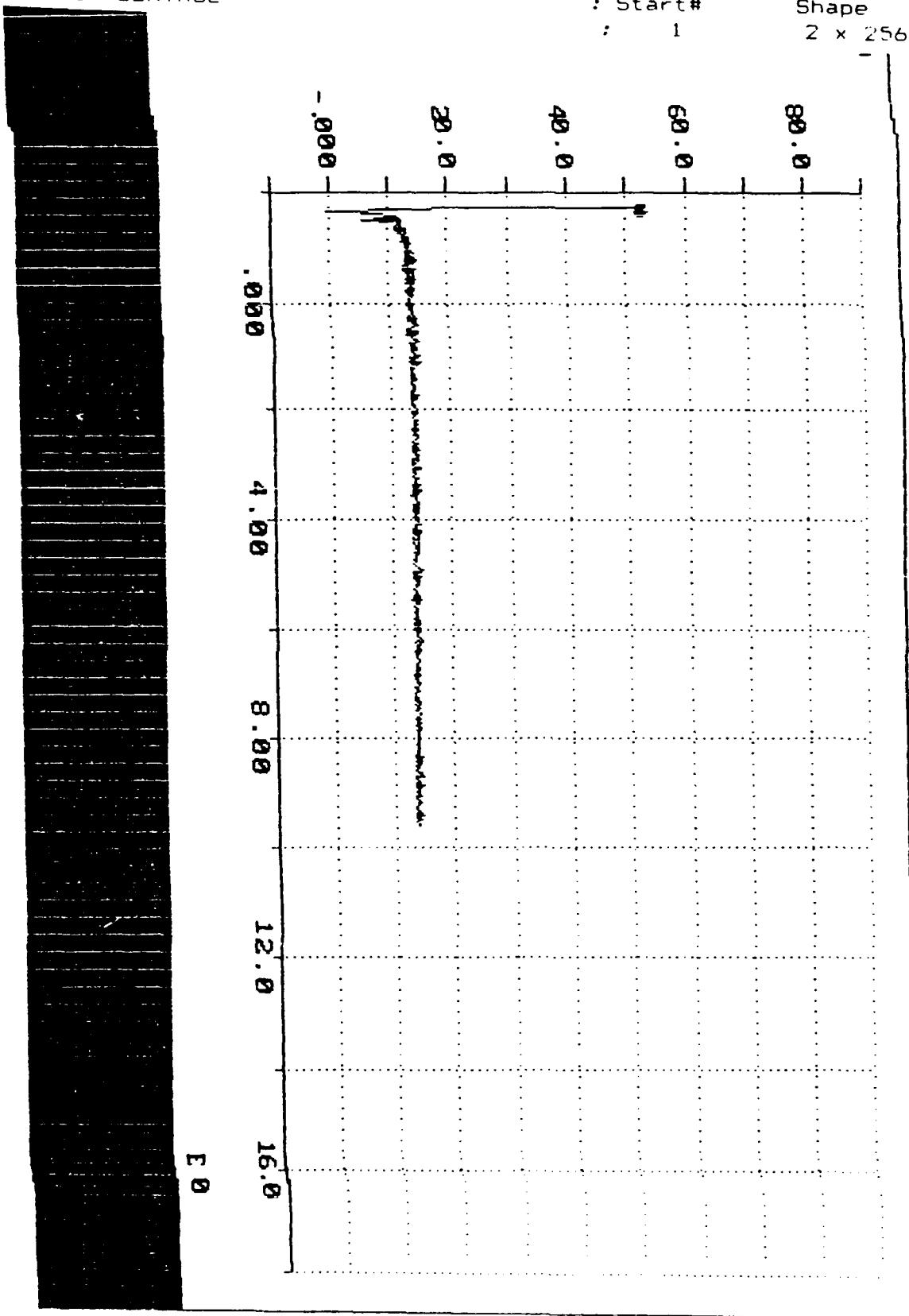
Quit/Continue: *

8 Comments

Subfiles (Total #: 5) :

> TWO STRIP CONTROL

: Start# : Shape : #Repts :
: 1 : 2 x 256 : 5 :



2 strip control

File Name: <B:AUG28C.USR>

Quit/Continue: *

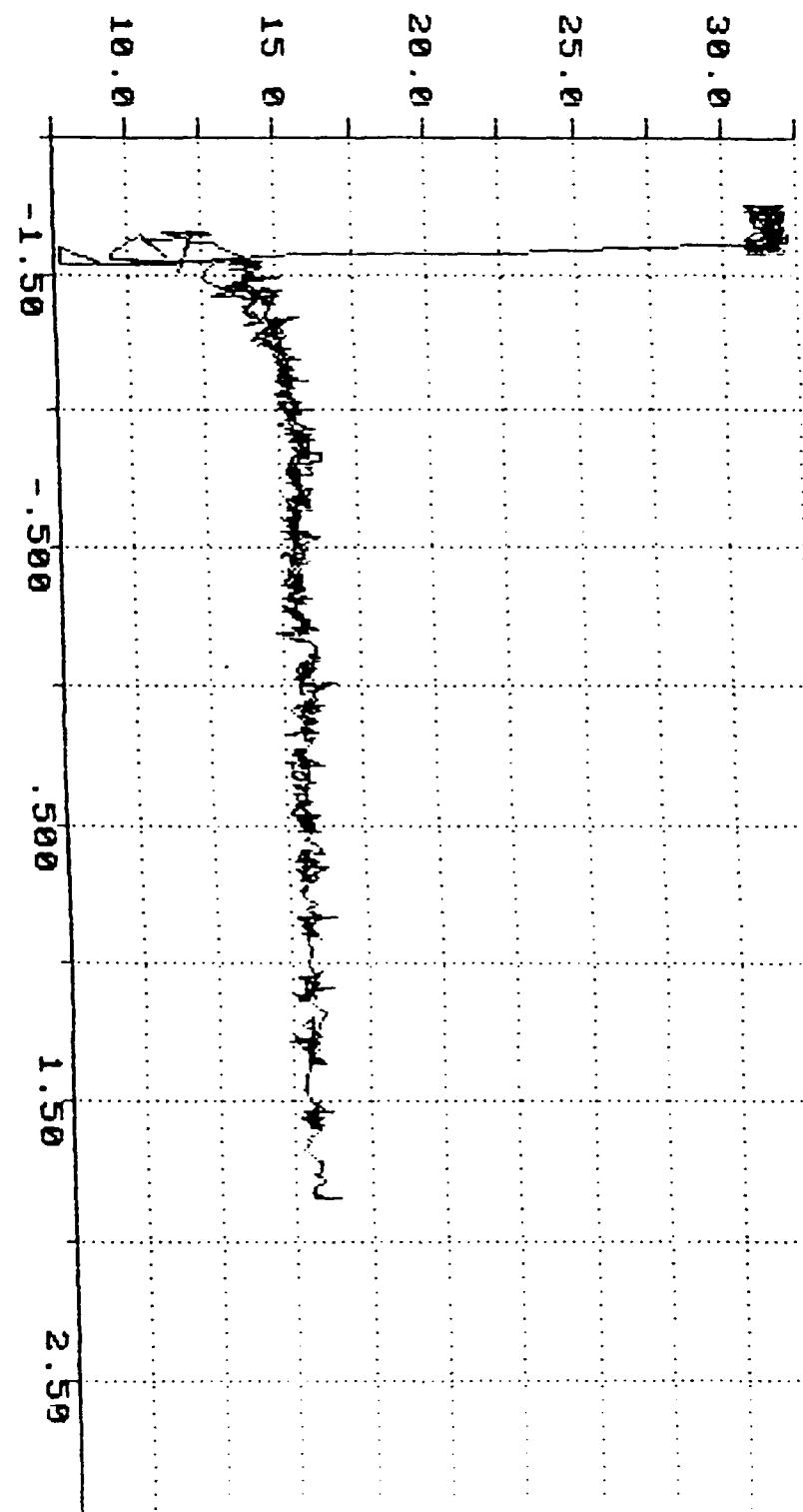
8 Comments

Subfiles (Total #: 4) :

LASER CONTROL

: Start# Shape #Repts :
2> DID NOT LOOK LIKE MUCH OF A WELD : 1 2 x 256 4 :

laser control - 2 steps



XXXXXXXXXXXXXXXXXXXX File Input/Output

File Name: <B:AUG28D.USR>

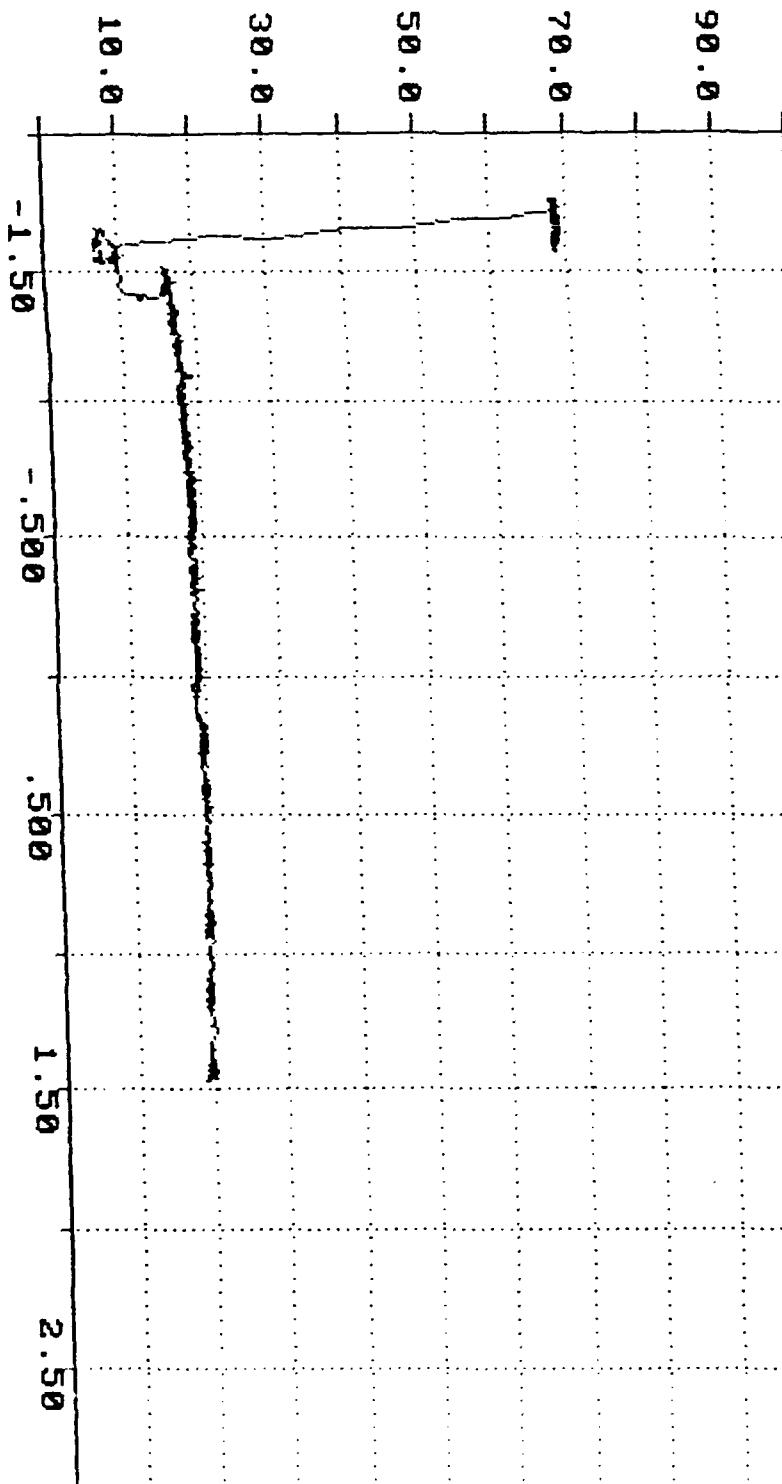
Quit/Continue: *

8 Comments

INDIA INK LIGHT

NOT A GOOD WELD

Subfiles (Total #: 4)

: Start#
: 1Shape
2 x 256#Repts :
4 :

Aug 28D

India ink (light)

4-5-90

E 9

7.00 5.00 3.00 1.00

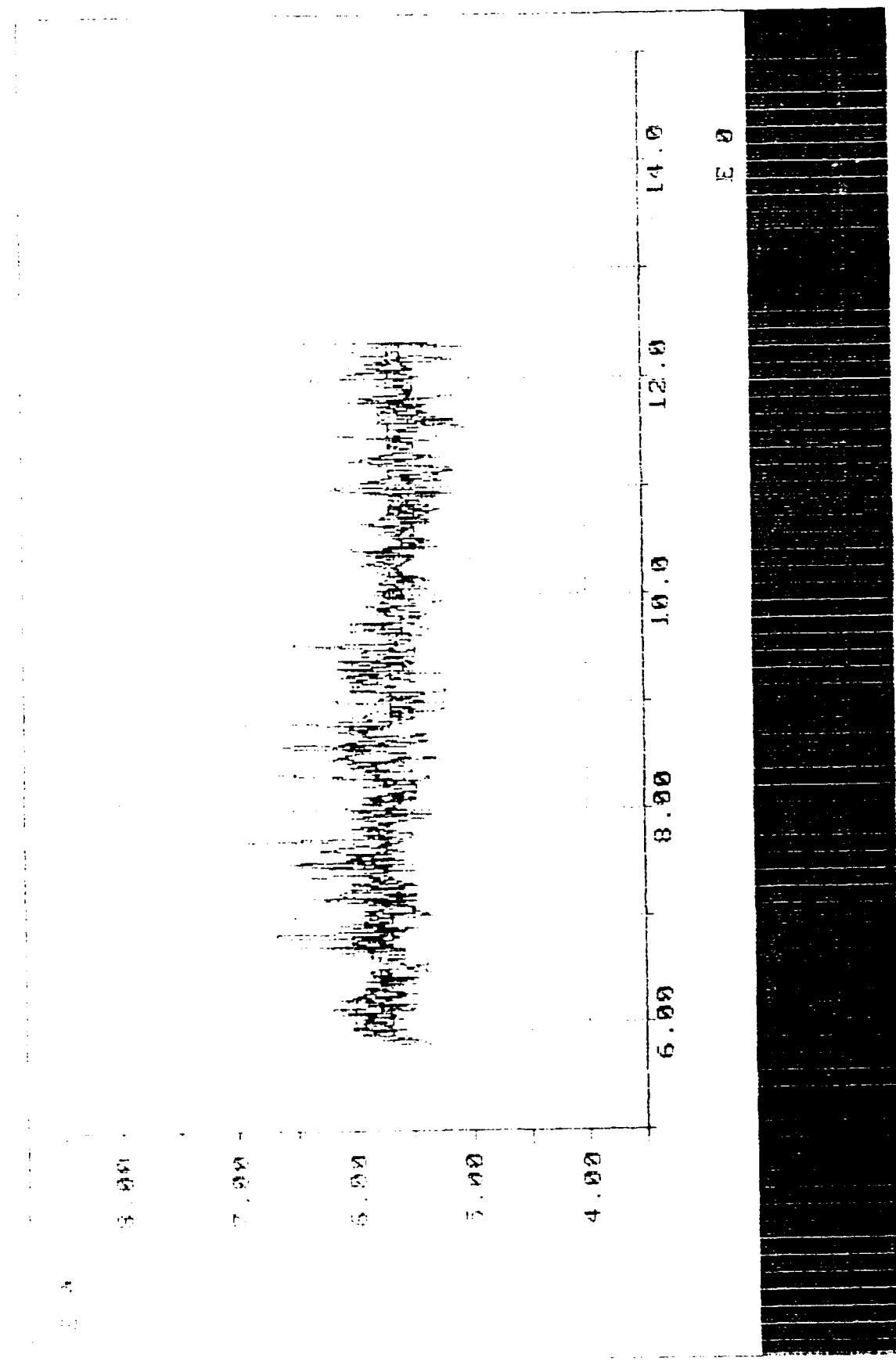
4.00

3.00

2.00

1.00

0.00



20

1.000 4.00 8.00 12.00 16.00

-2.00

-0.00

2.00

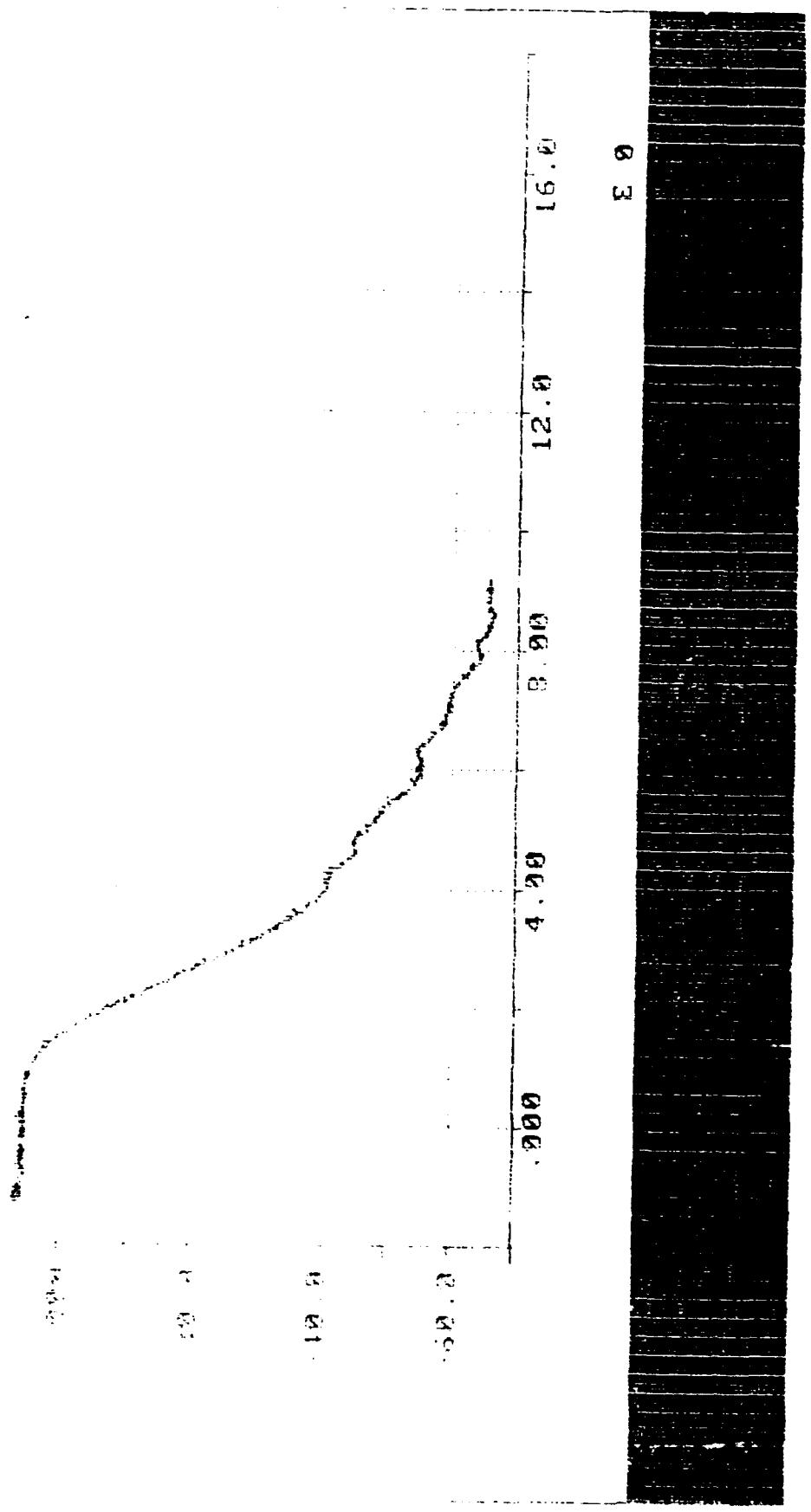
4.00

6.00

8.00

10.00

12.00



1000 = 1000000
1000000

200 0

200 0

-600 0

9.62 10.6 11.5 12.6 13.6

E 0

E 0

3.96
2.96
1.96
0.96

5.96

6.96

7.96

8.96

9.96

10

11.96

12.96

E 3

7.09

5.89

5.99

6.01

6.02

4.00

3.99

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-3.00

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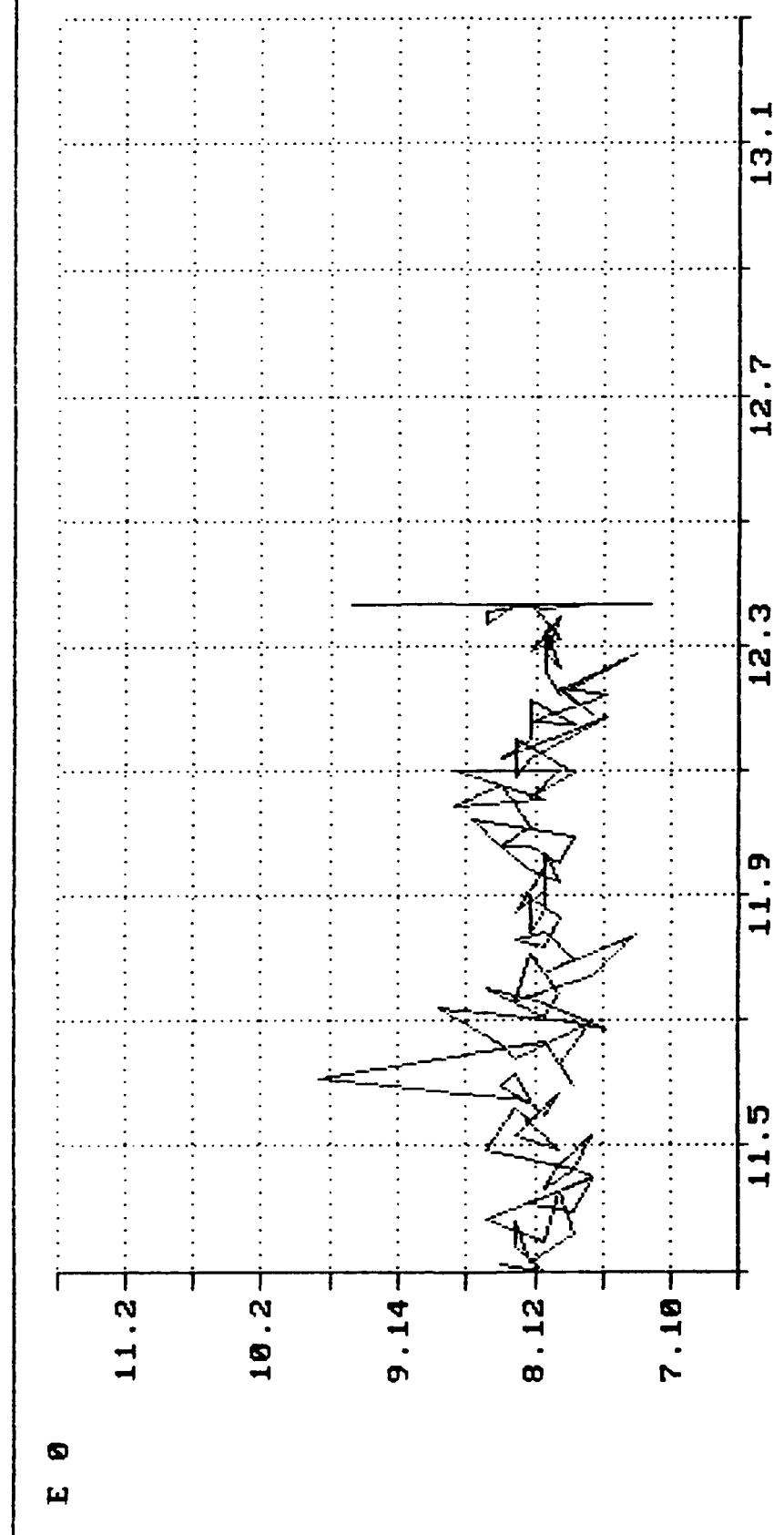
7.19

6.18

5.19

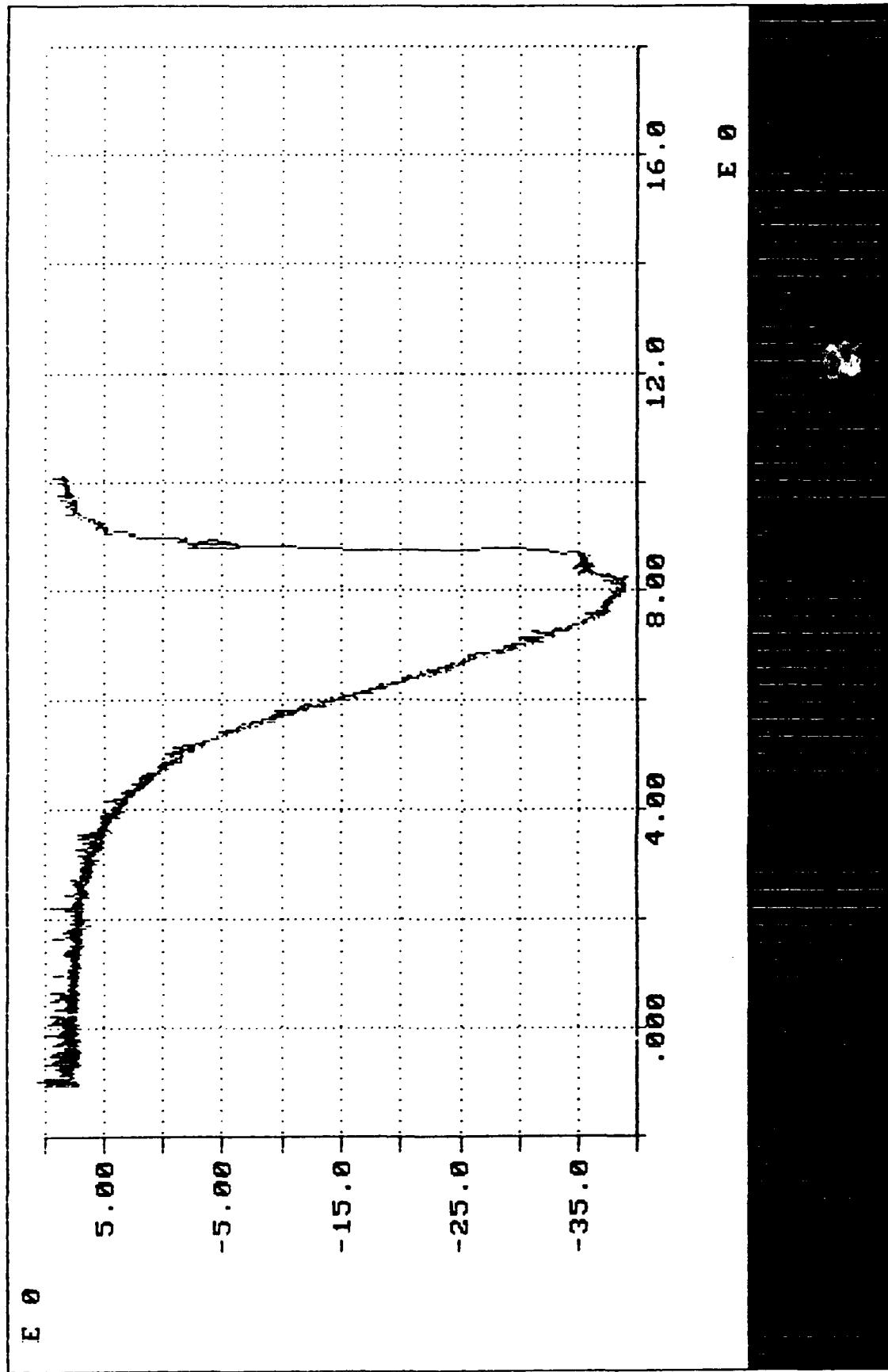


(2500 = 100)
WCF



E 0



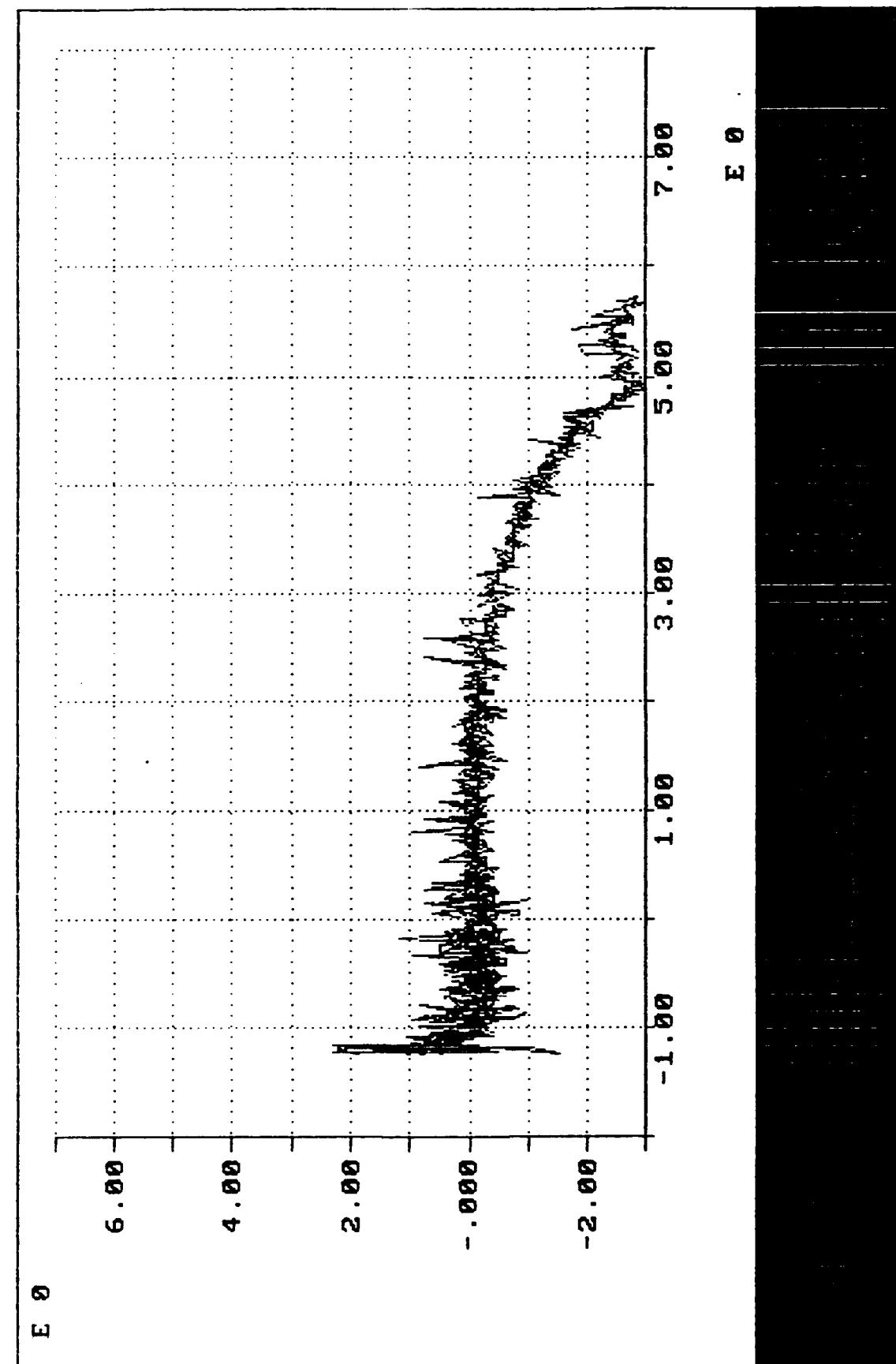


slimy

Sep 5 N
one snip unweeded
cold saline control

Baruch

Sep 50
India gun conducted at 50°
(main weld)



1. Burris

Arne Manning, classmate

$\overline{SS} = 55$

India ink set - 60

1525

E 0

5.00

3.00

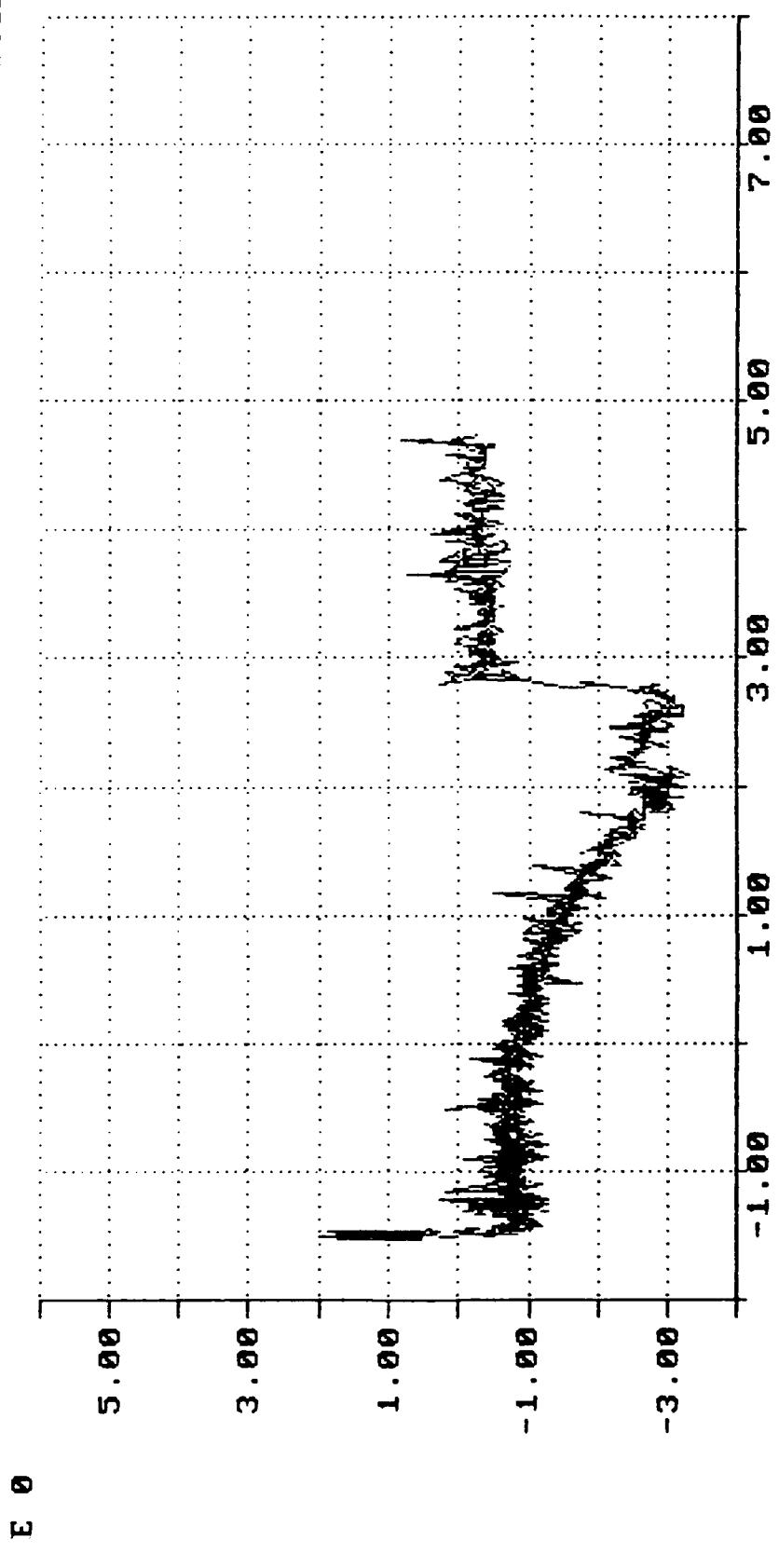
1.00

-1.00

-3.00

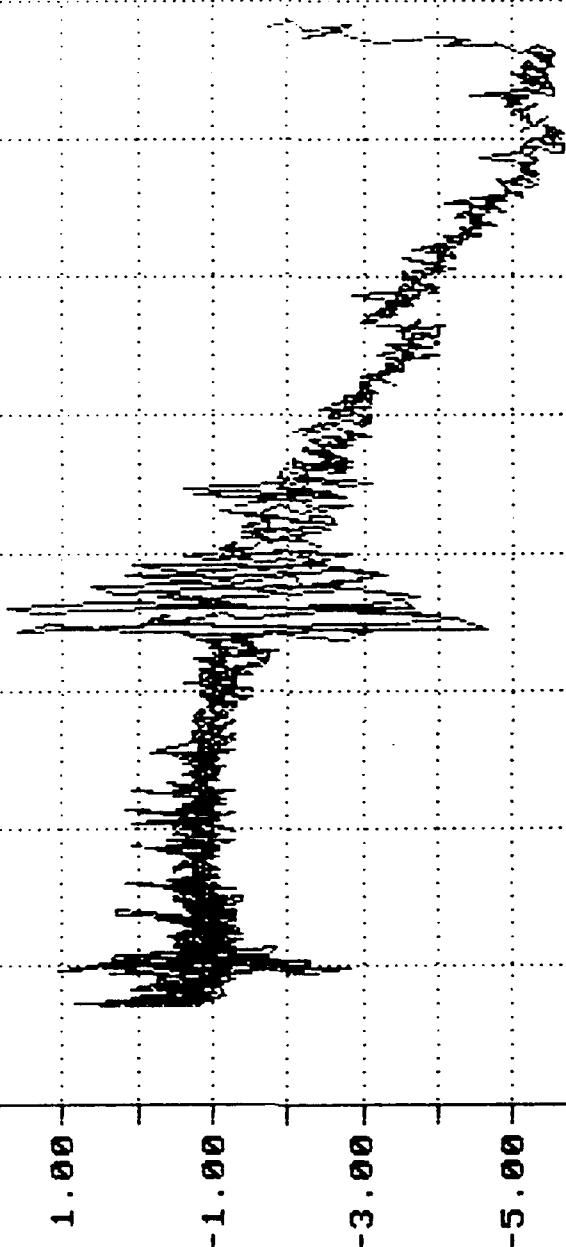
E 0

7.00
5.00
3.00
1.00
-1.00



E 9

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5.00
3.00
1.00
-1.00
-3.00
-5.00



E 9

SEPS 5
Indice de convolution

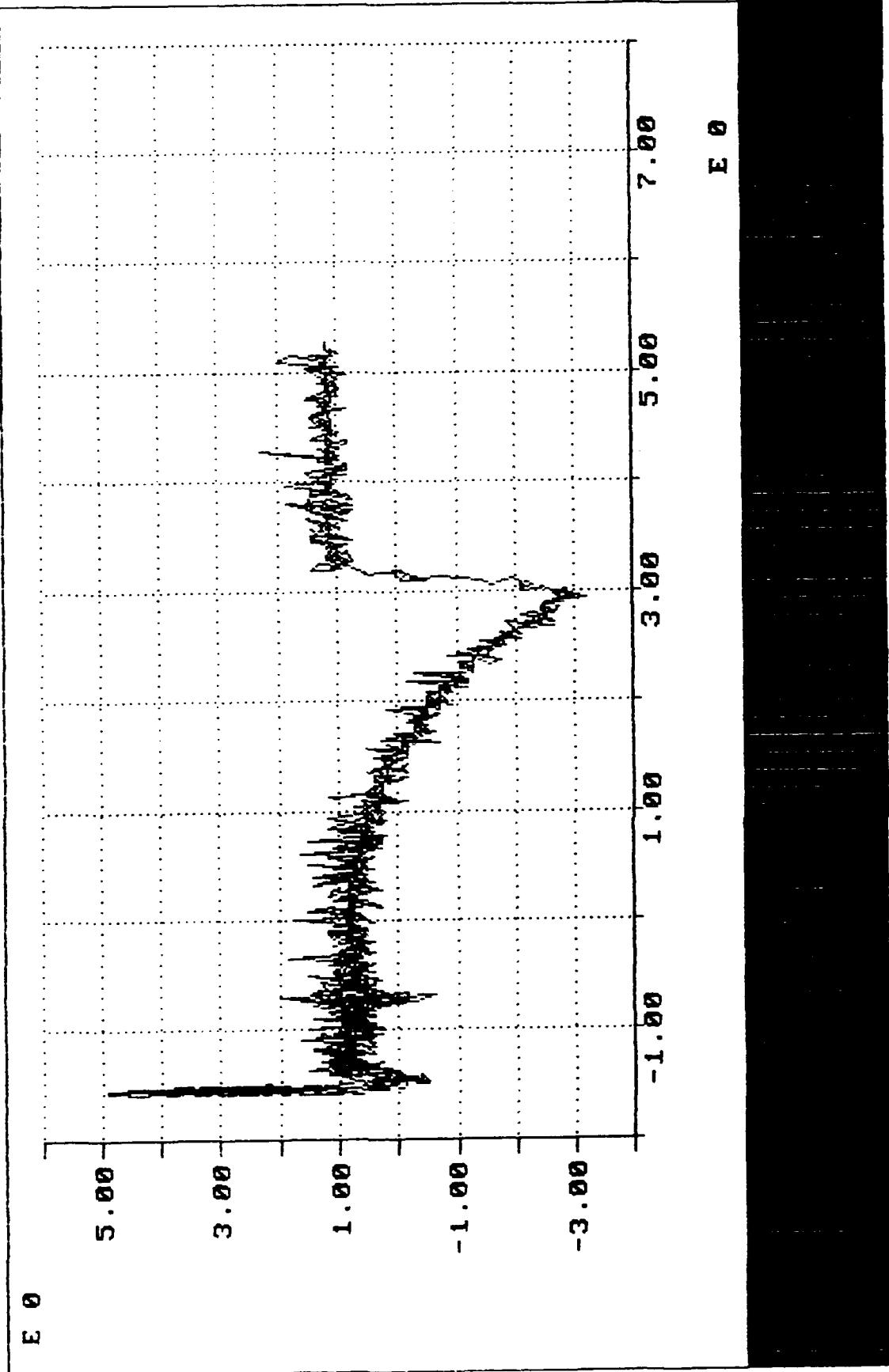
blandium, dessication
wooden pane
mean well

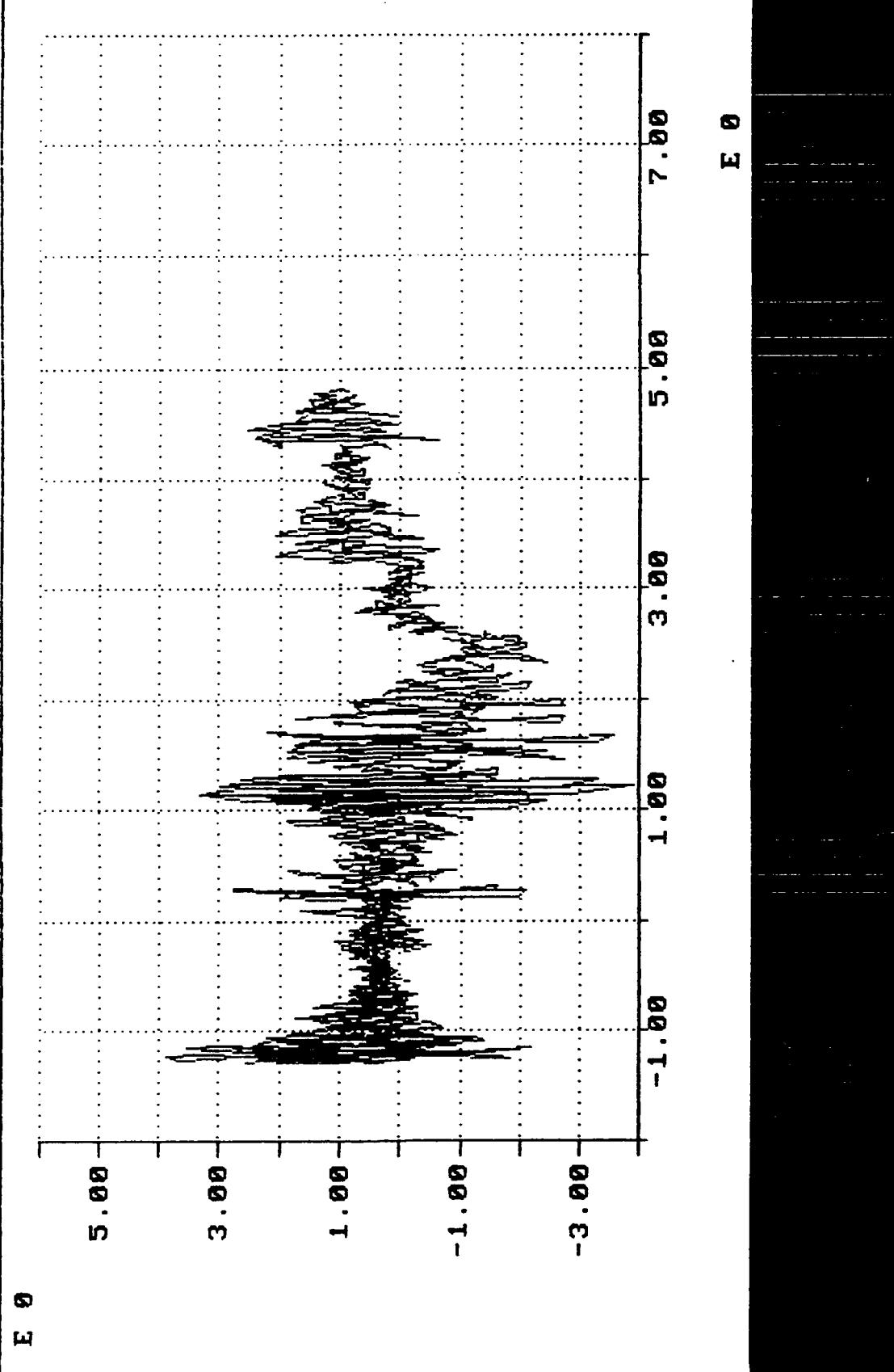
No. visible channels

TC max = 60

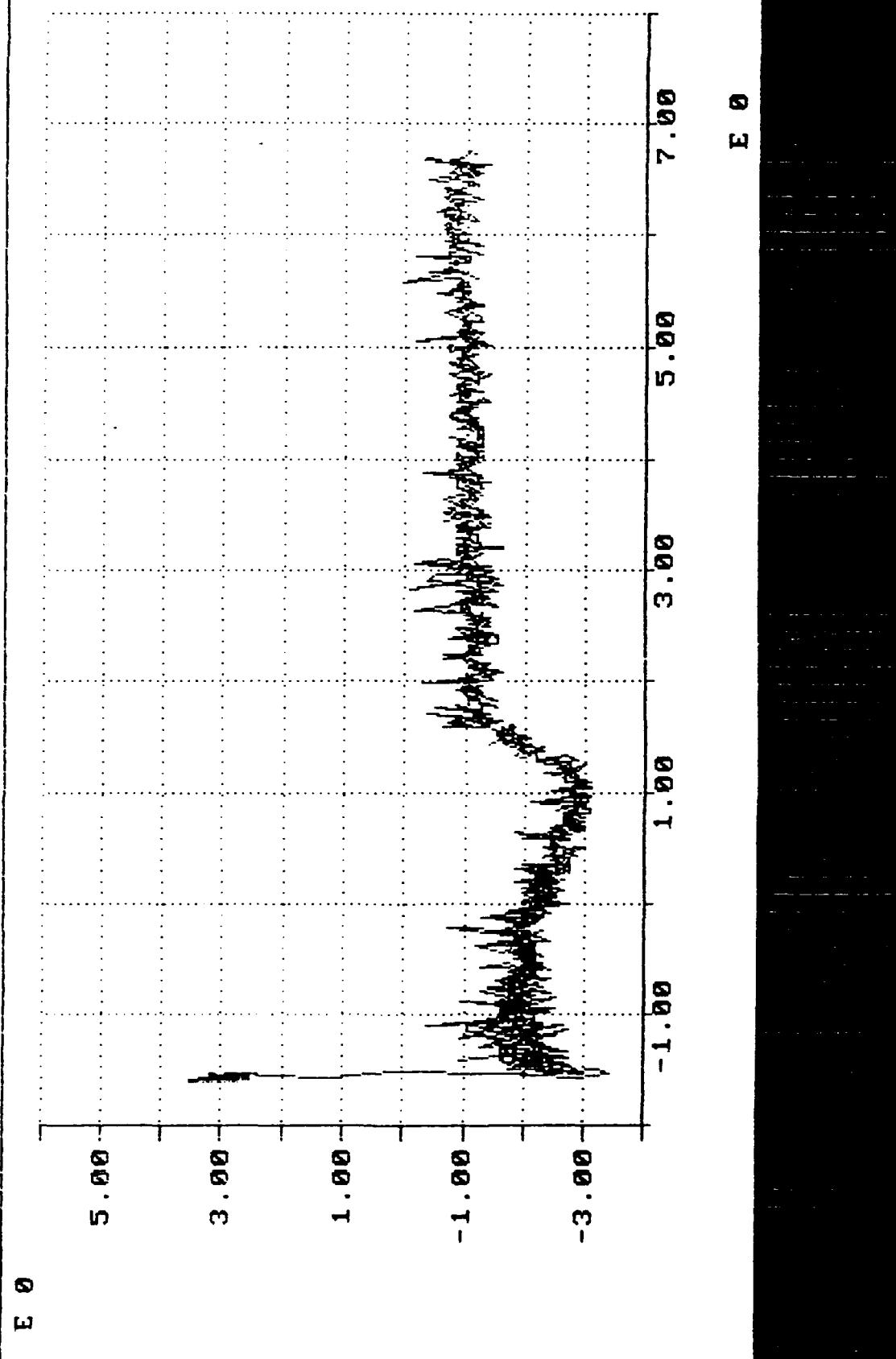
Detector gain set = 20³

SEPF5R



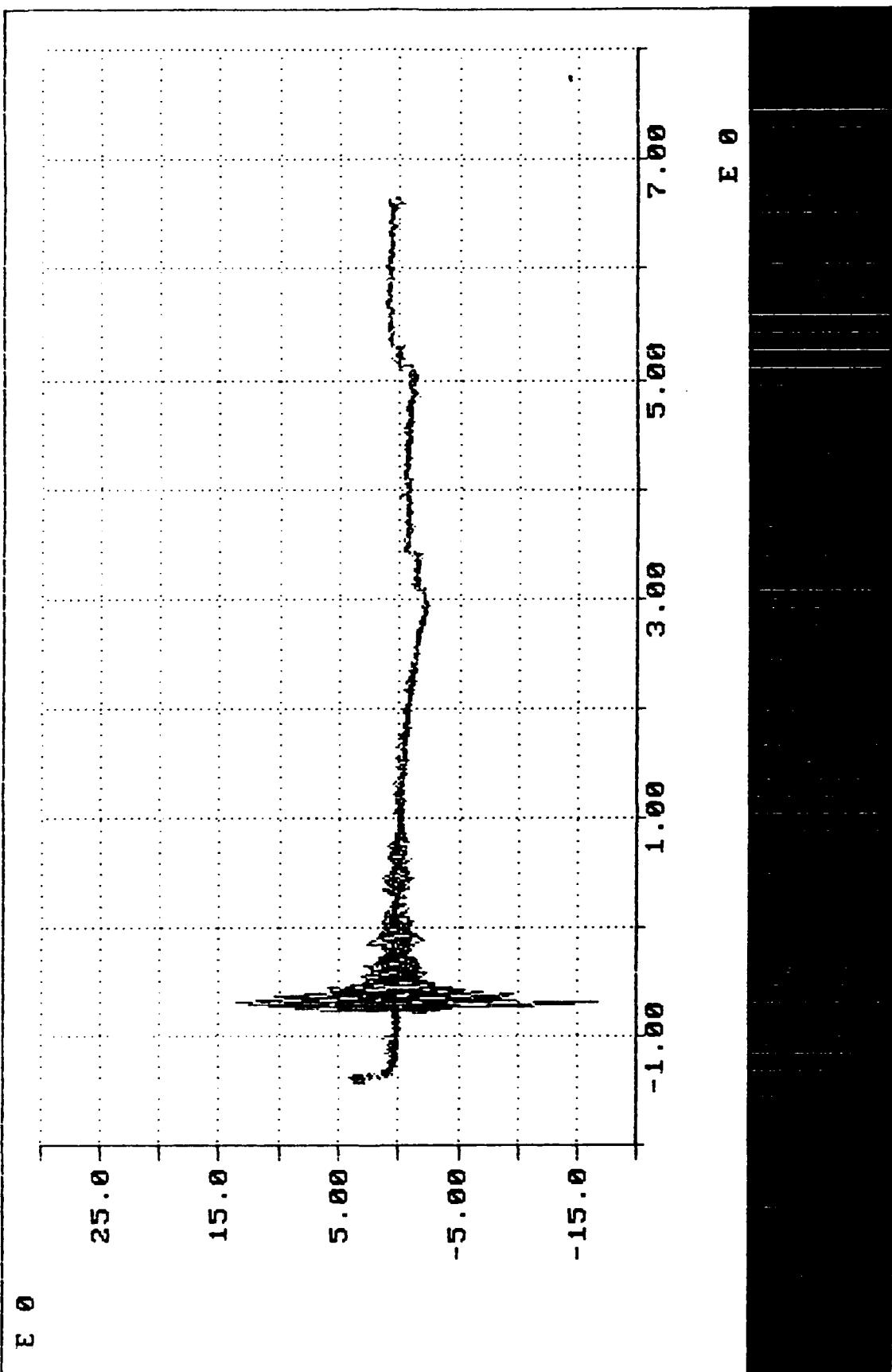


JEP 55
India rock controlled at 50°
No visible changes
very weak (well)



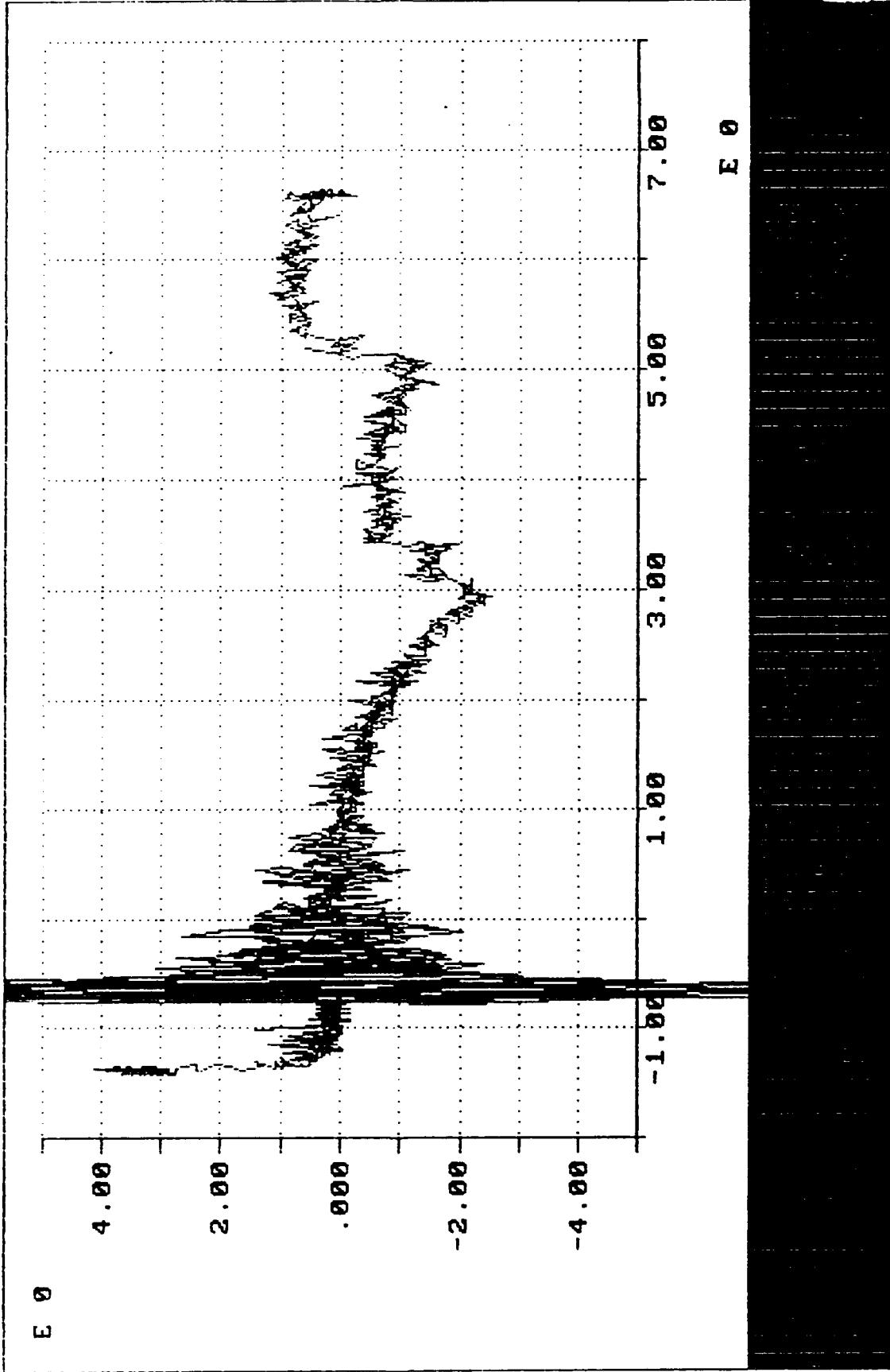
SEPSIT
Anais Only Set = 100°
 $T_c \max = 55^\circ$

Blanking, dissipation, smoke
weak weld
transient load welds

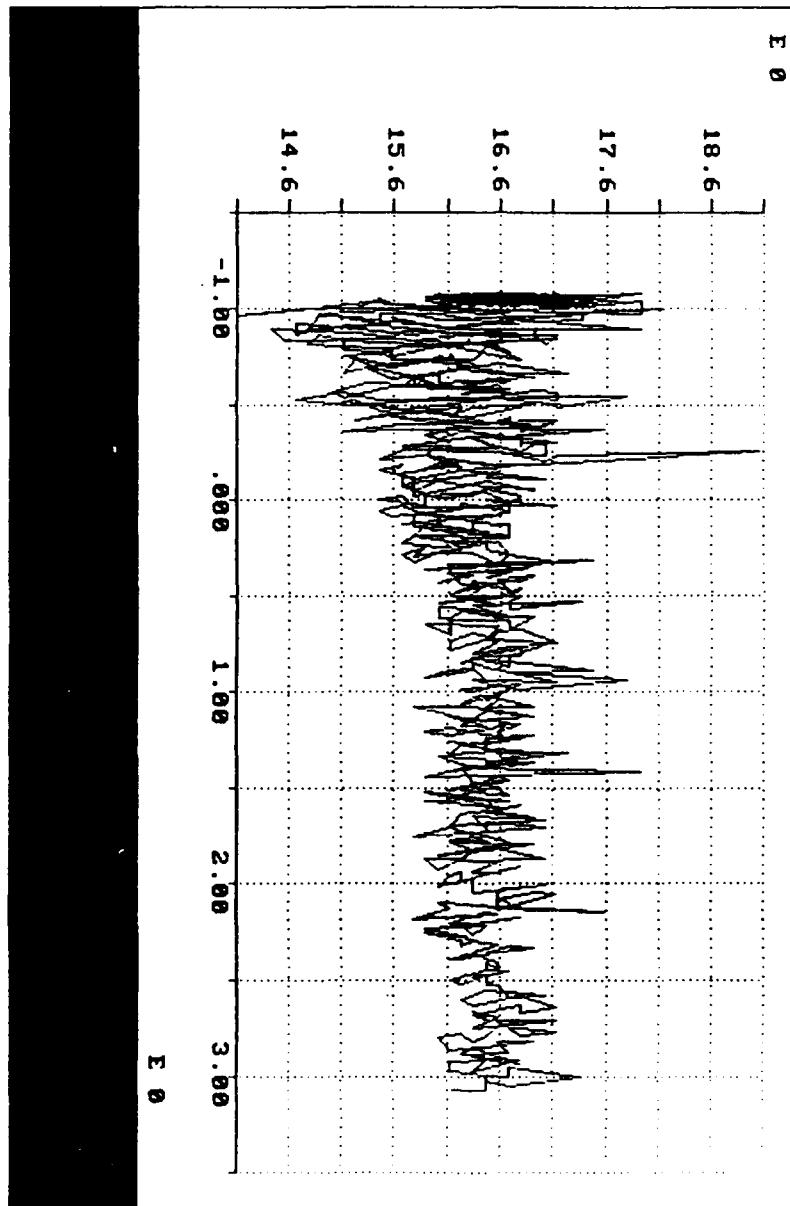


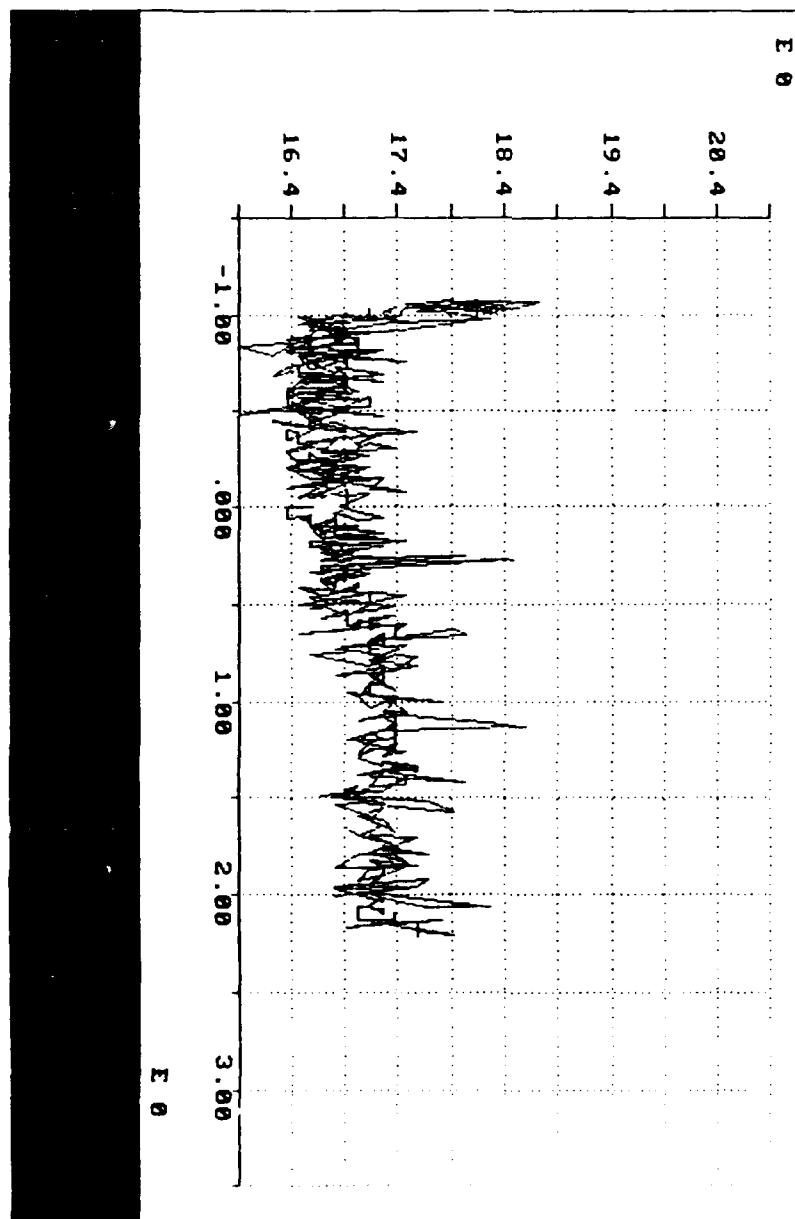
SEPSU
Dowco link
Controlled at 1000
Decimation rate 100x
wave used

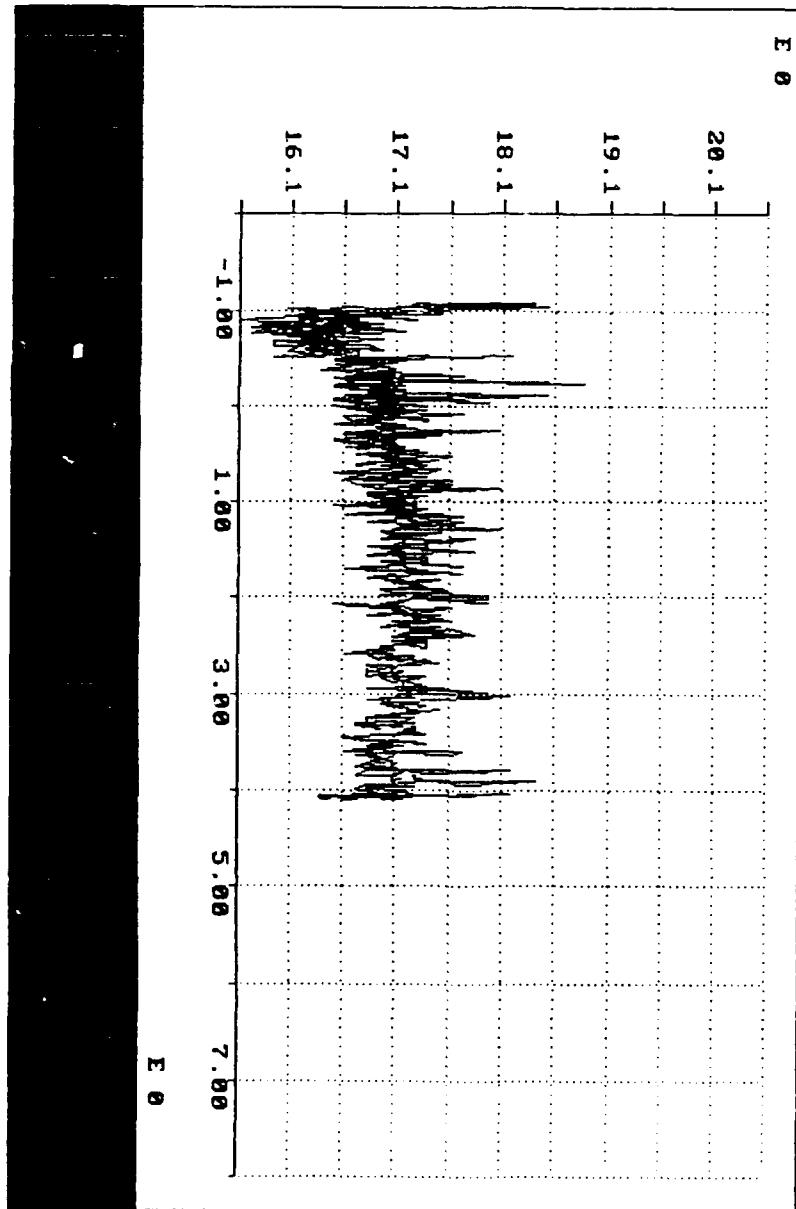
SEPSU
(replotted)

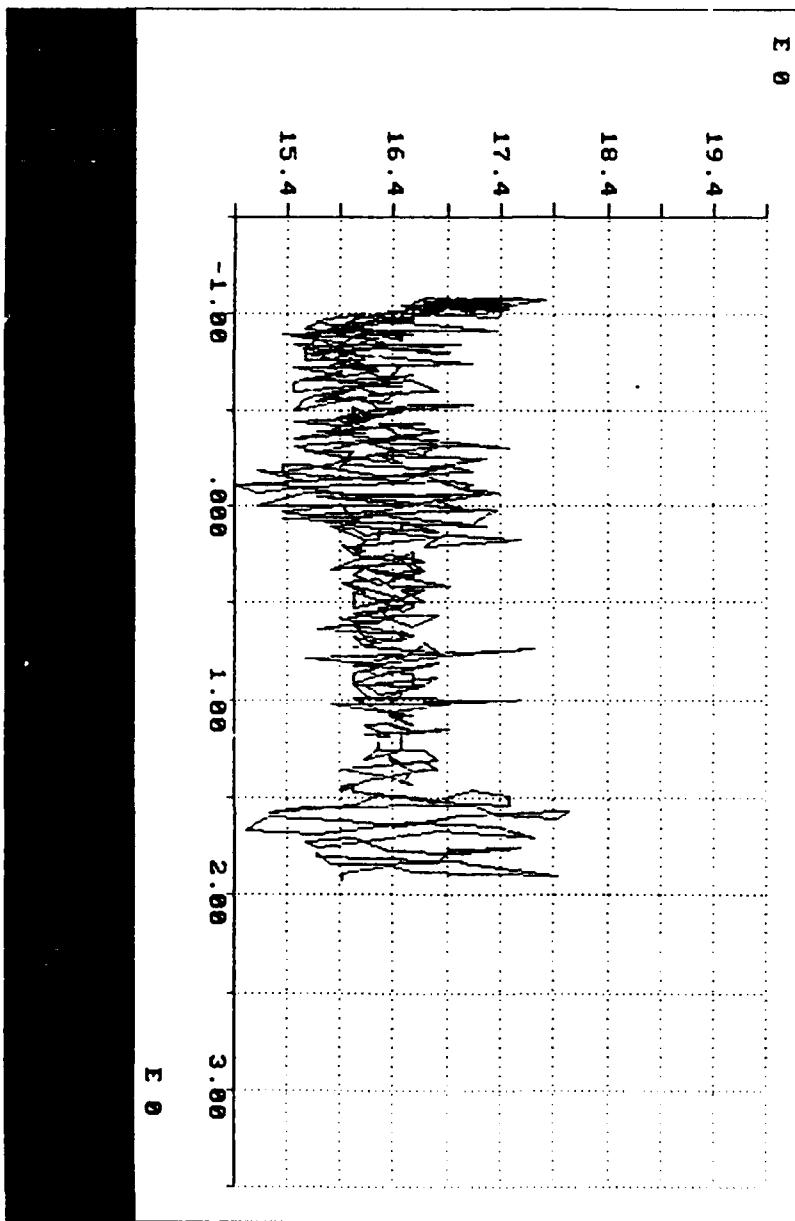


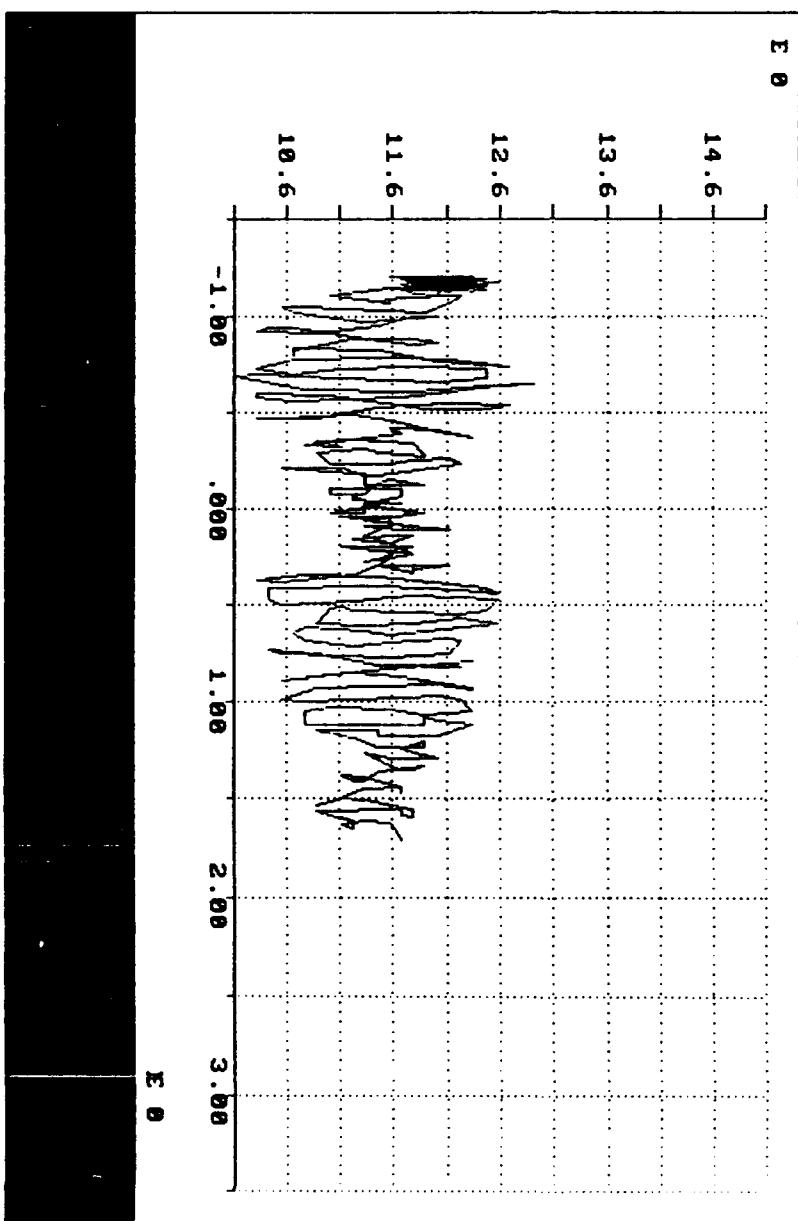
```
: File Name: <C:SEF7U.USR> Quit/Continue: * :  
:  
: 8 Comments Subfiles (Total #: 1) :  
: 1> INDIA INK : Start# 1 Shape 2 x 256 #Repts :  
: 2> CONTROLLED AT 60 :  
: 3> SLIGHT DESSICATION :  
: 4> NOT VERY IMPRESSIVE :  
: 5> :  
: 6> :  
: 7> :  
: 8> :  
: I/O Variable ( R - Z ): 1024 REAL :  
:  
: Subfile: < 1 > 2 x 256 (Max Length: 768 ) :  
: Row (0=all): < 0 > :  
: Start Column: < 1 > :  
: # of Columns: < 256 > :  
:  
: Read/Write/Append/Scroll/Plot/Edit/List/New file/Quit: *
```

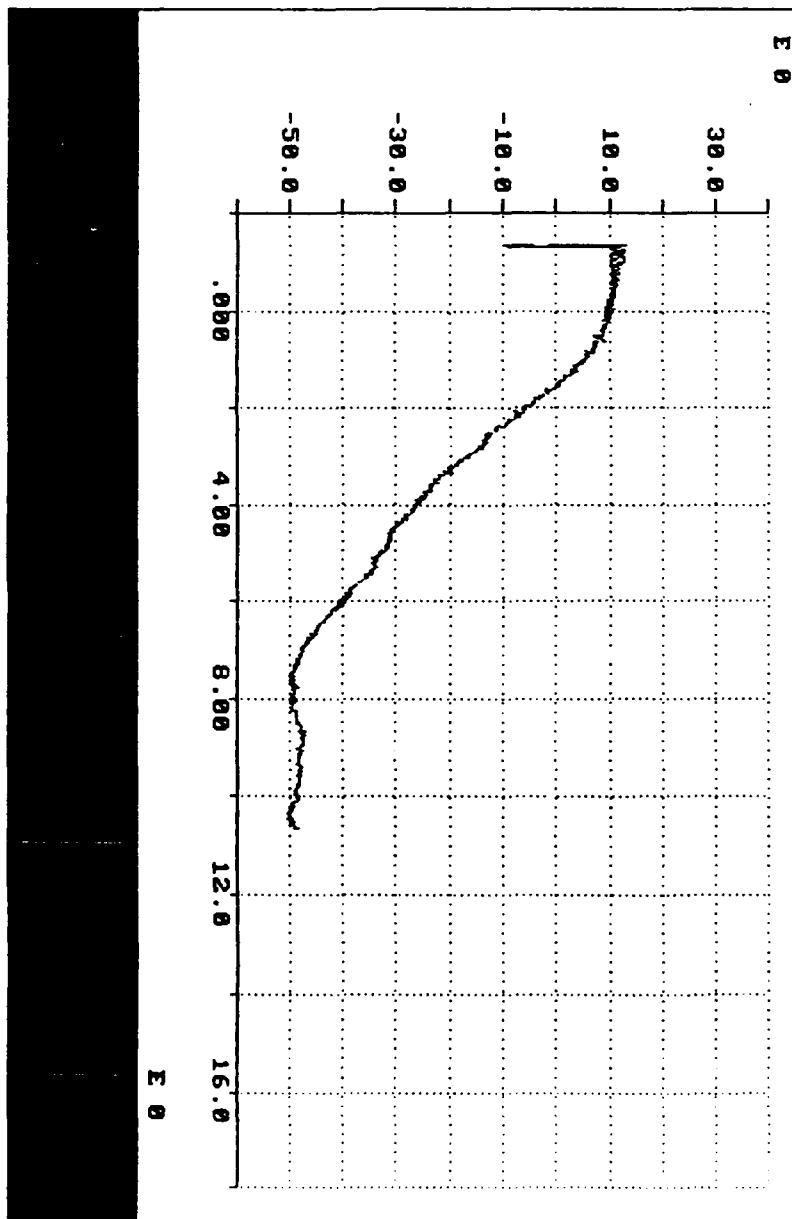


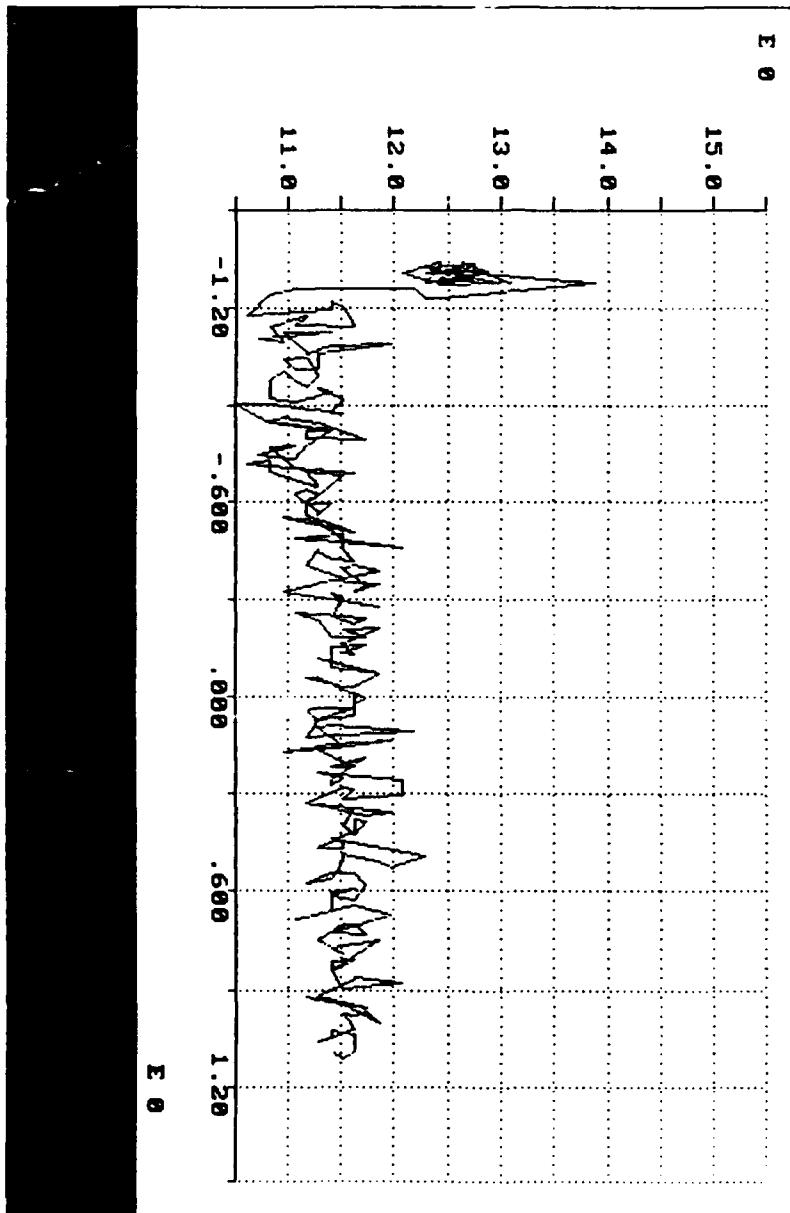


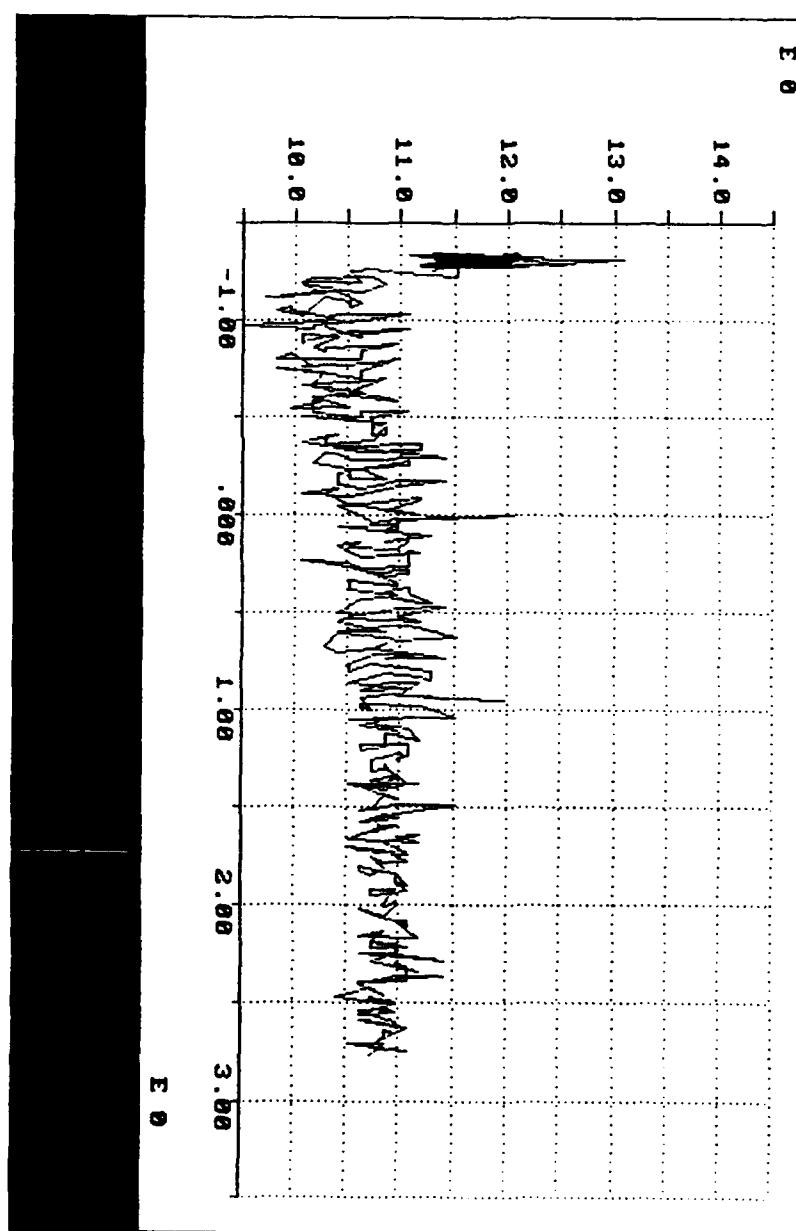


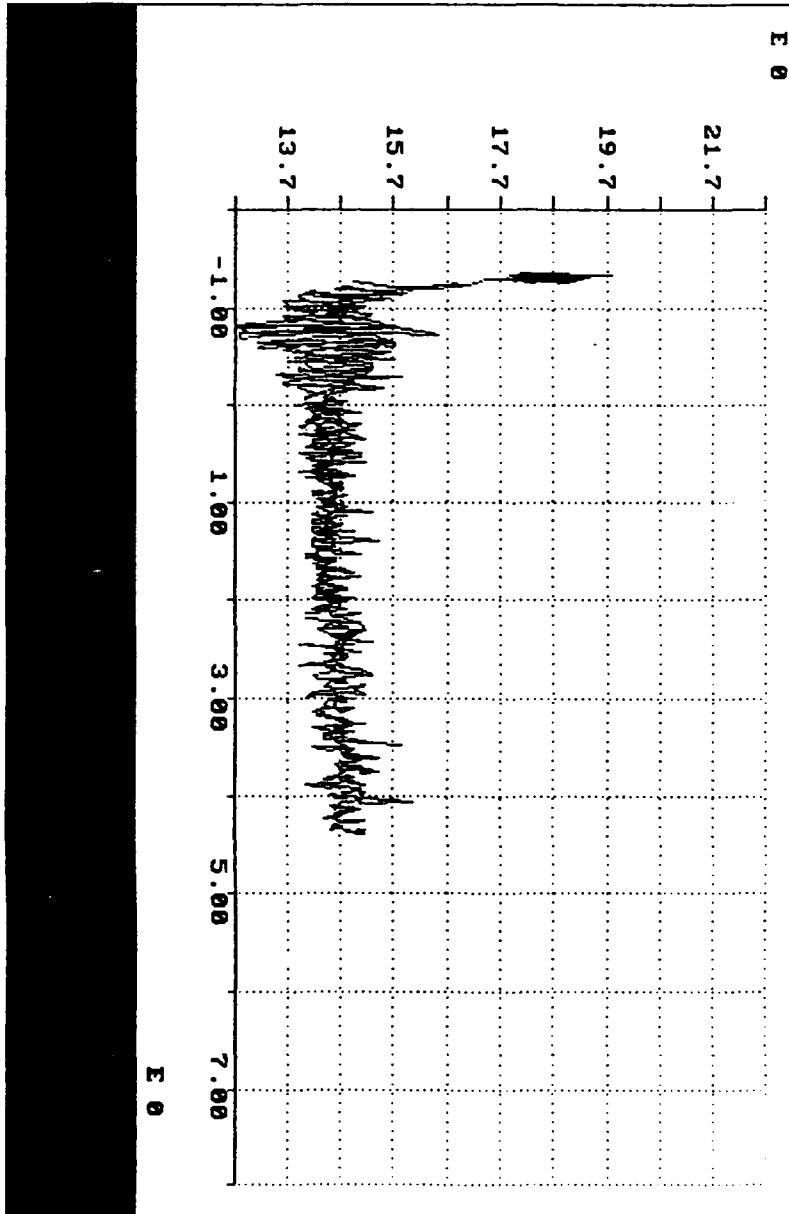




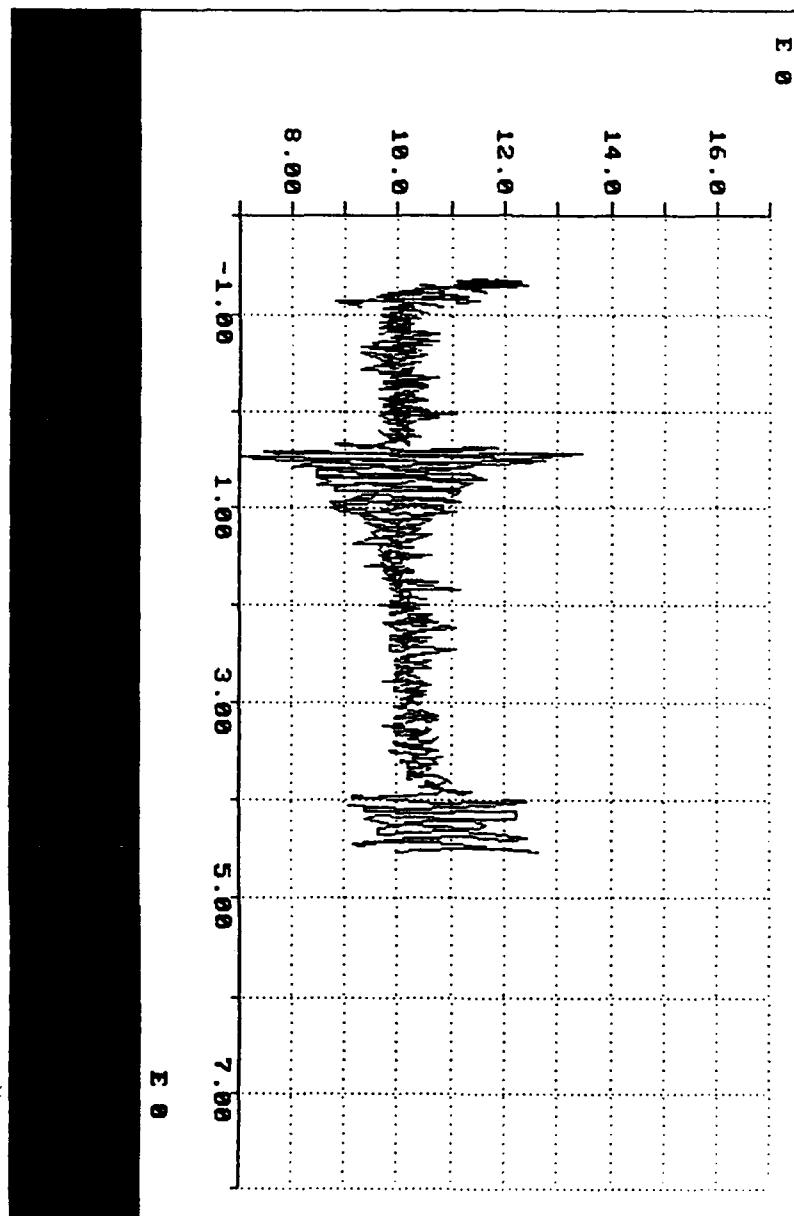


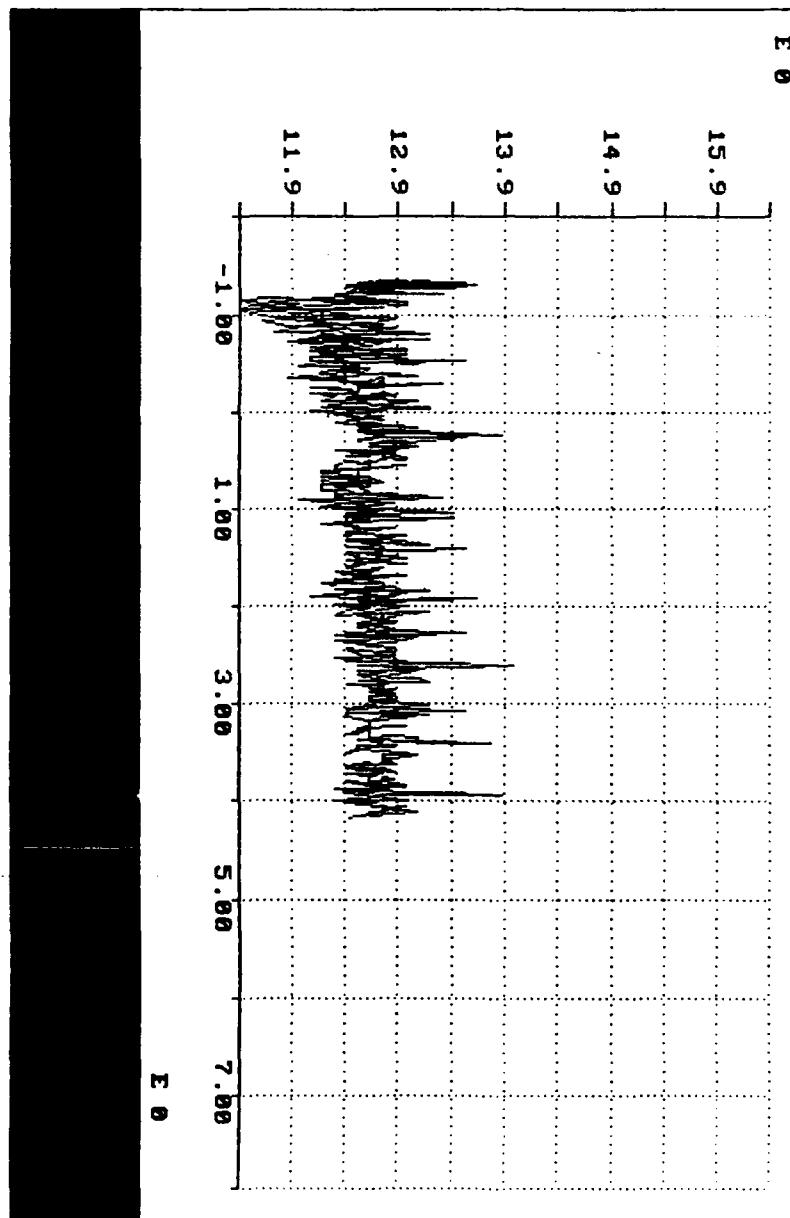


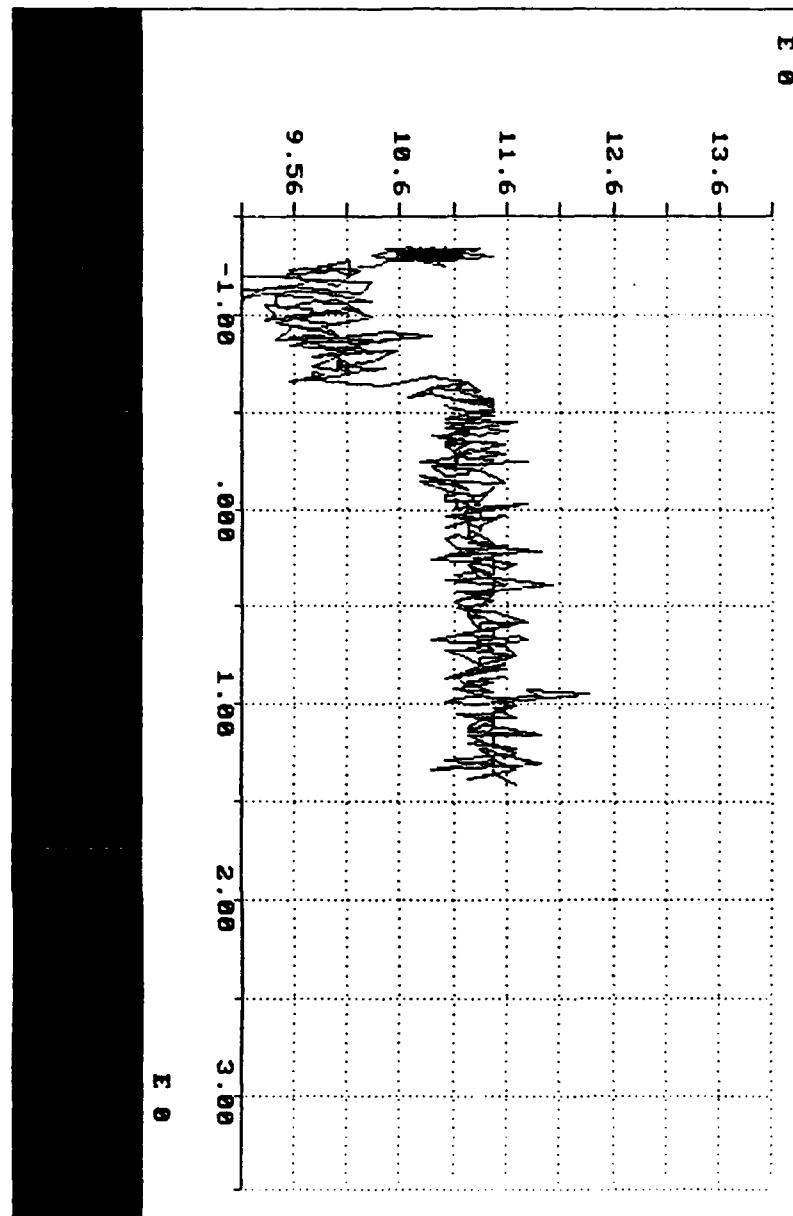


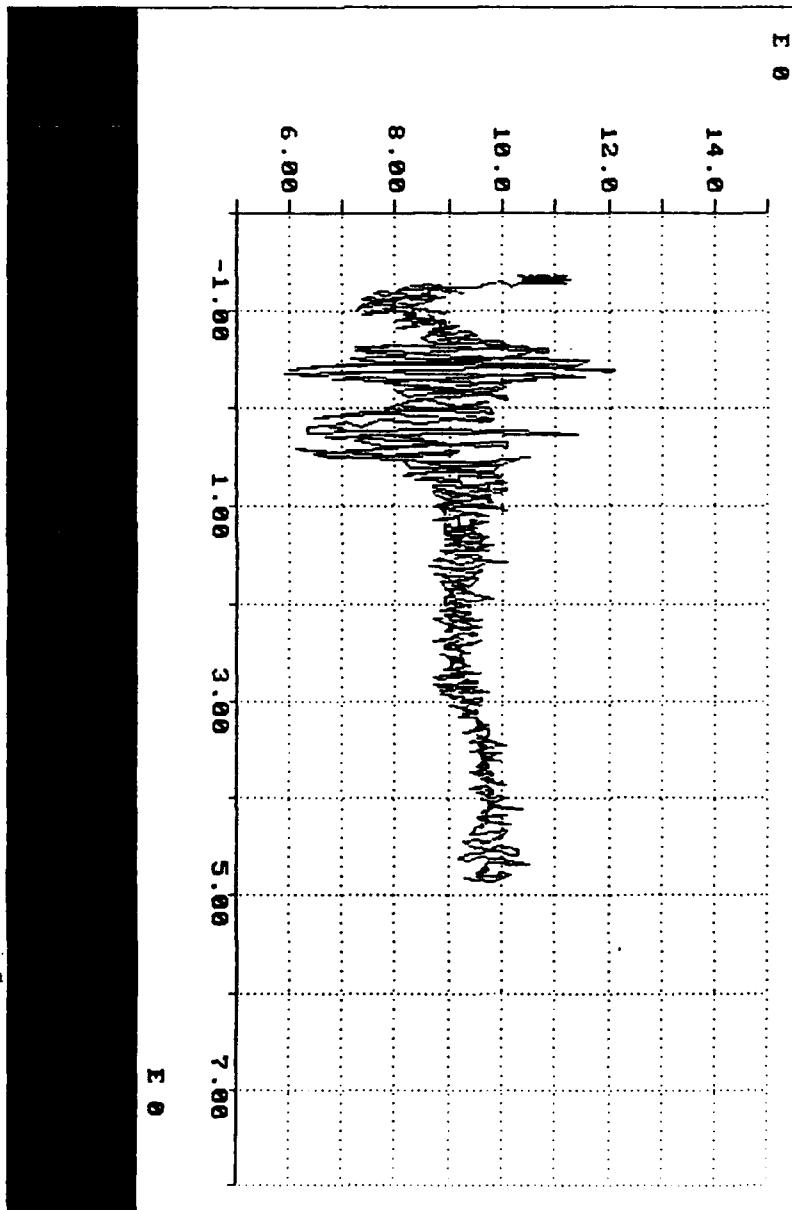


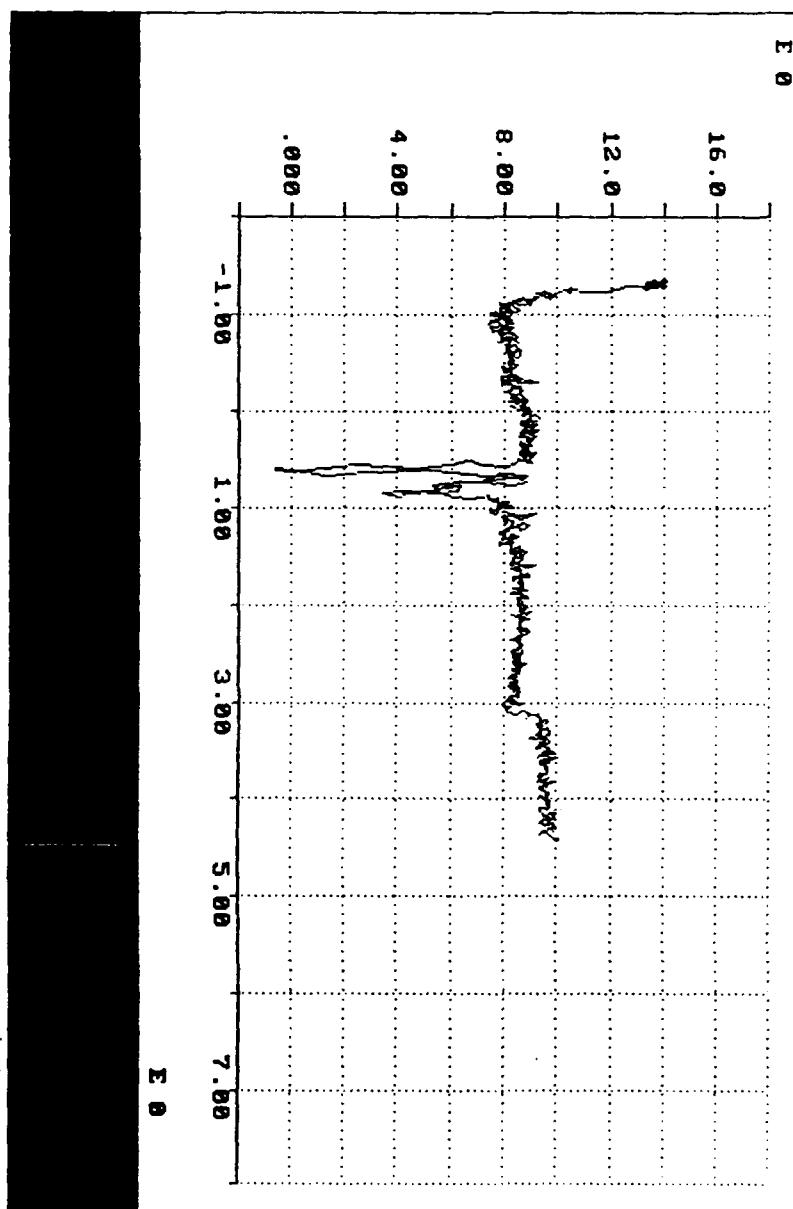
IMMMmm
 : File Name: <B:SEP12F.USR> Quit/Continue: *
 :
 : 8 Comments Subfiles (Total #: 3)
 :> INDIA INK CONTROLLED AT 70 : Start# Shape #hearts :
 : 2> SLIGHT DESSICATION : 1 0 ~ 0.46 :
 : 3> TORE APART FROM SERUSA :
 : 4> MEDIUM STRONG WELD :
 : 5> Small Aperture :
 : 6> SW :
 : 7> :
 : 8>
 LMmm
 : I/O Variable (R - Z): > 768 REAL :
 :
 : Subfile: < 1 > 2 x 256 (Max Length: 768) :
 : Row (0=all): < 0 >
 : Start Column: < 1 >
 : # of Columns: < 256 >
 :
 : Read/Write/Append/Scroll/I'lot/Fedit>List/New file/Quit: *

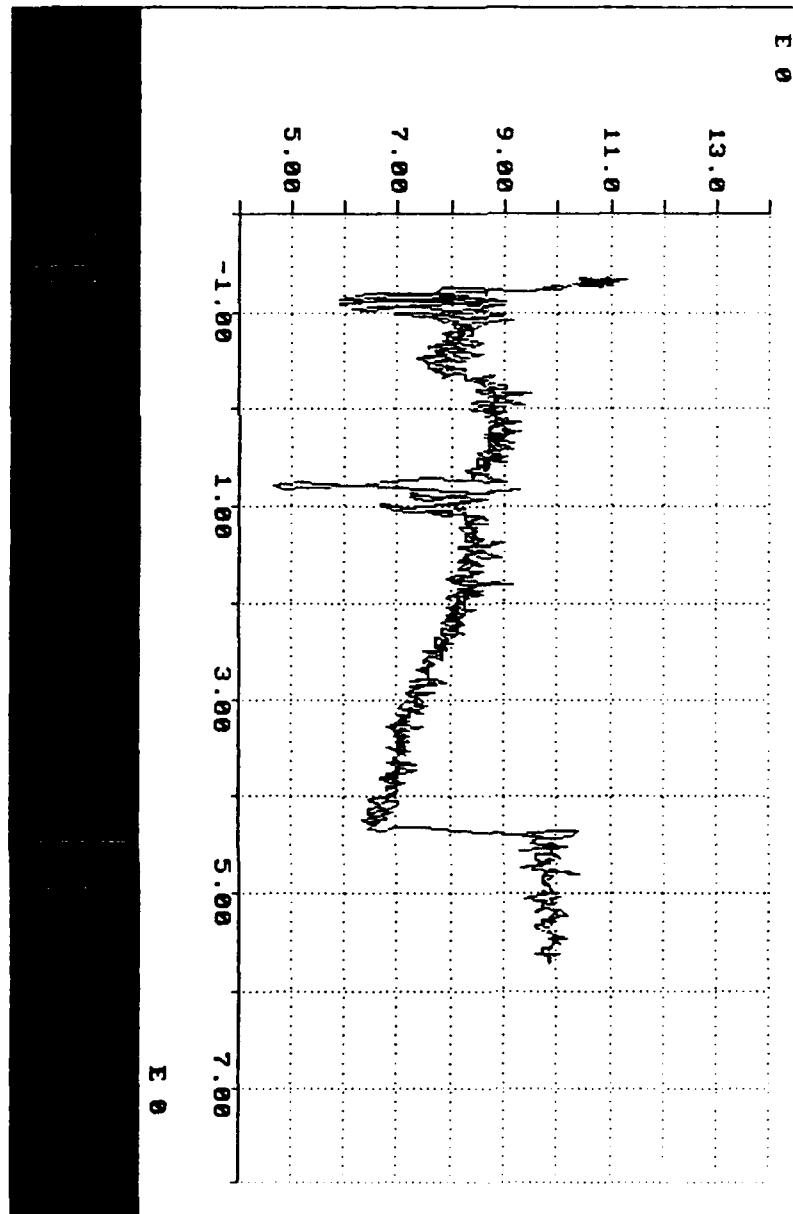


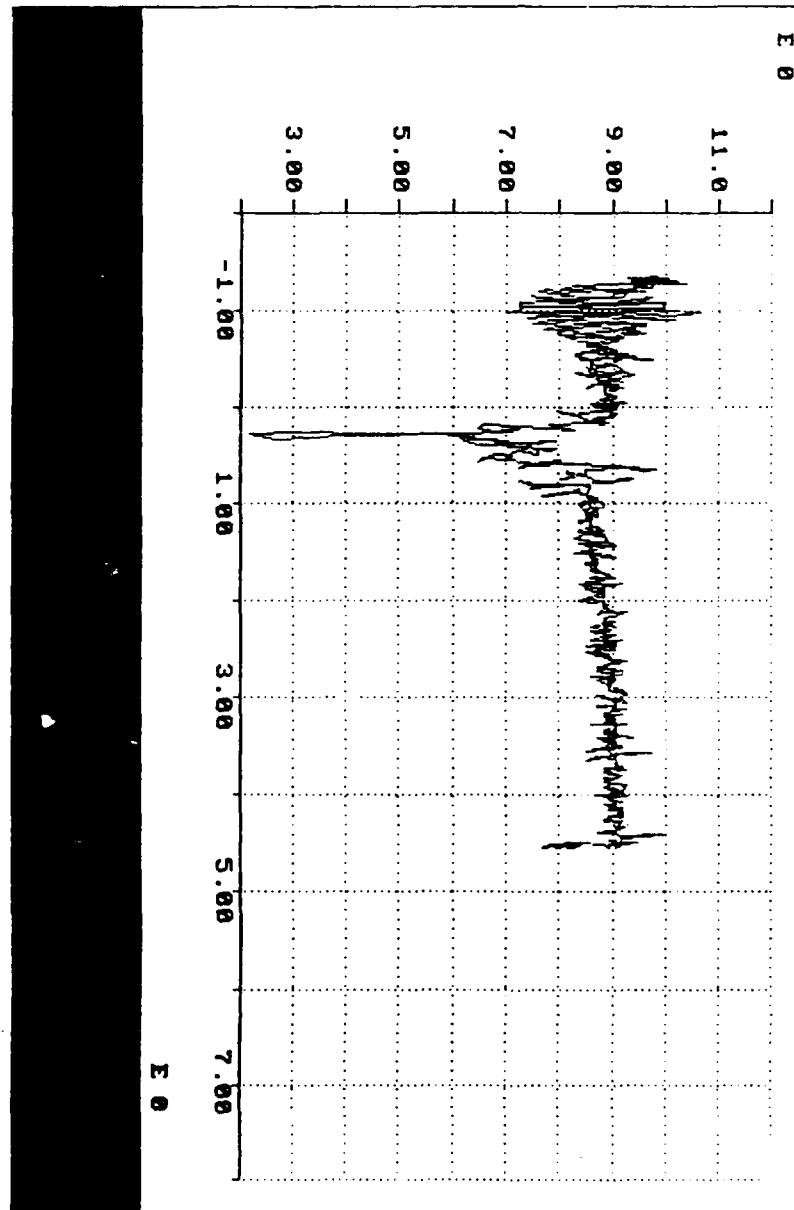


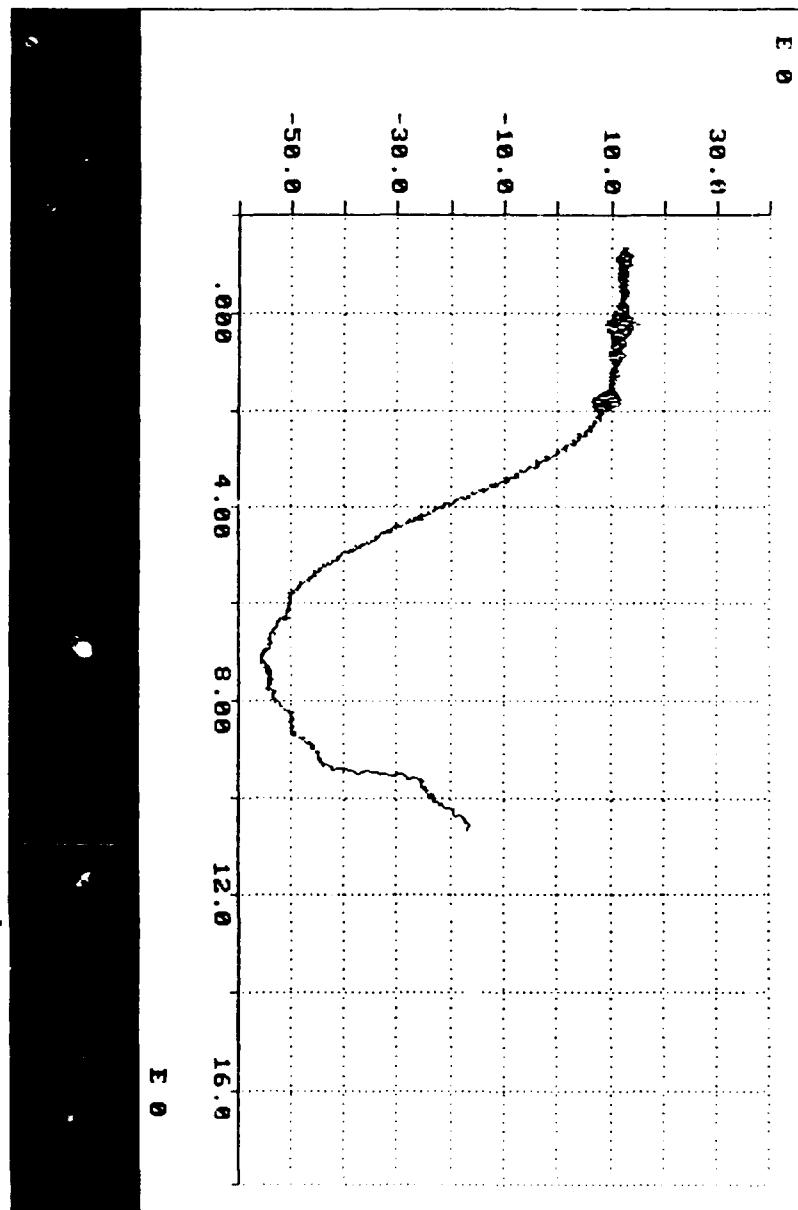








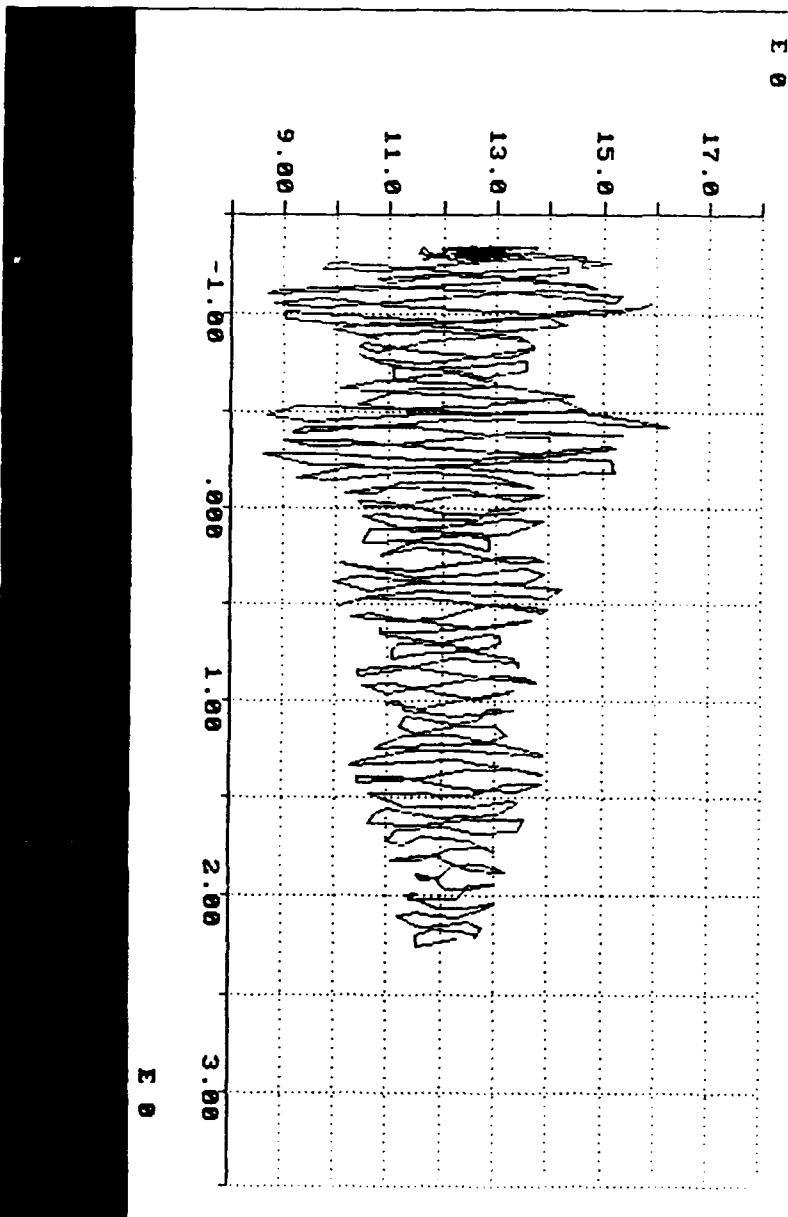


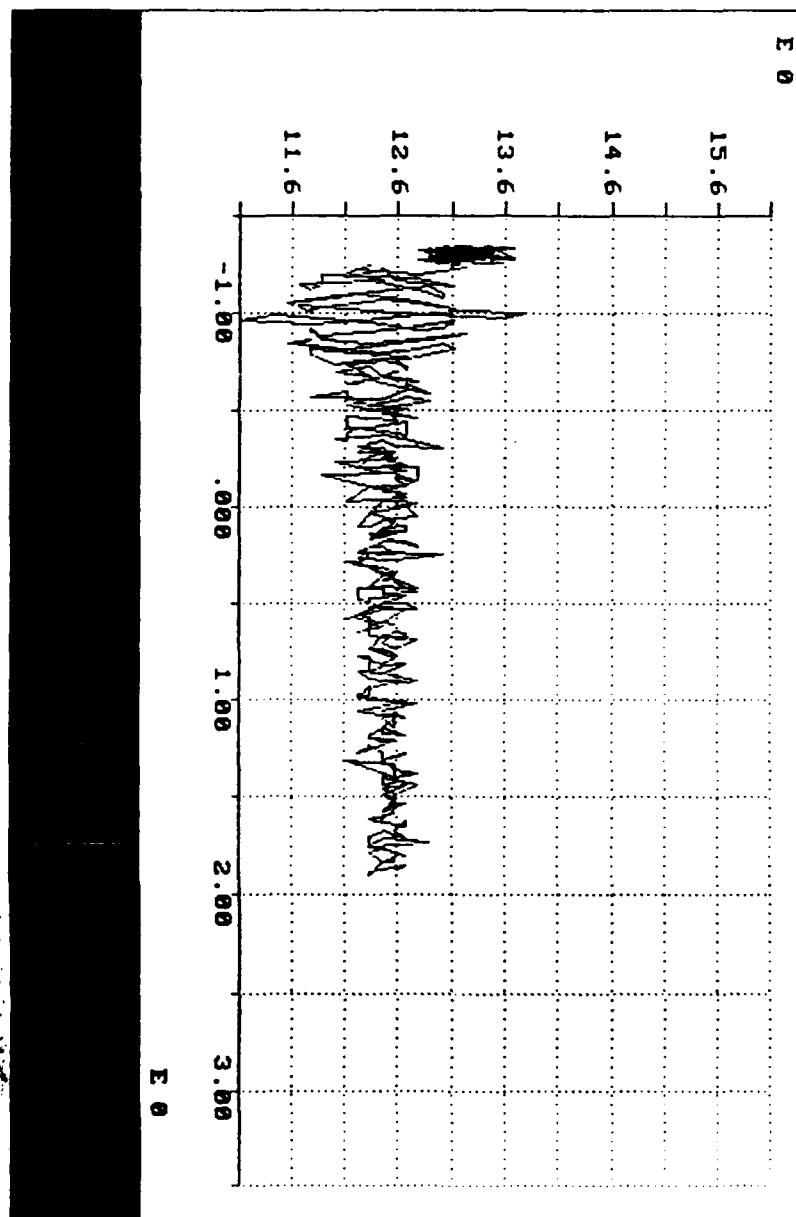


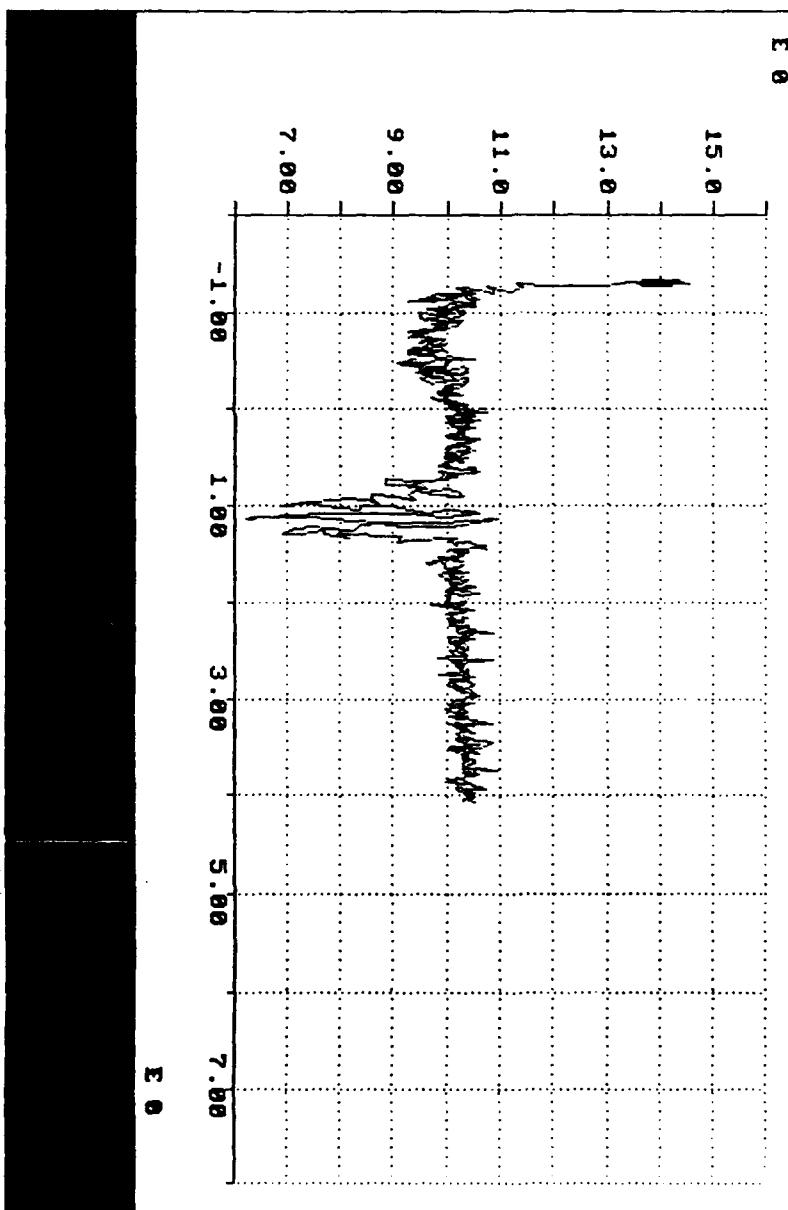
```

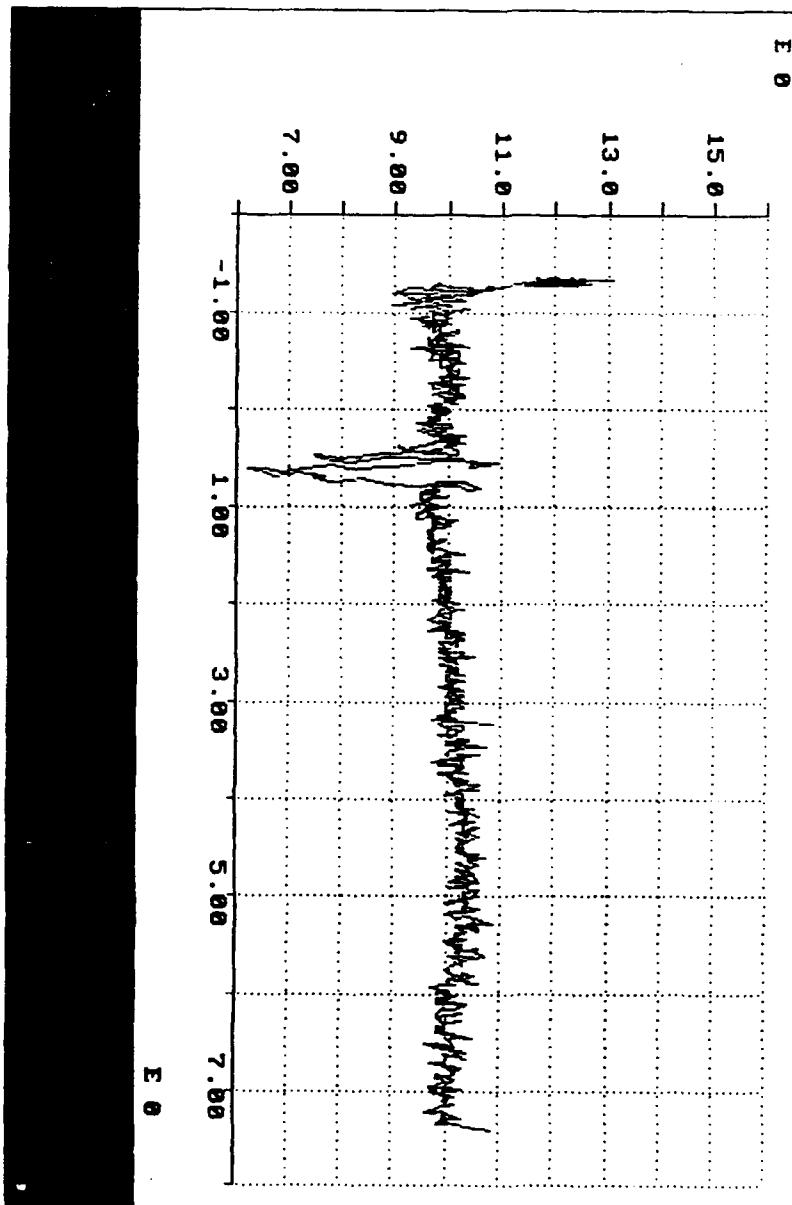
IMMMmmmmmmmmmmmmmmmmmmmmmmmmmm File Input/Output MMmmmmmmmmmmmmmmmmmmmmmmmmmm
:
: File Name: <B:SEPICN.USK>                                Quit/Continue: *
:
: 8 Comments
LMmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmm
: 1' TWO STRIP CONTROL UNWELDED : Start#      Shape      #Repts :
: 2' LARGE APERTURE      : 1          1          1000
: SW
: 3'
: 4'
: 5'
: 6'
: 7'
: 8'
Lmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmm
: I/O Variable ( R - Z ): 1024 REAL
:
: Subfile: < 1 >      2 x 256 (Max Length: 512)
: Row (0=all): < 0 >
: Start Column: < 1 >
: # of Columns: < 256 >
:
: Read/Write/Append/Scroll/Plot/Edit/List/New file/Quit: *
:
Hmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmmm

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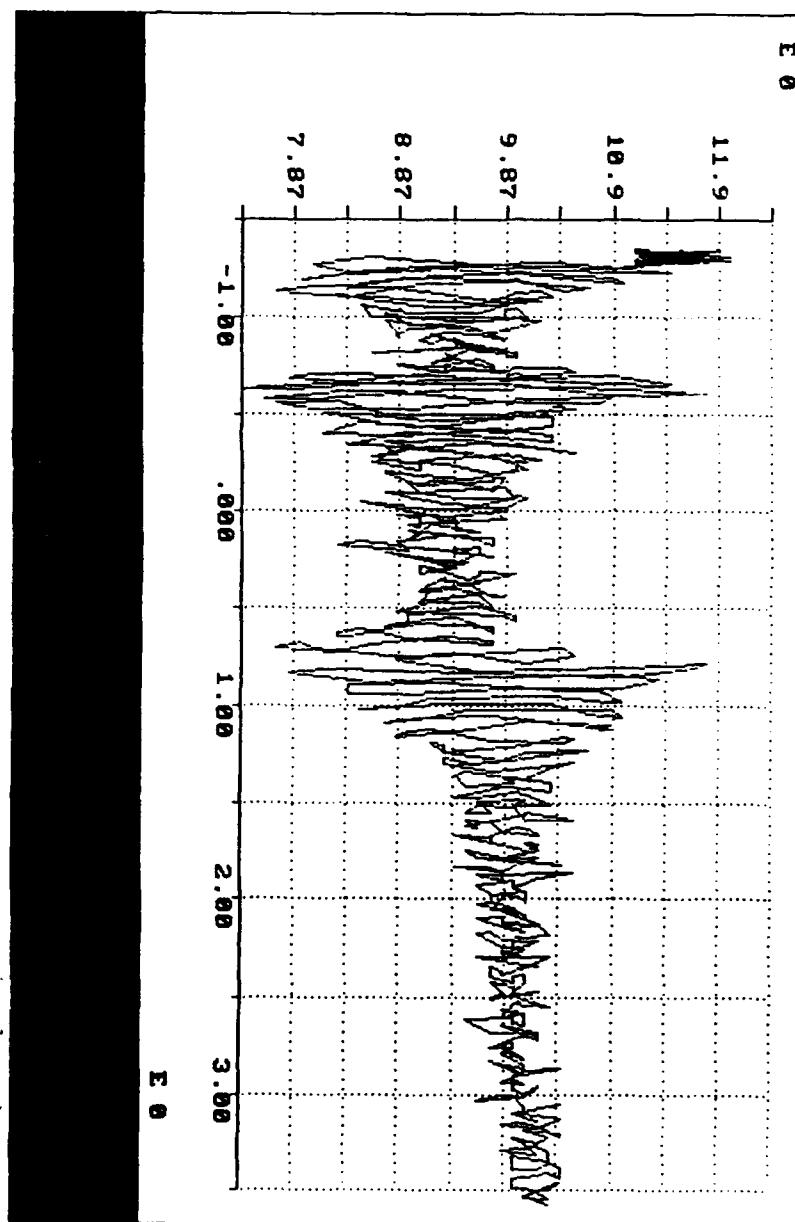


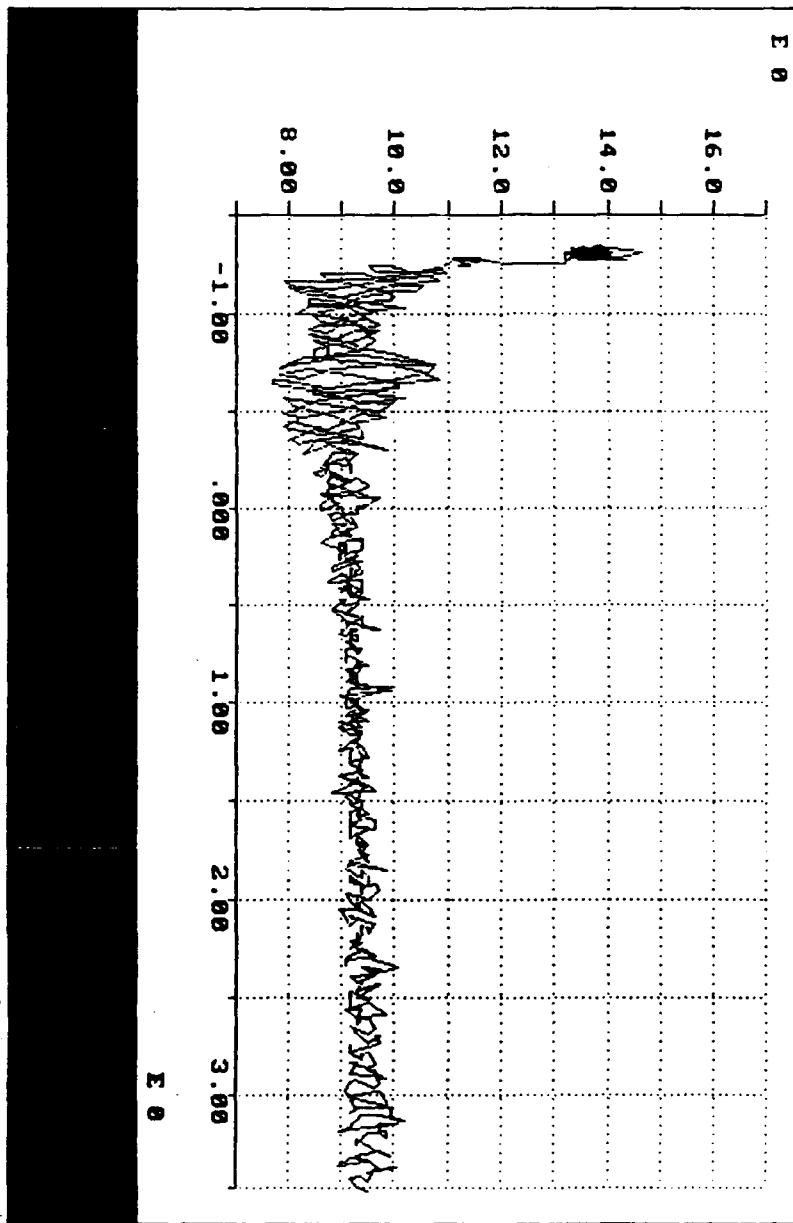


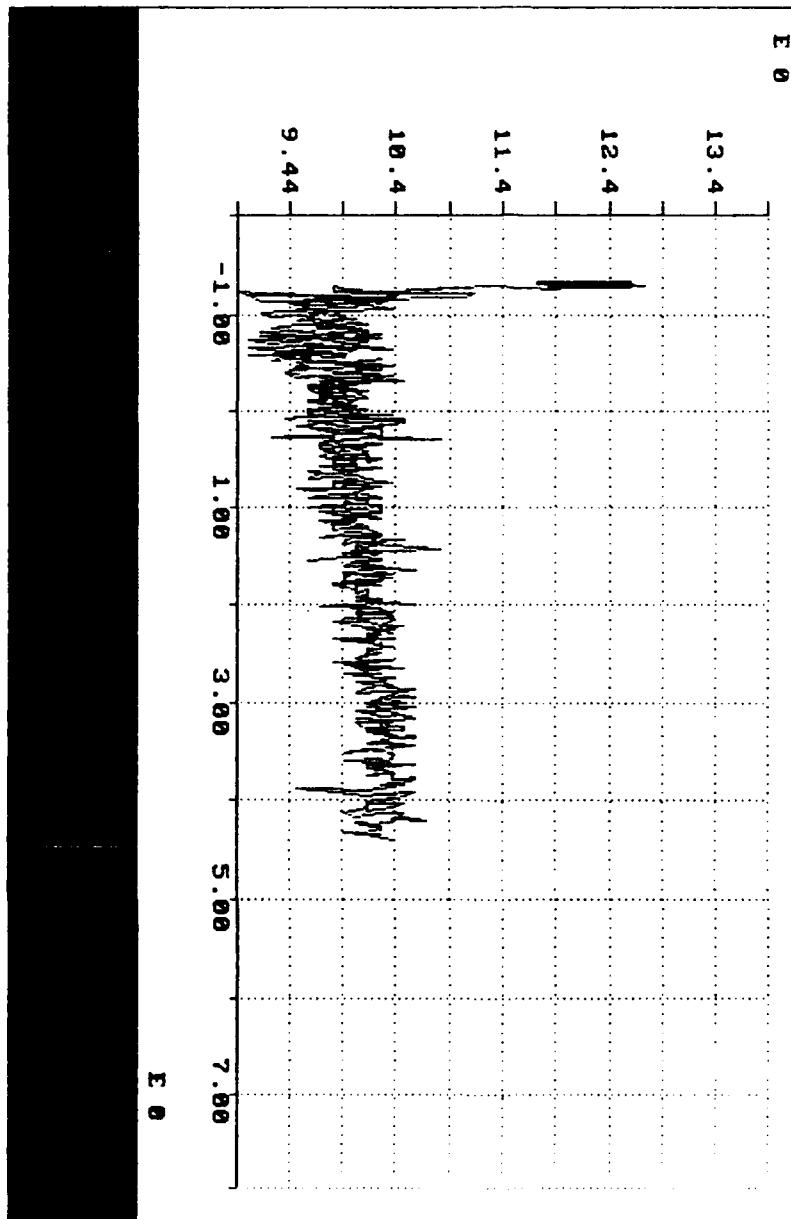


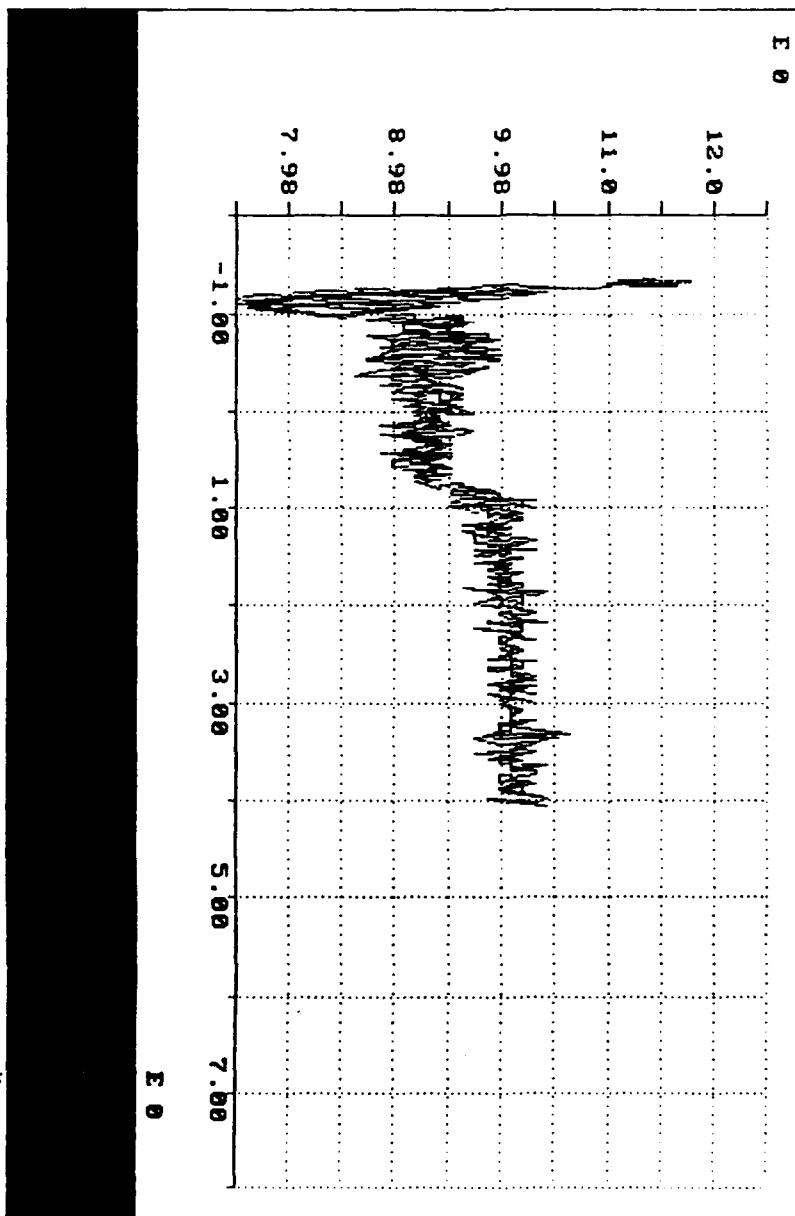


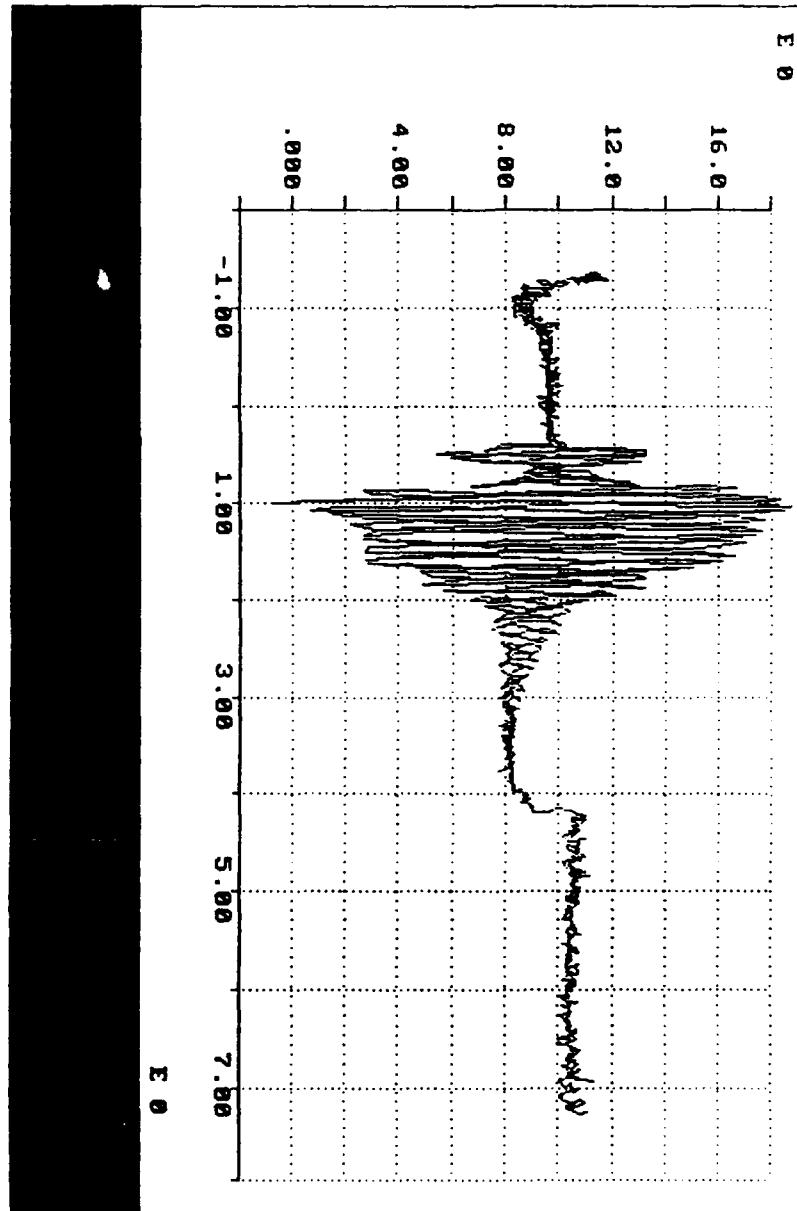
```
: IXXXXXXXXXXXXXXXXXXXXXXXXXXXXX File Input/Output XXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXX;
: : File Name: <B:SER12R.USR           Quit/Continue: *      :
: : B Comments                                Subfiles (Total #: 3 )   :
LXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXX;
: 1> INDIA INK SET=70 TC MAX=60       : Start#      Shape      #Knots   :
: 2> LARGE APERTURE                  :      1          2 x 256    :
: 3> SLIGHT DESSICATION             :      2          2 x 256    :
: 4> WEAK WELD                      :      3          2 x 256    :
: 5> SW                           :      4          2 x 256    :
: 6>                               :      5          2 x 256    :
: 7>                               :      6          2 x 256    :
: 8>                               :      7          2 x 256    :
LXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXX;
: I/O Variable ( R = Z ):               1024 KBAL   :
: : Subfile: < 1 >          2 x 256 (Max Length: 168)   :
: : Row (0=all): < 0 >          2 x 256 (Max Length: 168)   :
: : Start Column: < 1 >          2 x 256 (Max Length: 168)   :
: : # of Columns: < 256 >          2 x 256 (Max Length: 168)   :
: : Read/Write/Append/Scroll/Plot/Edit>List/New file/Quit: *   :
LXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXX;
```

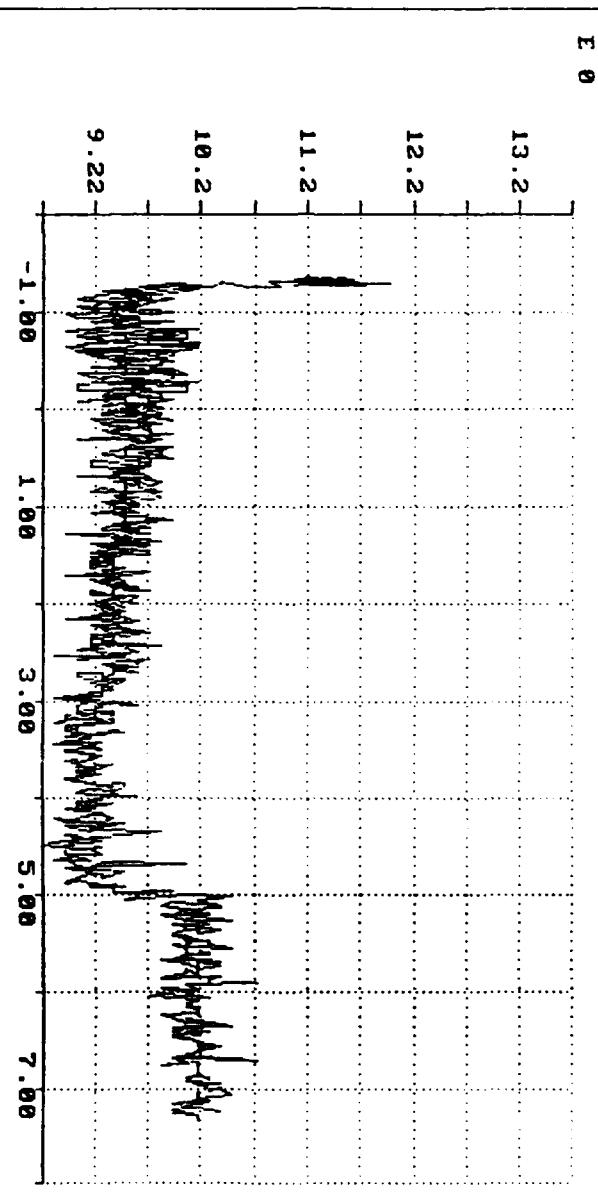


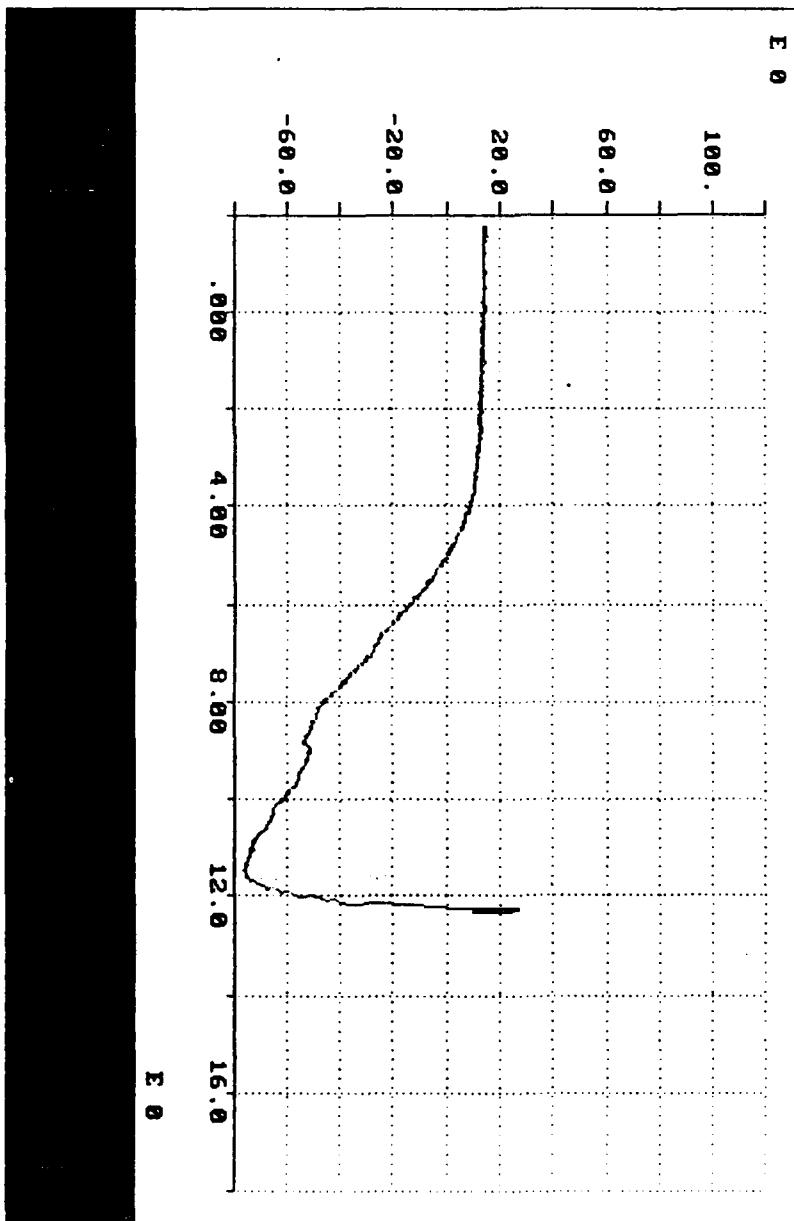


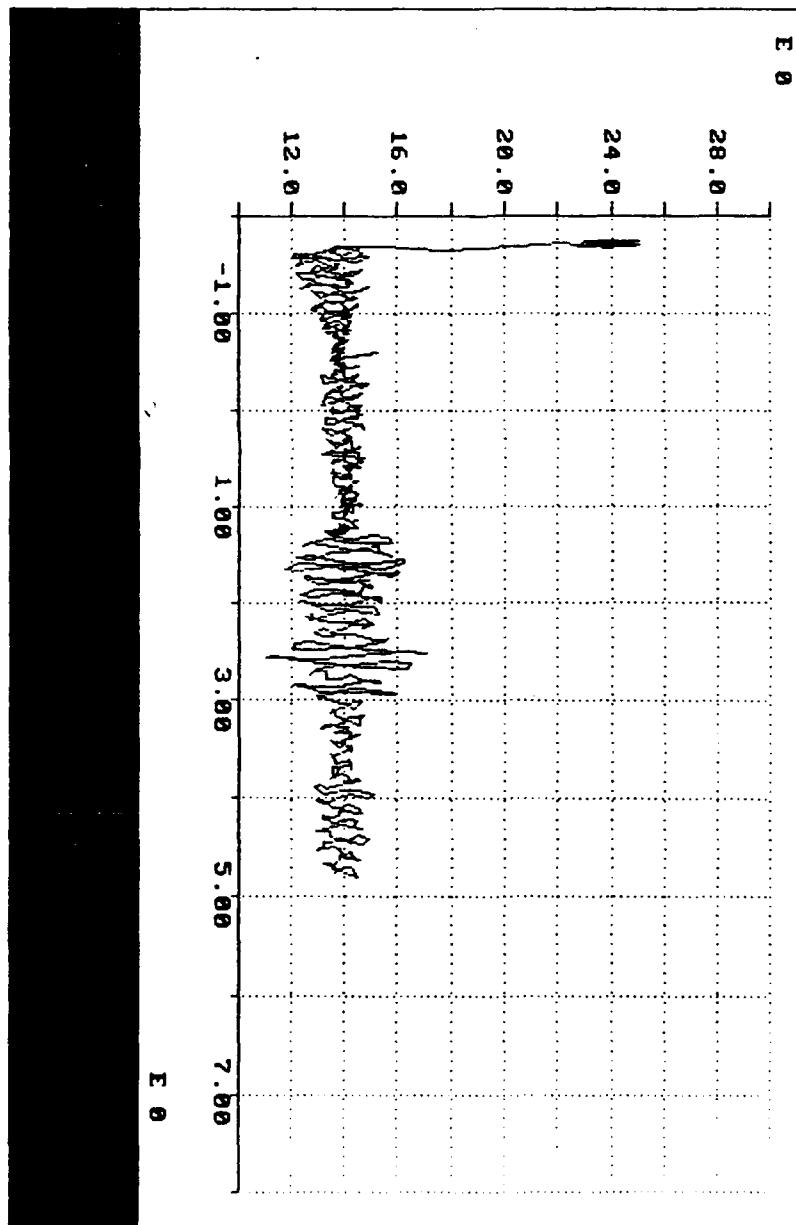


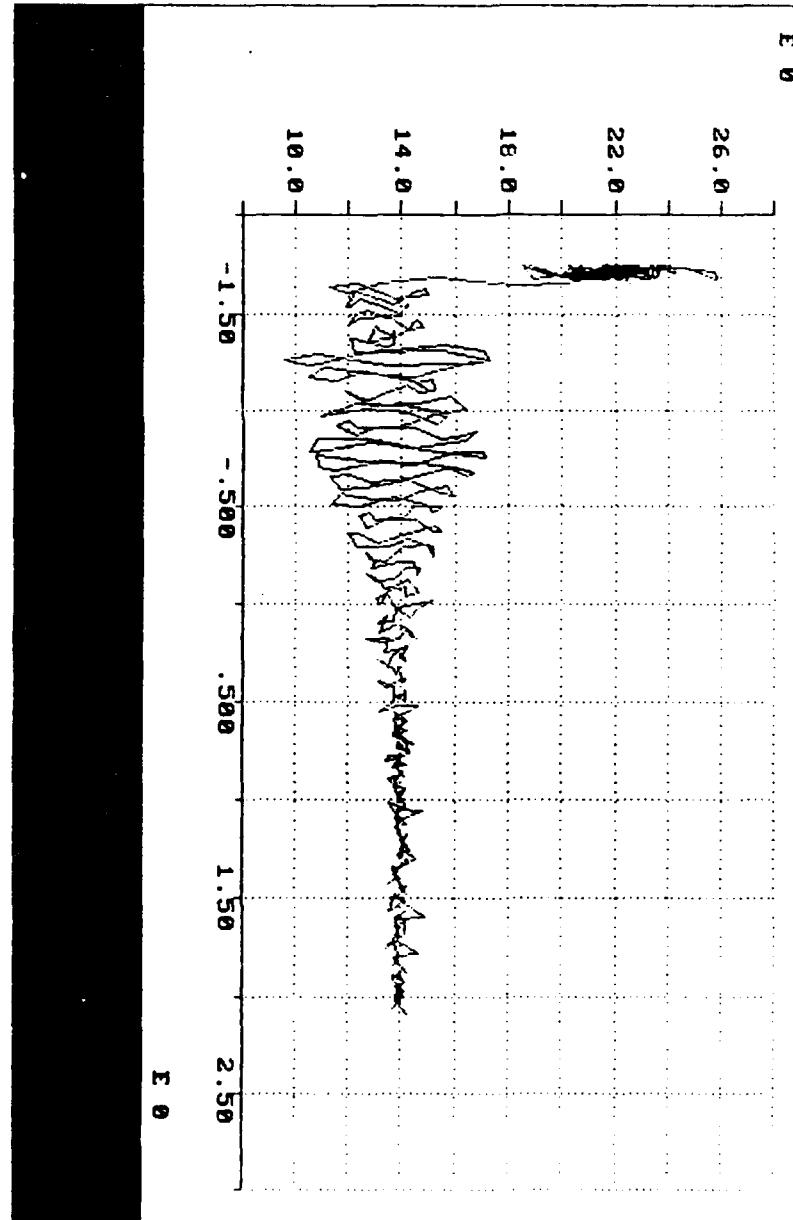








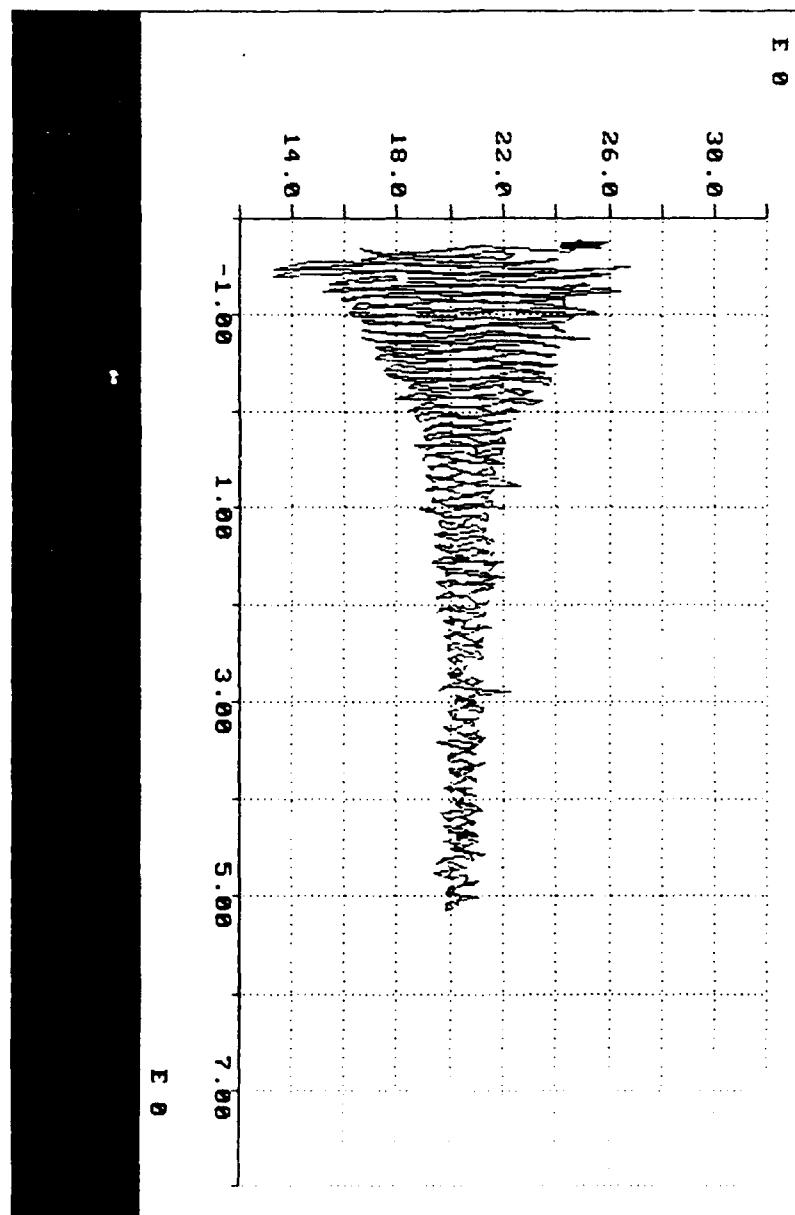




```

: File Name: C:\SER14D.DAT          Date: 10/10/95   Description: INDIA INK CONTROLLED AT 50% 100% VISION
: Subfile: C:\SER14.DAT             # Lines: 4
: 8 Comments
: 1 INDIA INK CONTROLLED AT 50% : Start#      Shape    #F exite
: 2 NO VISUAL CHANGES           : 1          2 + 256
: 3 NO SMOKE
: 4 NO OBVIOUS WELL SPOTS
: 5 BOWEL LOOKS GOOD
: 6
: 7
: 8
: I/O Variable ( F = 7 ):       S1: 1E0.
: Subfile: 1                      C = 256  Bla Length: 128
: Row (0=all): 0
: Start Column: 1
: # of Columns: 256
: Read/Write/Append/Scroll/File/Edit/List/New file/Quit: *

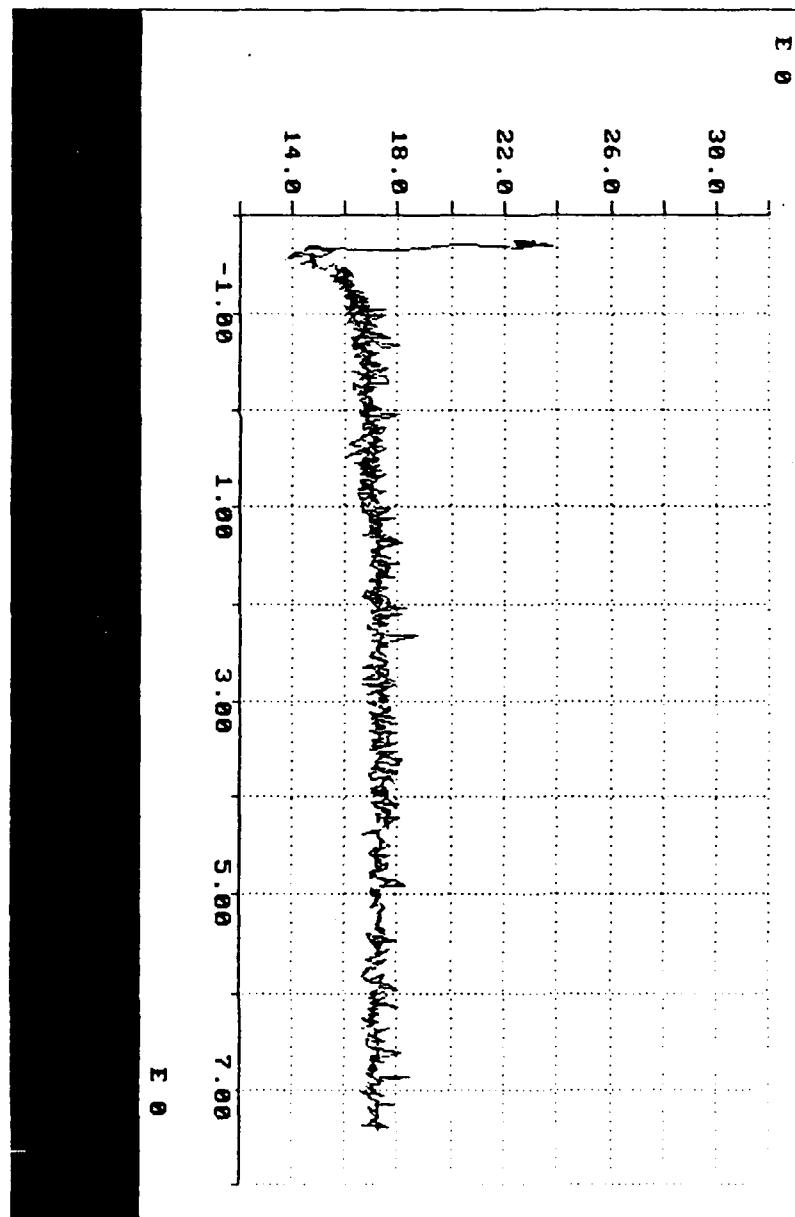
```

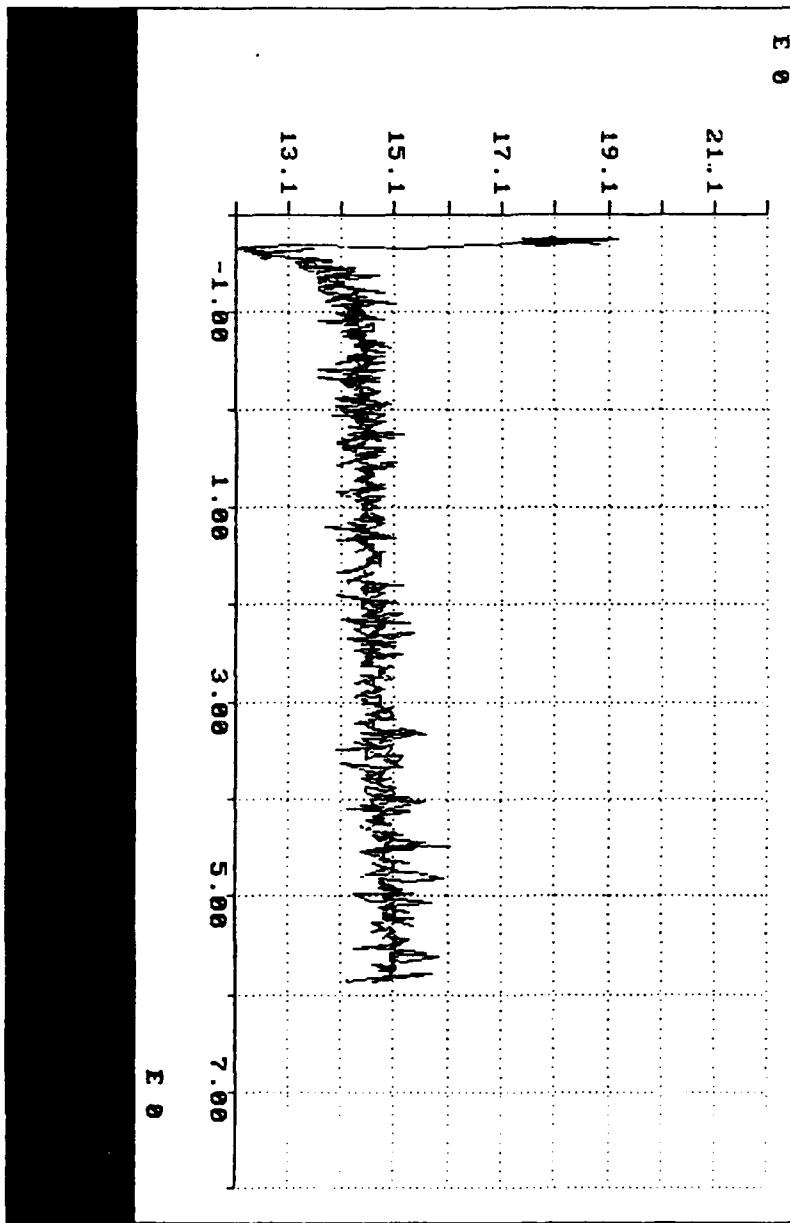


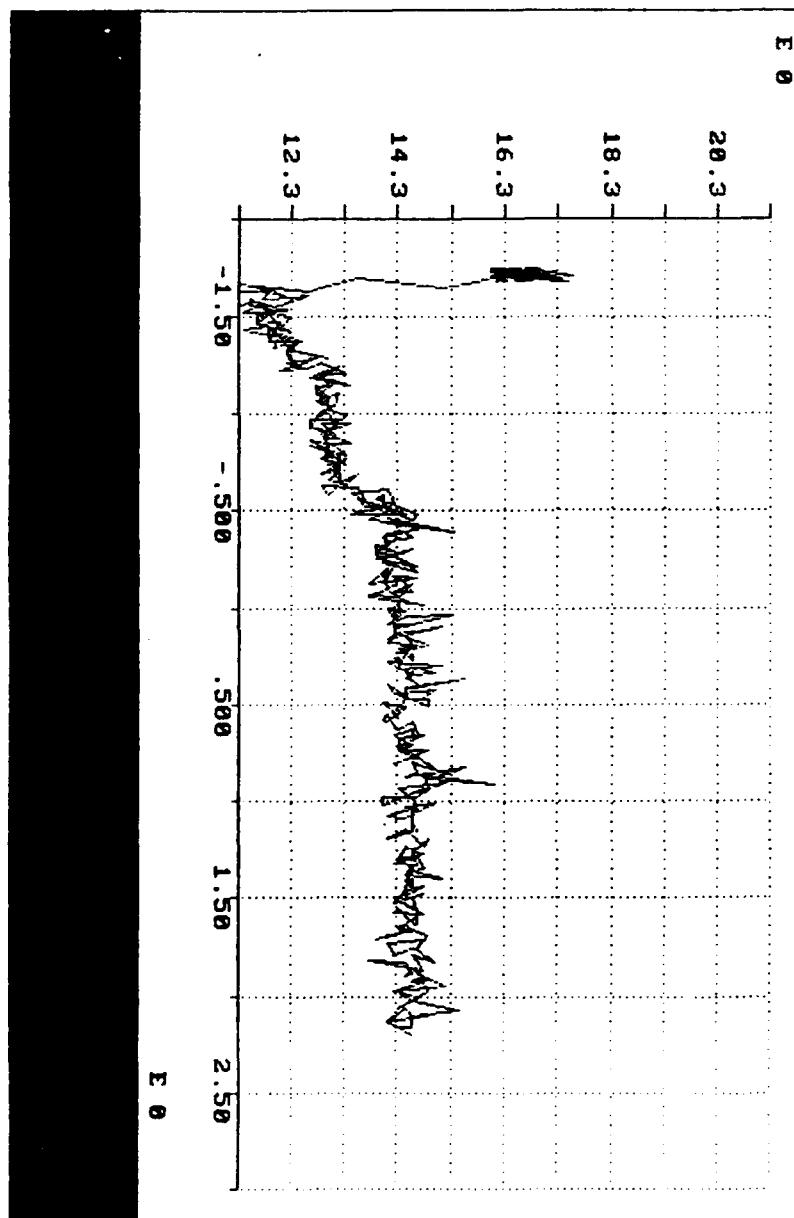
```

: File Name: <C:SEF14E.USK>          Quit/Continue: *
: B Comments                                Subtitles (Total #: 1)
: 1 INDIA INK CONTROLLED AT 60             : Start#      Shape    Effects:
: 2 NO VISUAL CHANGES                      : 1           2      158
: 3 NO SMOKE
: 4
: 5 WEAK WELD
: 6 ERODE FROM SIDE(S) - 100% DEPTH
: 7
: 8
: I/O Variable ( R = Z ):                  768  BLK
: Subfile: < 1 >                         2 :: 256 (Max Length: 768 )
: Row (0=all): < 0 >
: Start Column: < 1 >
: # of Columns: < 256 >
: Read/Write/Append/Script/Plot/Edit/List/New file/Quit: *

```



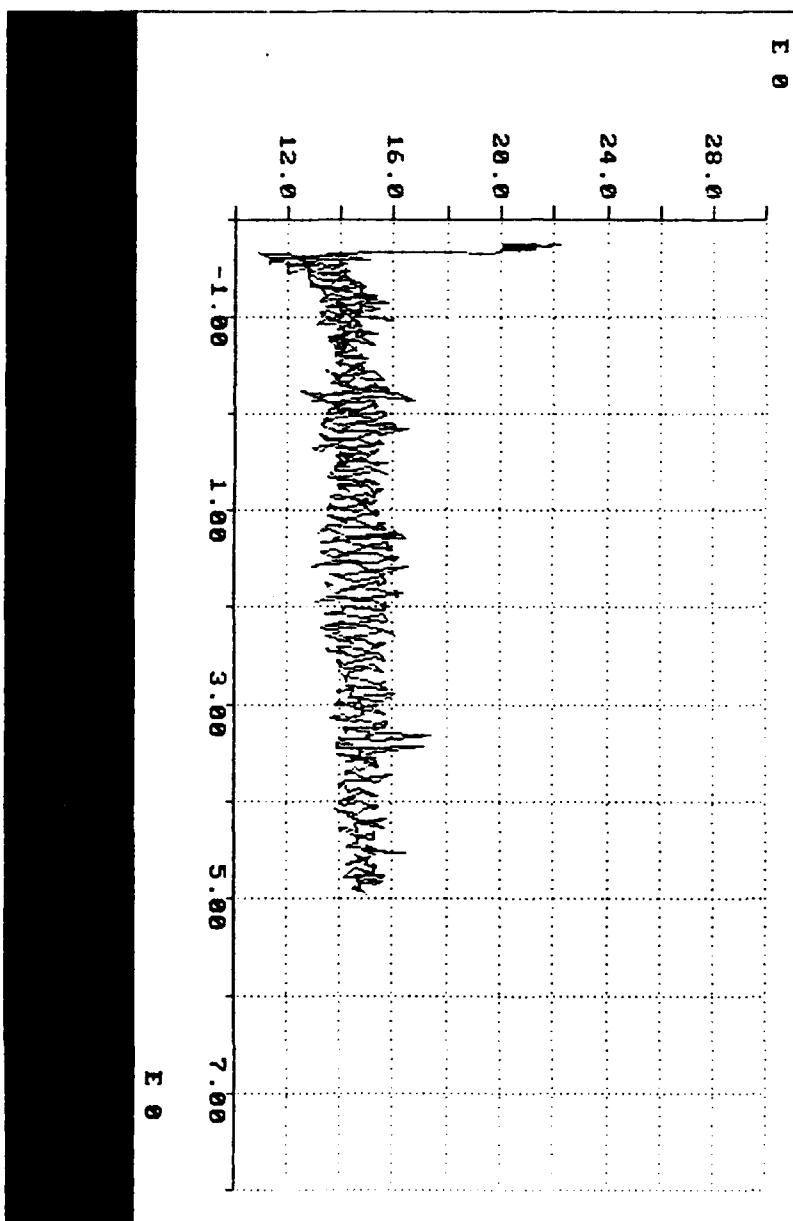


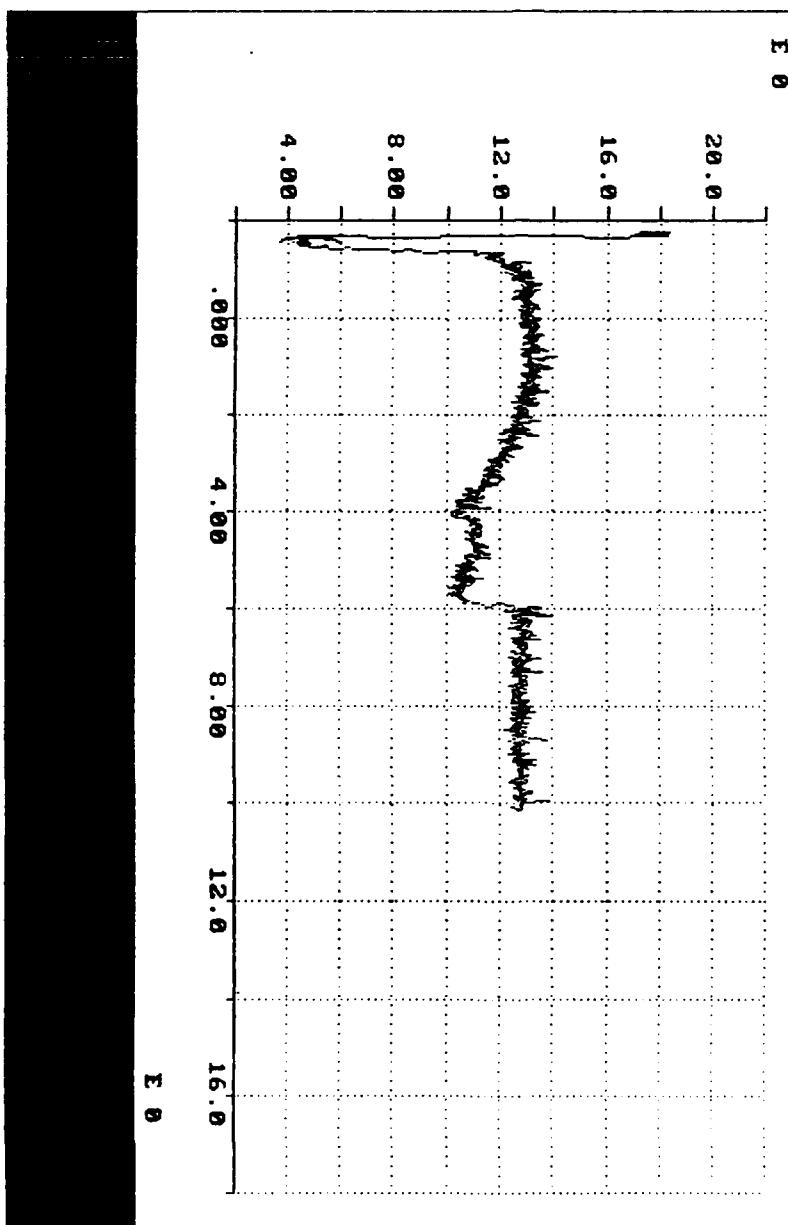


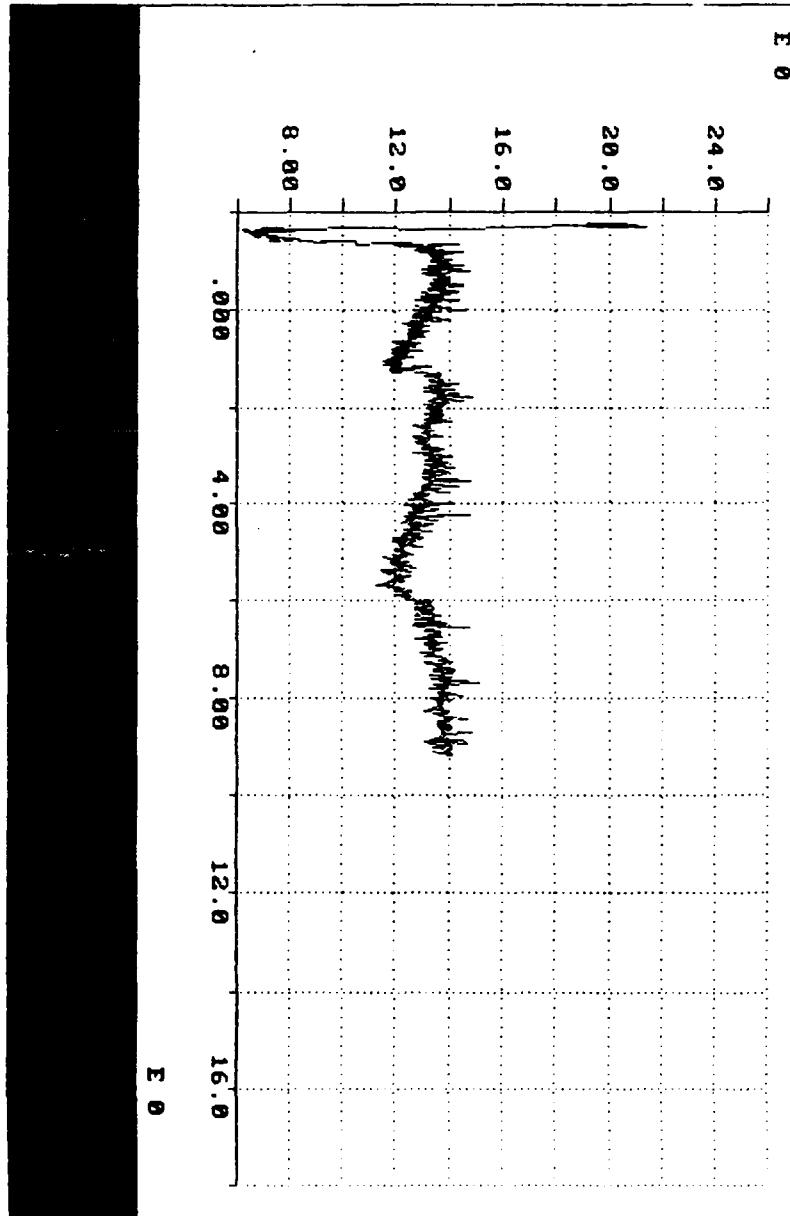
```

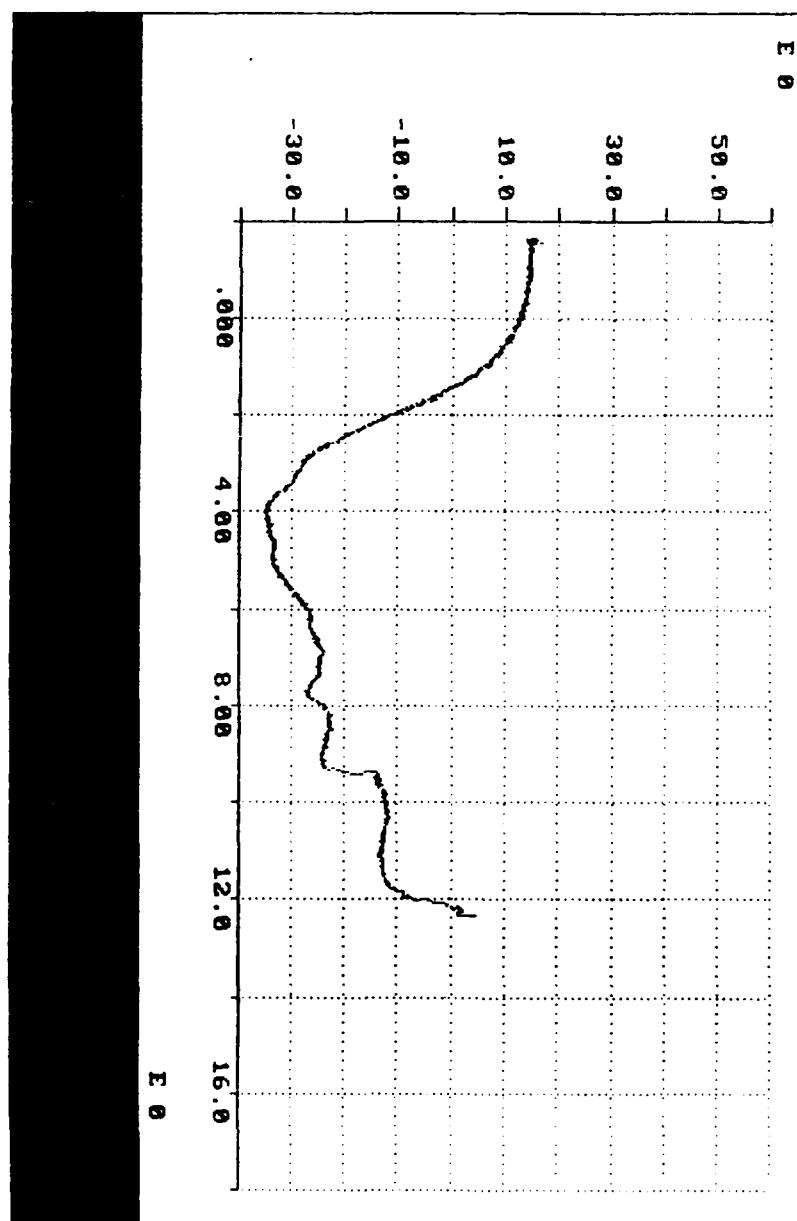
: File Name: <C:SEF14H.USP>          Reut/Confidence: *      :
: Subfile: 1 Total #: 31      :
: 8 Comments      :
: 1 INDIA INK SET=BO TC MAX=65      :
: 2 SLIGHT DESSICATION      :
: 3 NO CARBONIZATION      :
: 4 SLIGHT BLANCHING      :
: 5 NO SMOKE      :
: 6      :
: 7 BD LEANED ON TABLE      :
: 8 NO EVIDENCE OF A WELD      :
: I/O Variable ( R = 2 ):      :
: Subfile: 1      :
: Row (0=all): 0      :
: Start Column: 1      :
: # of Columns: 256      :
: Read/Write/Append/Scroll/Plot/Edit/List/New file.Reut: *      :

```

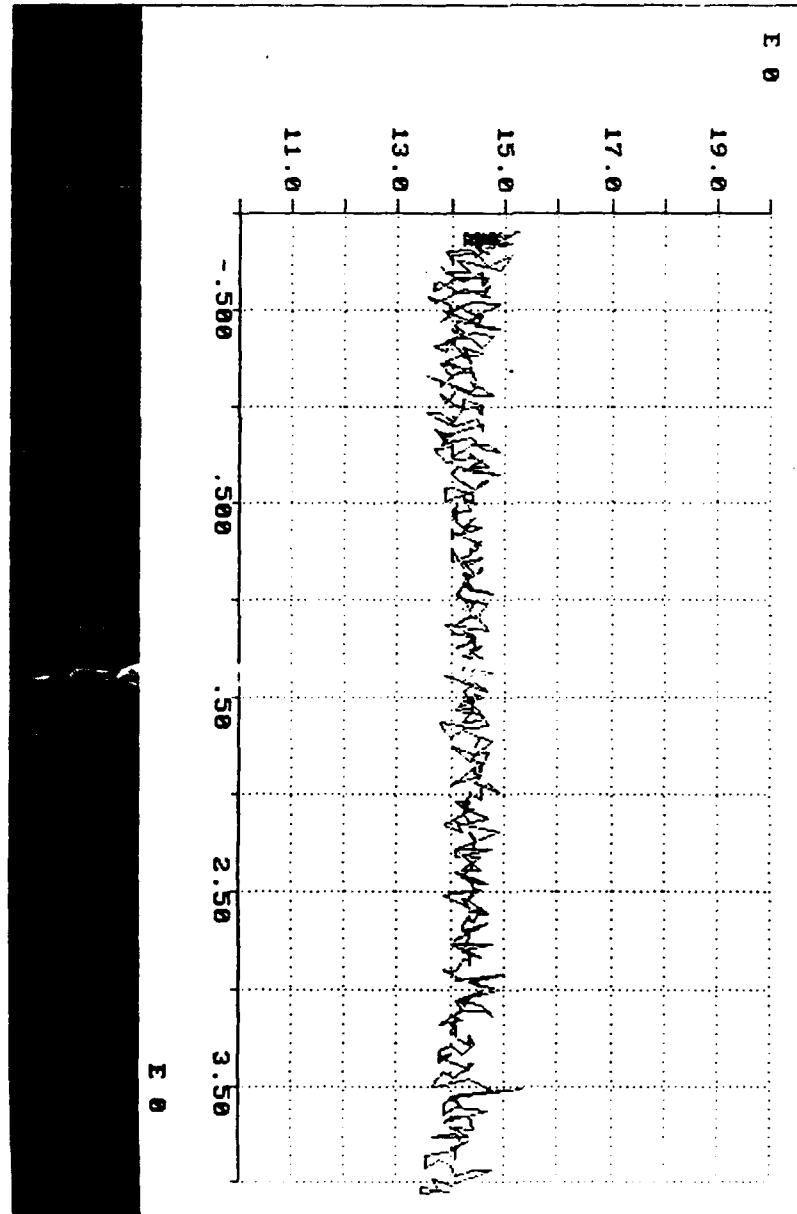






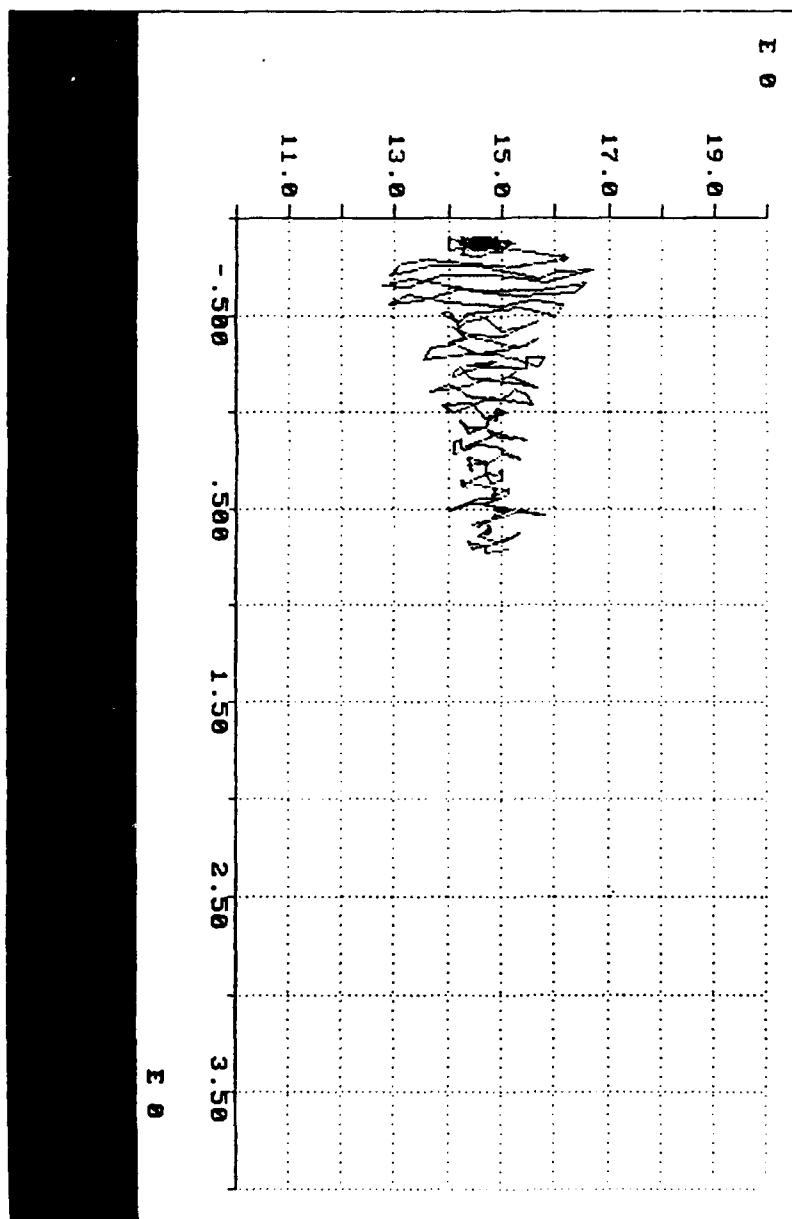


```
File Name: <C:SEF14.LIST          Edit/Cont/Line/Block/Select/Print/Exit/Help/Quit: *  
8 Comments  
1 TWO STRIP CONTROL UNITS.DAT  
11  
12  
13  
14  
15  
16  
17  
18  
19  
20  
21 Variable ( R = 1 )  
Subfile: - 1 -      7 x 2156 (Max. Length: 100)  
Row (0=All): < 0  
Start Column: - 1 -  
# of Columns: 100  
Read/Write/Append/Scroll/Left/Edit/List/New/ file/Quit: *
```



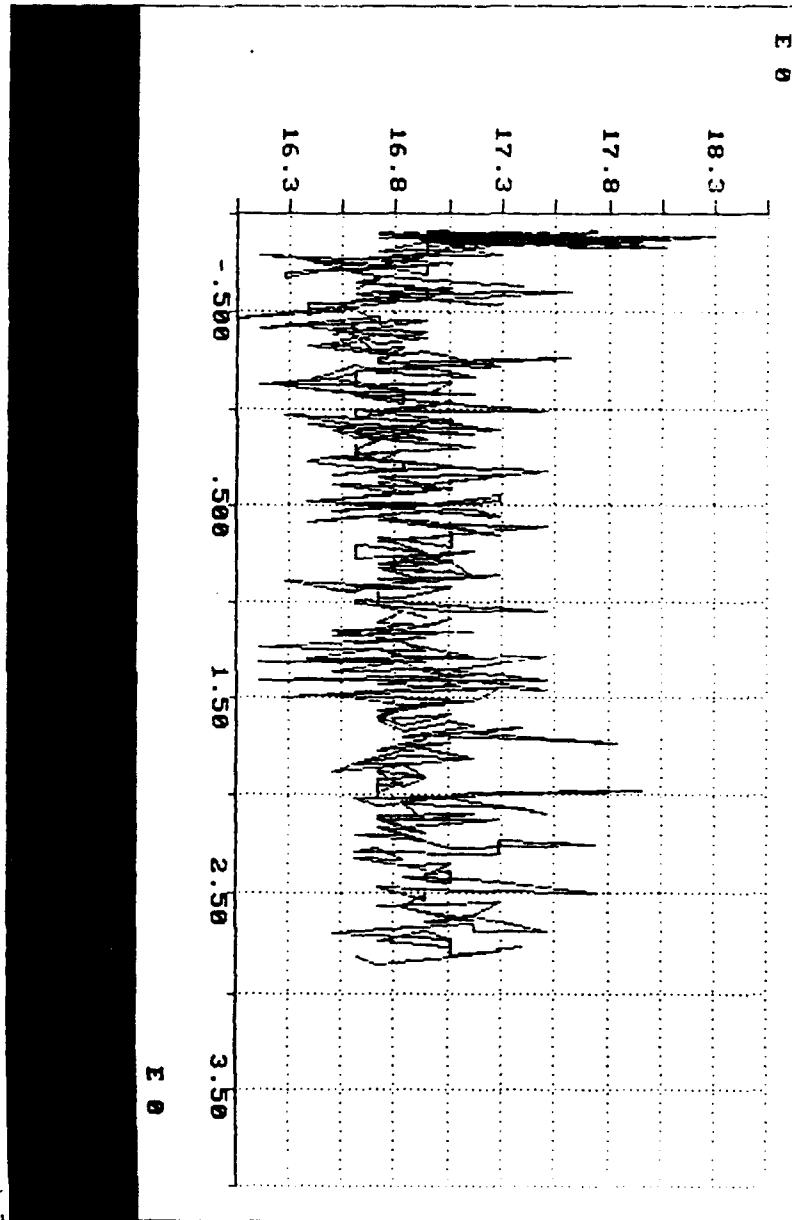
```

: File Name: C:\SER140\U10.DAT          Units/Decimals: *          :
:                                     Subfile: Listed at 1          :
:                                     Suffix:          :
: 8 Comments          :
: 1 TWO STRIP INDIA AND PIGUE DE L'INDIEN          :
: 2          :
: 3          :
: 4          :
: 5          :
: 6          :
: 7          :
: 8          :
: I/O Variable ( R = 2 ):          :
:                                     Subfile:          :
:                                     Row (0=all):          :
:                                     Start Column:          :
: # of Columns: 256          :
:                                     Read/Write/Append/Scratch/Fmt/Right/Left/New/Ext/Trunc: *          :
:
```



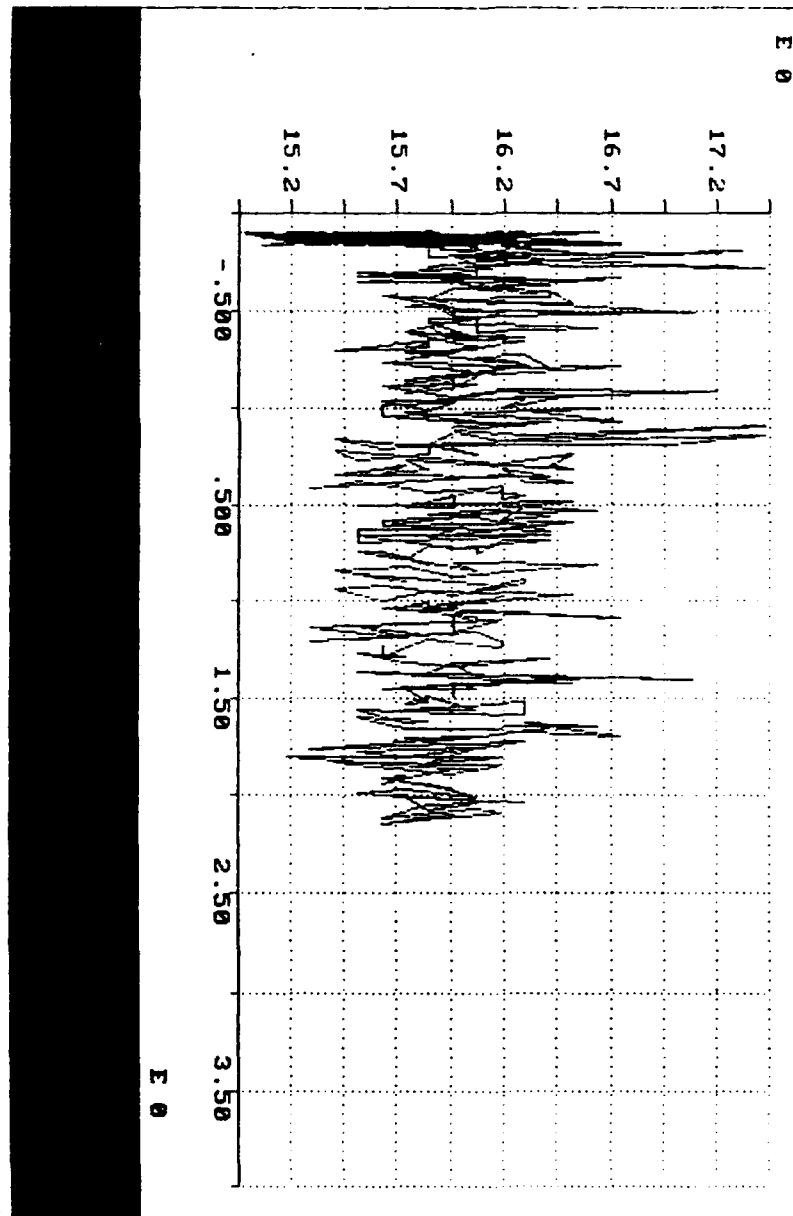
```

: File Name: INDIA.DAT
: 
: B Comments
: 
: 1 INDIA INK CONTROLLED AT 50
: 2 SMALL APERTURE
: 3 NO VISUAL CHANGE
: 4
: 5 NO WELD SEEN
: 6
: 7
: 8
: I/O Variable ( R = 1 ) : 25 1.00
: 
: Subfile: < 1 > 0 x 256 (Max Length: 256)
: Row (0=all): < 0 >
: Start Column: < 1 >
: # of Columns: < 256 >
: 
: Read/Write/Append/Screen: QUIT/Exit/Last/Next/First/Help
:
```



```

: File Name: C:\SER\14.JESR          Outfile (total # of lines): 4
: 
: 8 Comments
: 1 INDIA INK CONTROLLED AT 60      : Start #: 1      Chase #: 156
: 2 SLIGHT SMOKING                 : 
: 3 SLIGHT BLANCHING               : 
: 4 NO DESSICATION                : 
: 5 WEAK WELD                      : 
: 6 TORE AT SEROSA                 : 
: 7 
: 8 
: I-O Variable ( F = Z ):          : End #: 156
: 
: Subfile: 1                         : P = 156 (Max Length: 512)
: Row (0=all): 0                      : 
: Start Column: 1                     : 
: # of Columns: 256                  : 
: 
: Read/Write/Append/Replace (0=Read, 1=Write, 2=Append): 0
:
```



File Name: C:\SERV1\X11.DAT Date Created: 10/11/92 10:00 AM

Comments

1 INDIA INK CONTAMINANT
2 SMALL APERTURE
3 BLANCHING AND THERMAL
4 NO SMOKE

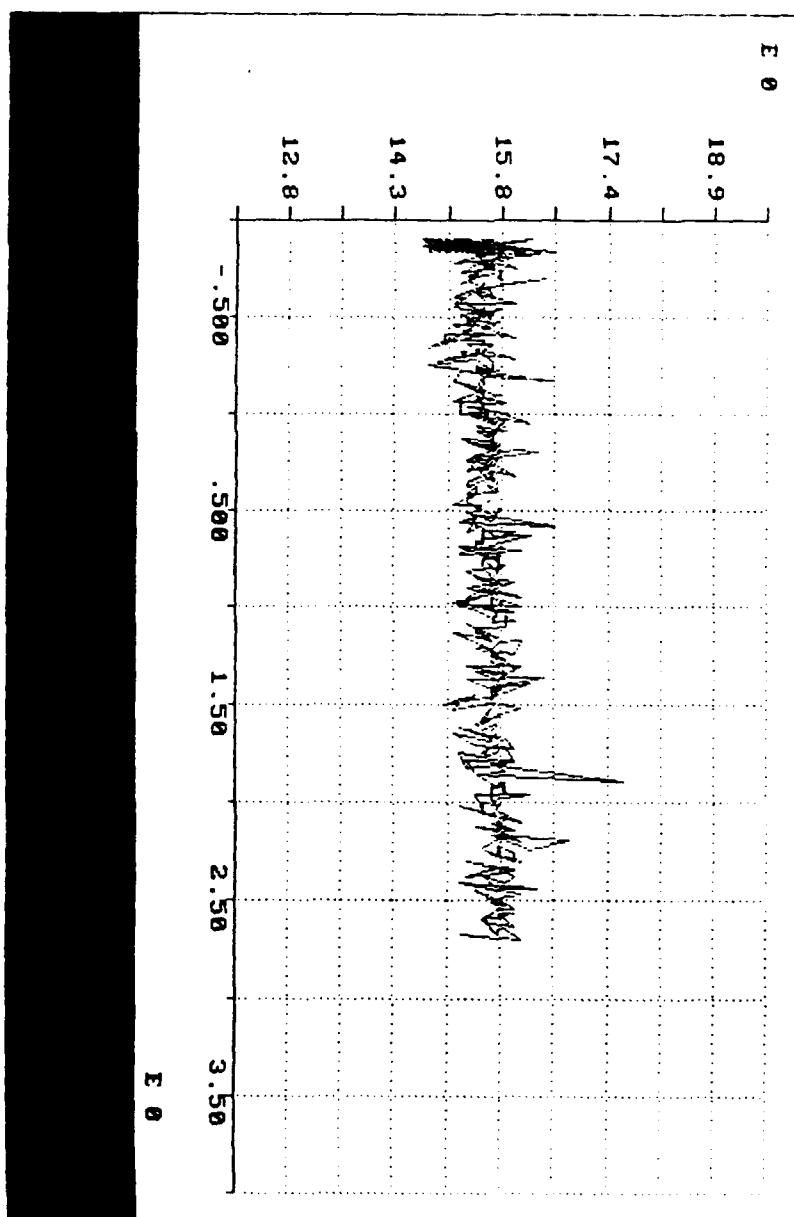
5 DID NOT LOAD IMAGE

6

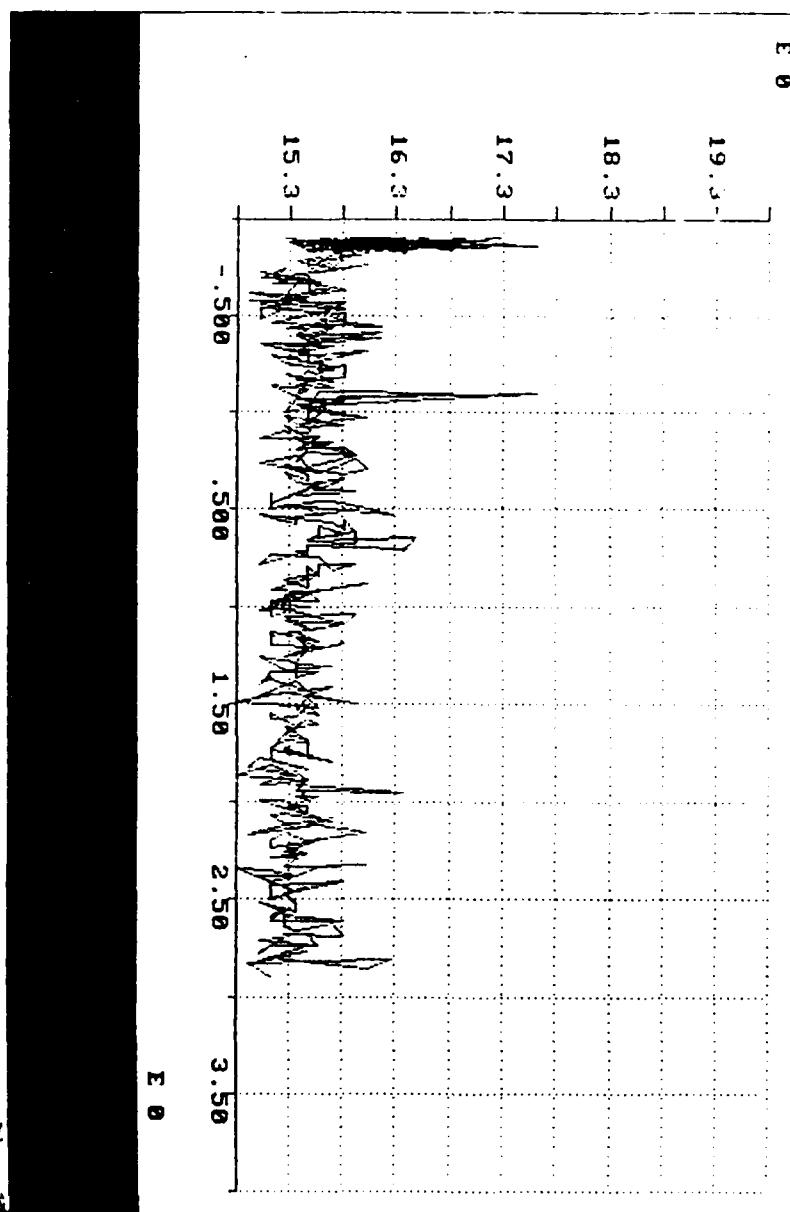
I/O Variable F R = Z 0

Subfile: 1 Row (0=all): 0 Start Column: 1 # of Columns: 256

Read/Write/Append/Replace/Format/Print/Save/Load

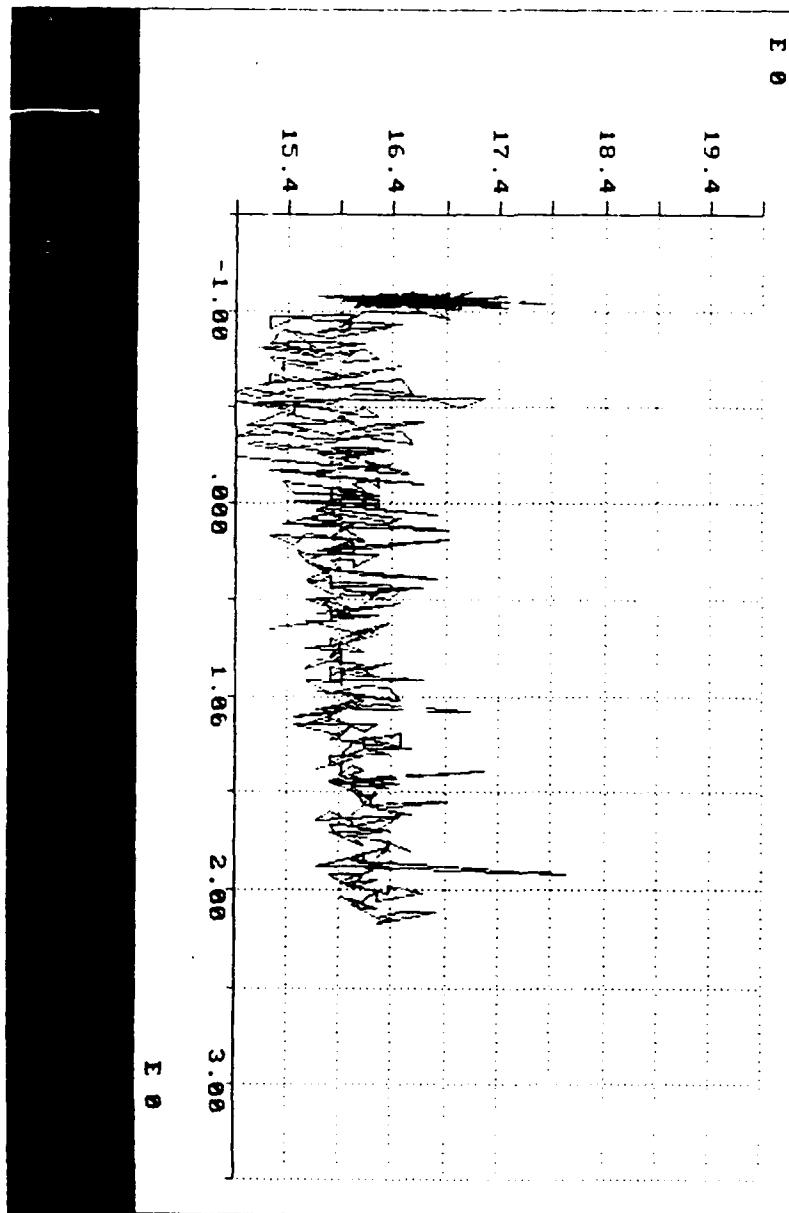


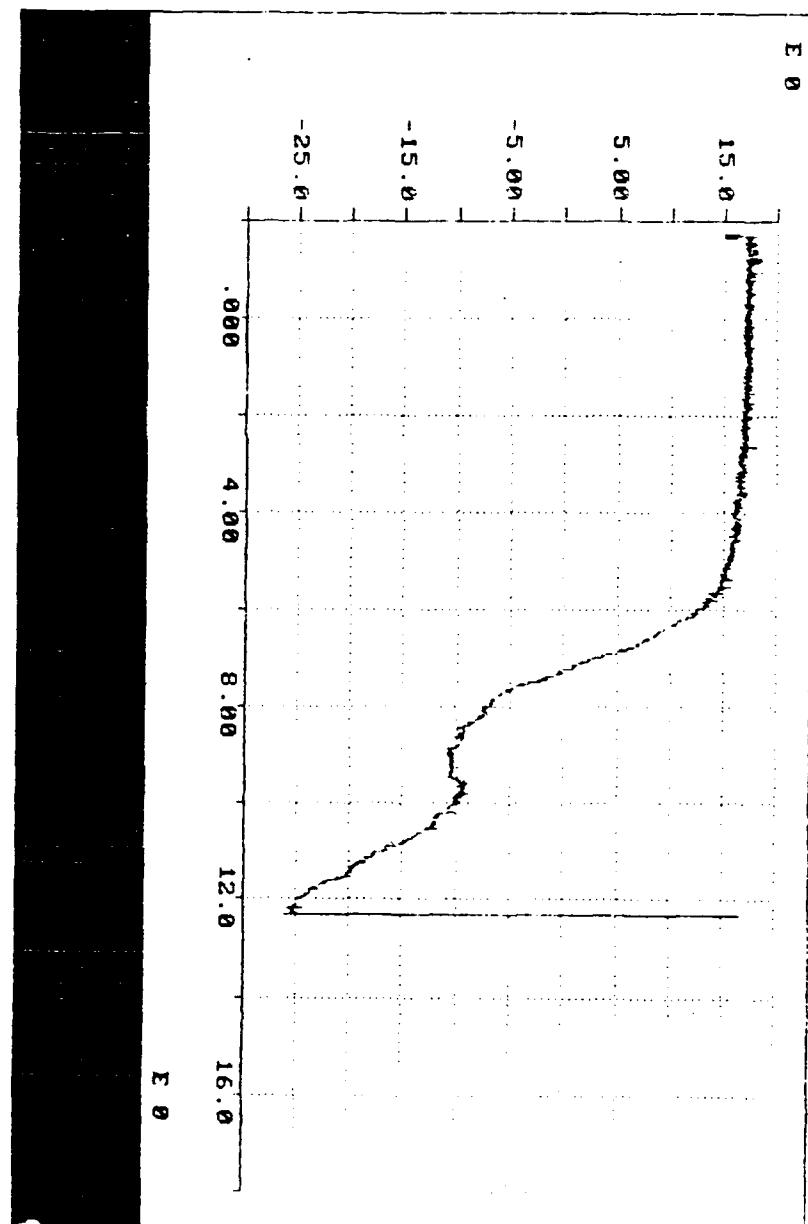
```
: File Name: C:\SEF\14\14.DAT          Output Device: 2
:
: 8 Comments
:
: 1 TRINA INC. COMPUTER SYSTEMS
: 2 SMALL ALERTS
: 3 CRASHED
: 4 STOP
:
: 5
: 6 MEDIUM ALERT - NO ERROR
:
: 7
:
: I/O Variable (R = Read):
:
: Subfile: 1          File: 14.DAT Length: 512
: Row (0=all): 0
: Start Column: 1
: # of Columns: 256
:
: Read/Write/Append: Read/Append/None
```

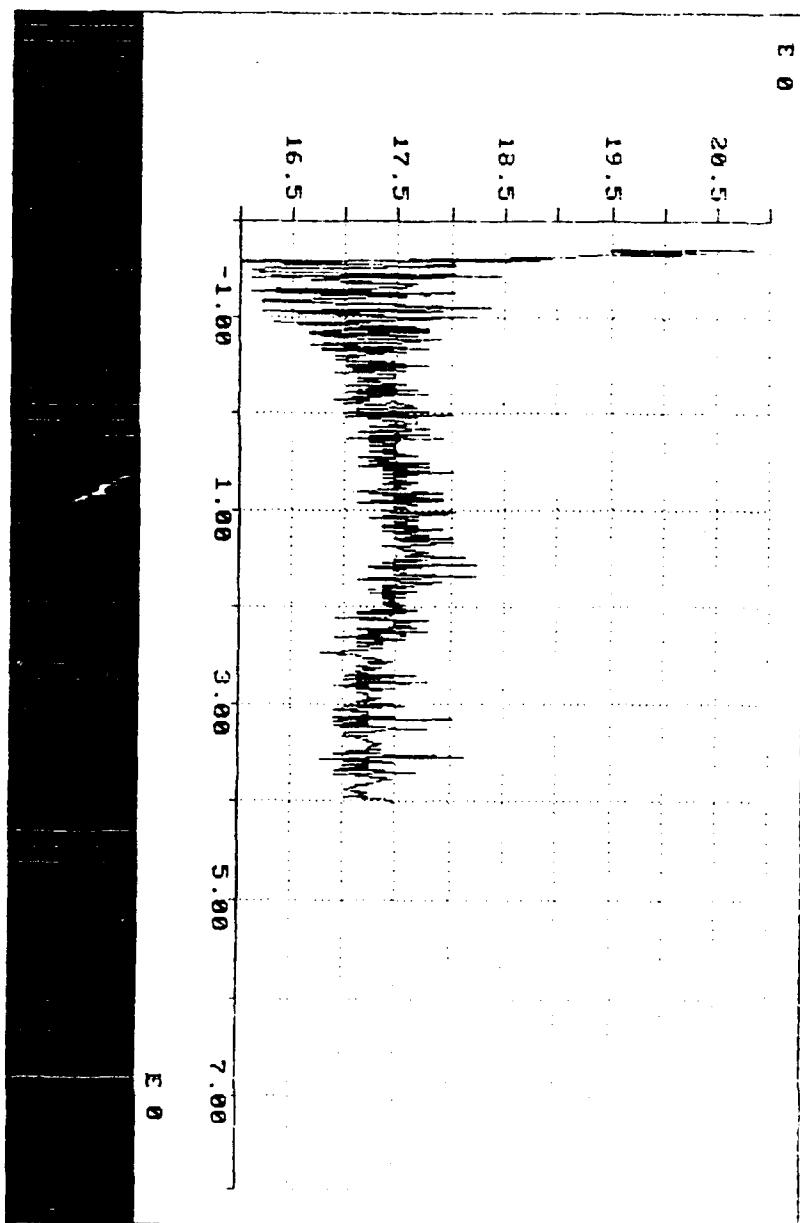


```
File Name: C:\...\INDIA.DAT
E Comments
LINDIA.DAT CONTAINS THE INDIA DATA SET FROM THE 1990 U.S. CENSUS.
E EMAIL: AF.EXTREME
E ELANERIN@AOL.COM
E IN STATE
E
E WEIR, JEFF D
E

INDIA Variable (1) 1990 U.S. CENSUS
E Profile: 1
E Row (Recall): 1
E Start Column: 1
E # of Columns: 1
E Read/Write/Ruplicate: R
E
```







File Name: C:\F1\T1.DAT

C Comments

Two entries (X,Y) per line, separated by a space.

I/O Variable (E, T, R)

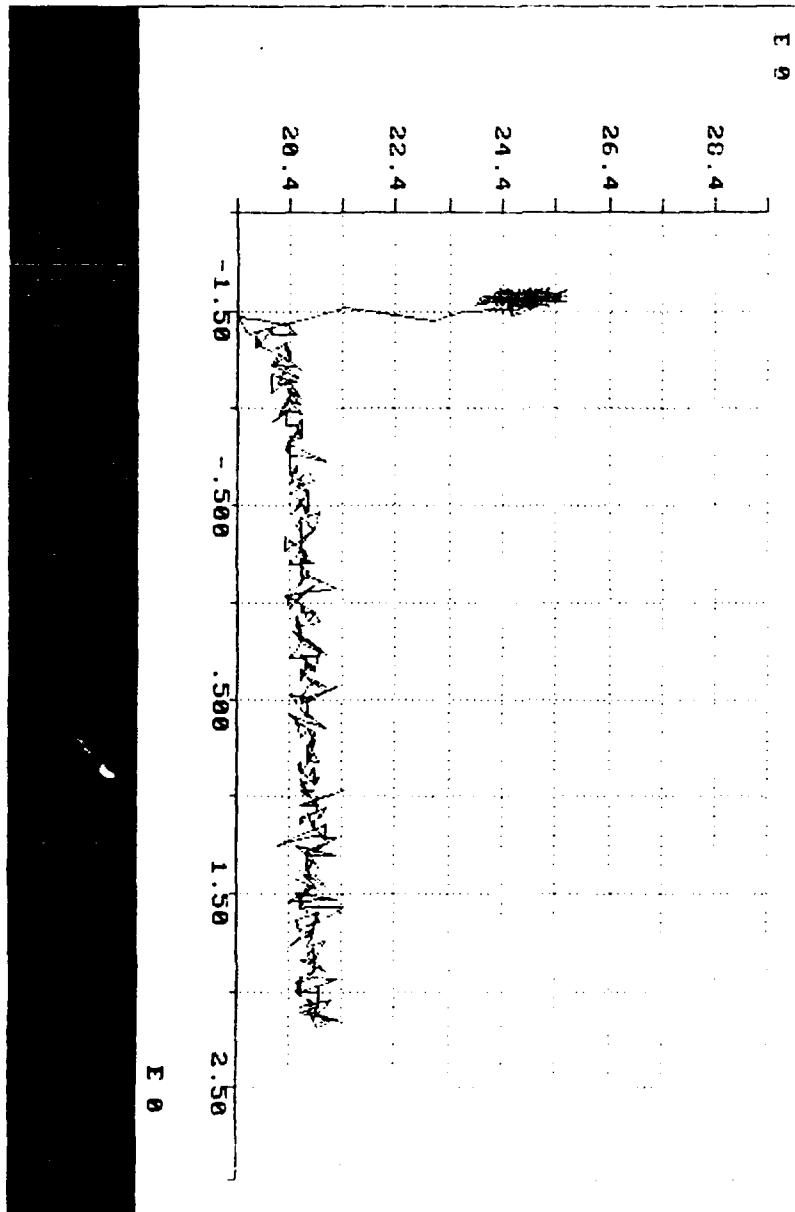
End File

Row (Index): 1

Start Column: 1

of Columns: 1000

Read/Write/Append/Overwrite/Replace/Reset/Close



XXXXXXXXXXXXXXXXXXXXXX

: File Name: C:\SP1\1.DAT

: 9 Comments

: 1 BY ICG CONTROLLED BY E

: 2 NO VISIBLE CHIMNEY

: 3 NO SMOKE

: 4 WIND

XXXXXXXXXXXXXXXXXXXXXX

: 5 10 Variable = 1

: 6 Subsite: 1

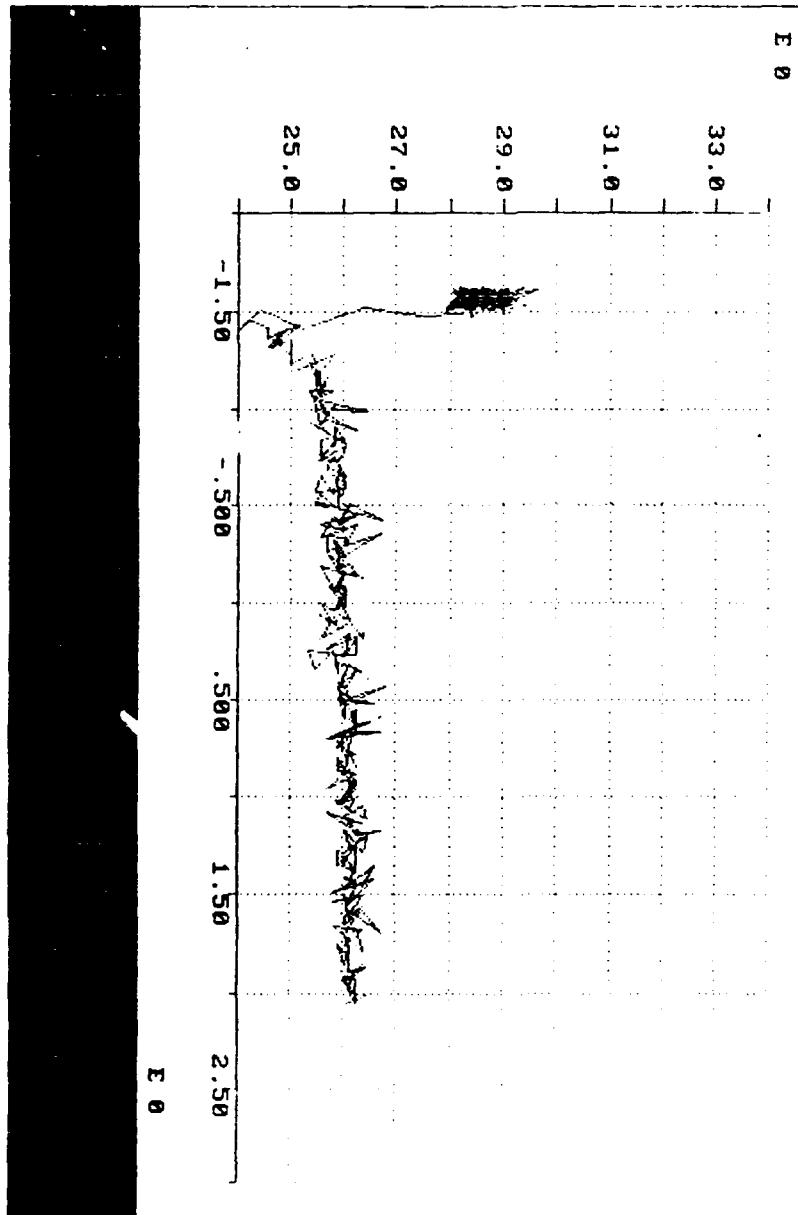
: 7 Row (0=all): 0

: 8 Start Column: 1

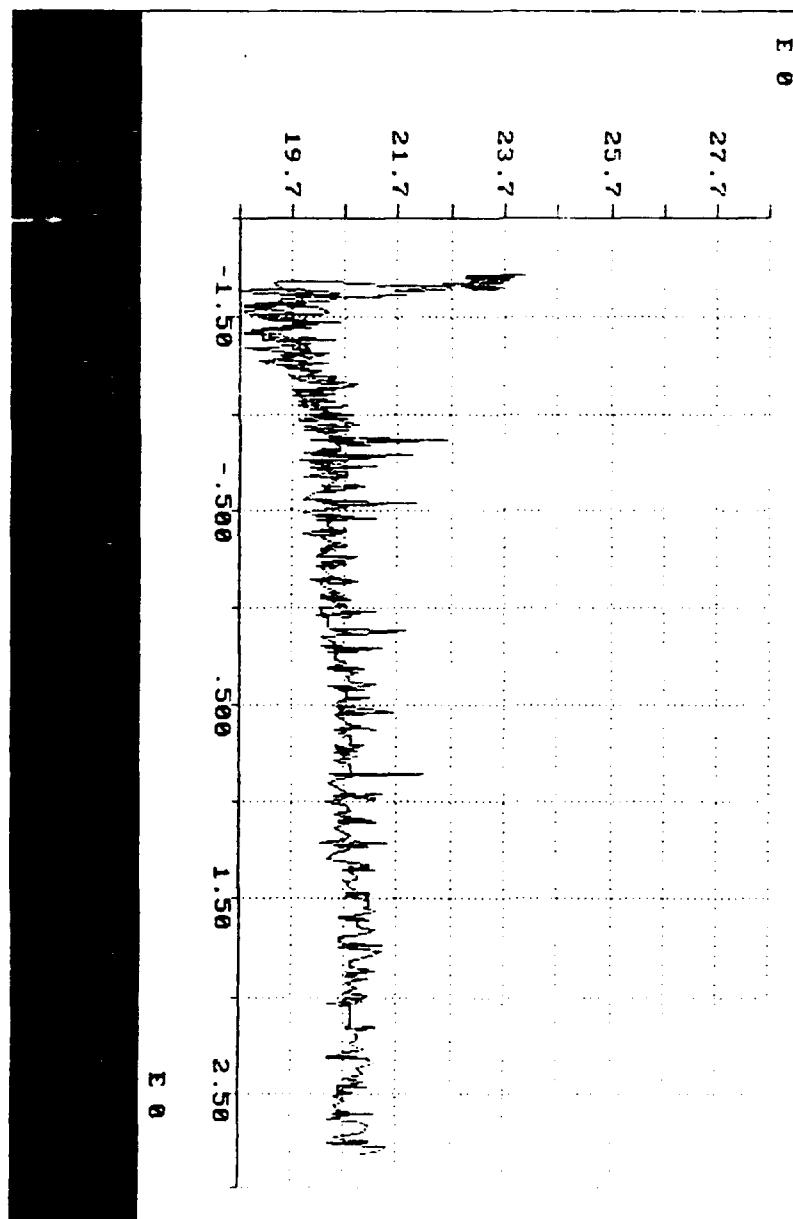
: 9 # of Columns: 2000

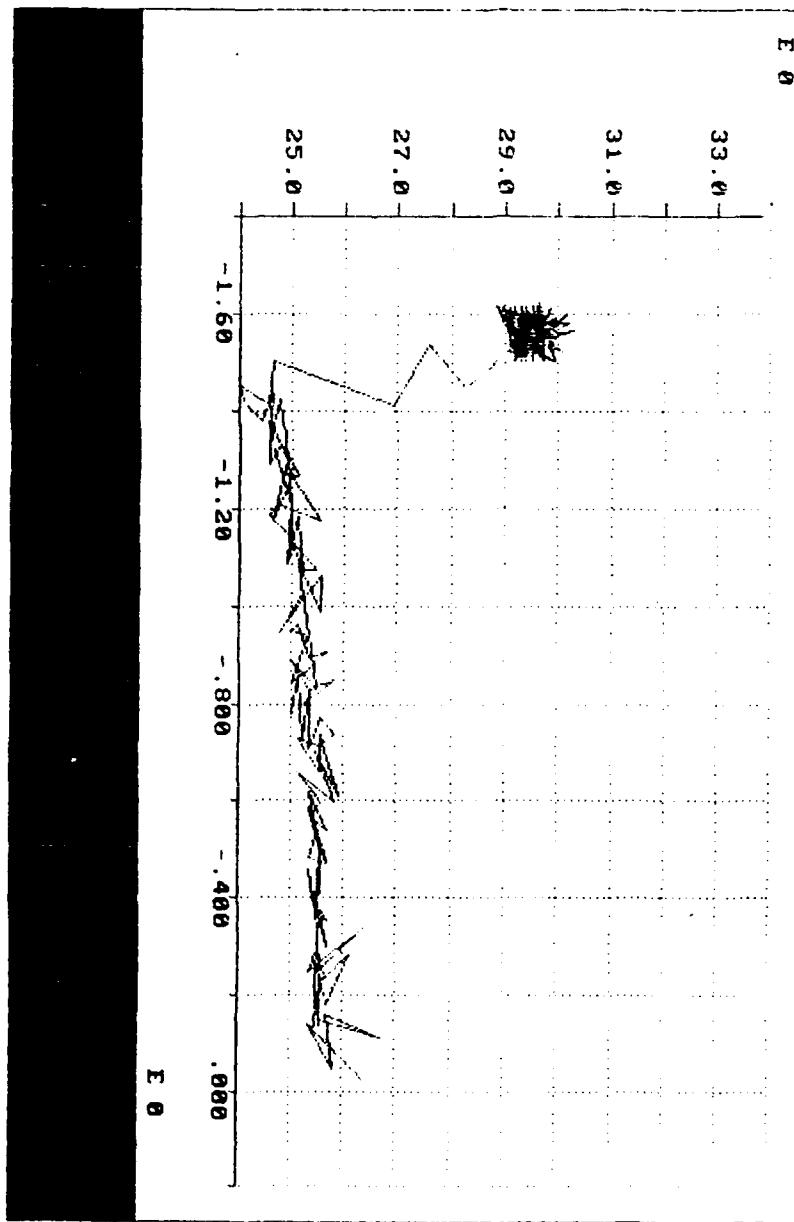
: 10 Read/Write/Append/Print/Exit/Erase/Save/Load/Plot/Graph/Help/Quit

XXXXXXXXXXXXXXXXXXXXXX



File Name: ESRF191.D
Comments:
1. FERRONYL+BLOOD CUFFED TO STOMACH
2. MINIMAL BLANCHING FROM CUFFED AREA
3. NO SMOKE
4. LESS CHROMOGENIC THAN PREVIOUS
5. WELDED, BUT NO
6. ONLY SEROUS SECRETION
7.
8.
9.
10. V ribble (S)
Subfile: 1
Row (0=all): 0
Start Column: 1
of Columns: 256
Read/Write/Append/Screen/1st/1st/1st/1st/1st/1st/1st/1st





REFERENCES AND NOTES

— 1 —

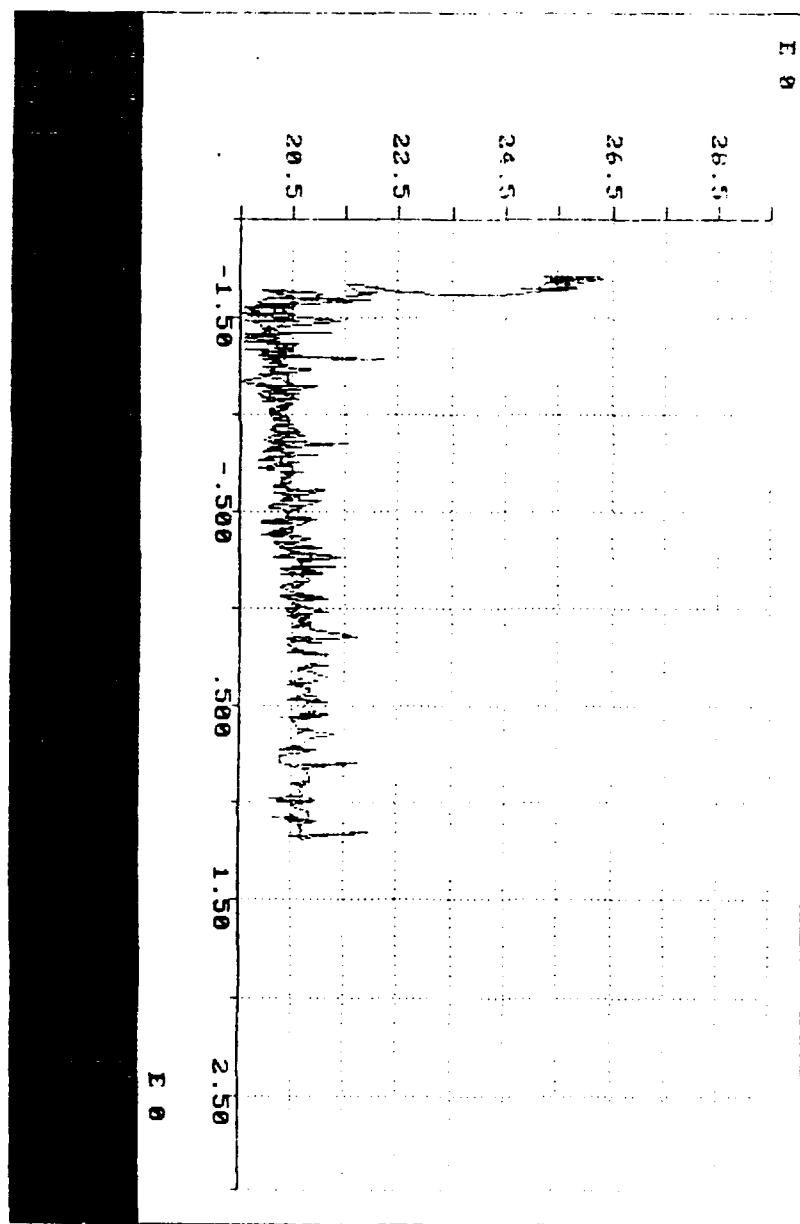
Journal of Health Politics, Policy and Law, Vol. 37, No. 1, January 2012
DOI 10.1215/03616878-37-1 © 2012 by The University of Chicago

For the first time, we have been able to measure the effect of the magnetic field on the energy loss function of a single molecule.

Format **Format** **Format**
Format **Format** **Format**

Read/Write Accessible Text in Full Text and Text-to-Speech API

Digitized by srujanika@gmail.com

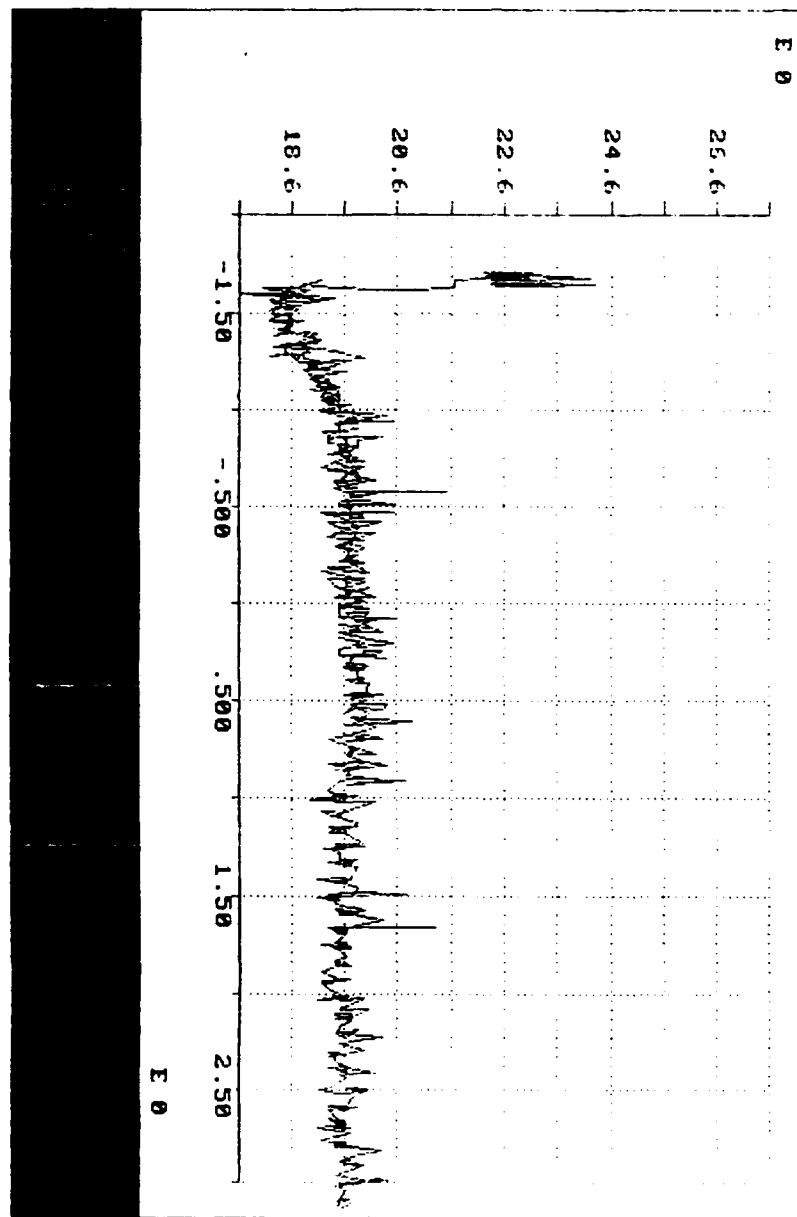


• File Name: C:\HARDWARE\PC\PC-1000\PC-1000.DAT
• Comments:
• 1 BURNDY WIRELESS
• 2 NO VISUALS CHARGE
• 3 NO SMOKE
• 4
• 5 NO WELD

¹ See, e.g., *State v. Gandy*, 100 N.W.2d 621, 624 (Iowa 1959) (“[T]he right to a trial by jury is a fundamental right which cannot be abridged.”); *State v. Johnson*, 100 N.W.2d 621, 624 (Iowa 1959) (“[T]he right to a trial by jury is a fundamental right which cannot be abridged.”).

```
# Subplot:          1
# Row (0=all):    0
# Start Column:   0
# # of Columns:   2x2
# Read/Write/Append/Skip all of local data from file and return it as
```

10. The following table summarizes the results of the study. The first column lists the variables, the second column lists the sample size, and the third column lists the estimated mean.



Journal of Management Education, Vol. 35, No. 7, November 2011, pp. 895–911
ISSN: 1052-5025 print / 1098-2633 online
DOI: 10.1177/1052502511415320
<http://jme.sagepub.com>

MORNING: 10:00 A.M.

2 Comments

Leptin and leptin receptor signaling in the brain: implications for the regulation of energy balance, metabolism, and behavior.

BLOOD ALONE CONT'D

• 2 • SAFETY PERCEPTION

2 NOVEMBER

2.6. $\langle R^2 \rangle$ AND $\langle H^2 \rangle$

S. S. WILDE

• • • • • • • • • • • •

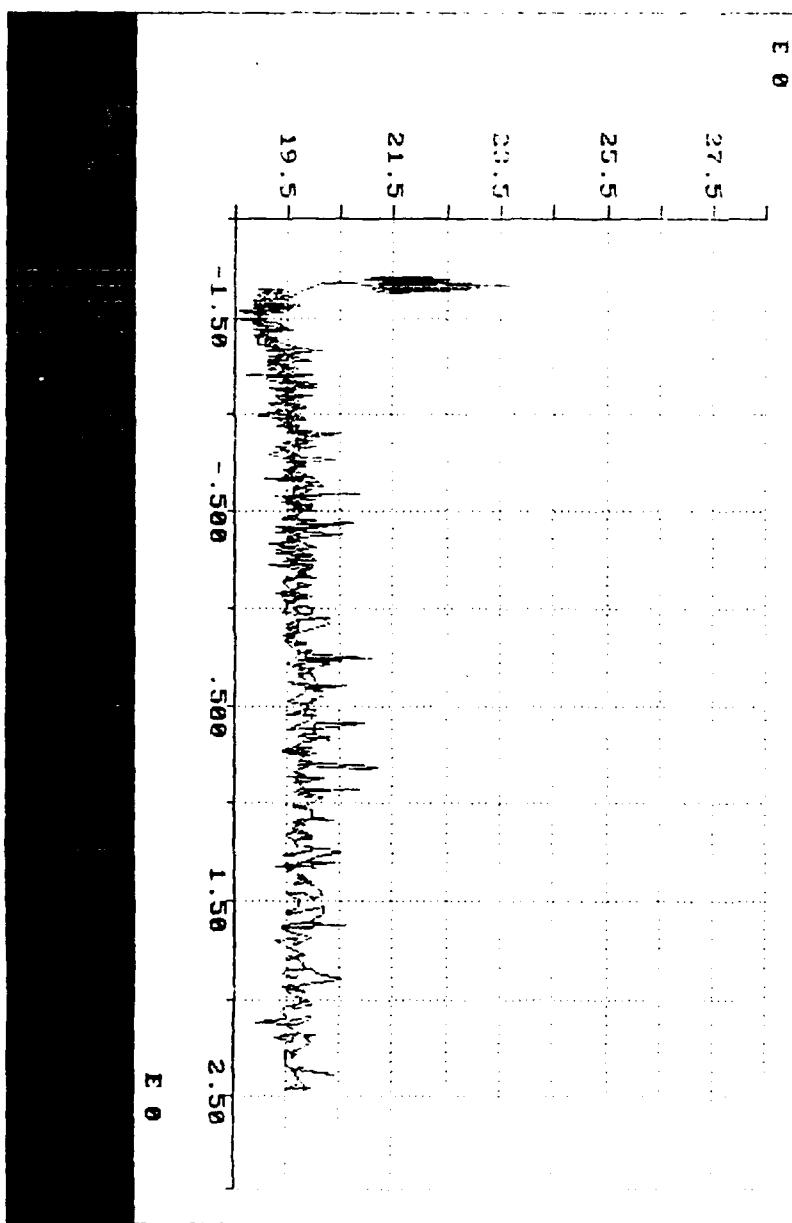
¹ See also *International Criminal Court, Statute of the International Criminal Court*, art. 12(2)(b), 1998, available at http://www.icc.int/iccwebportal/jsp/icc/statute/statute_en.jsp.

• On Marginalia •

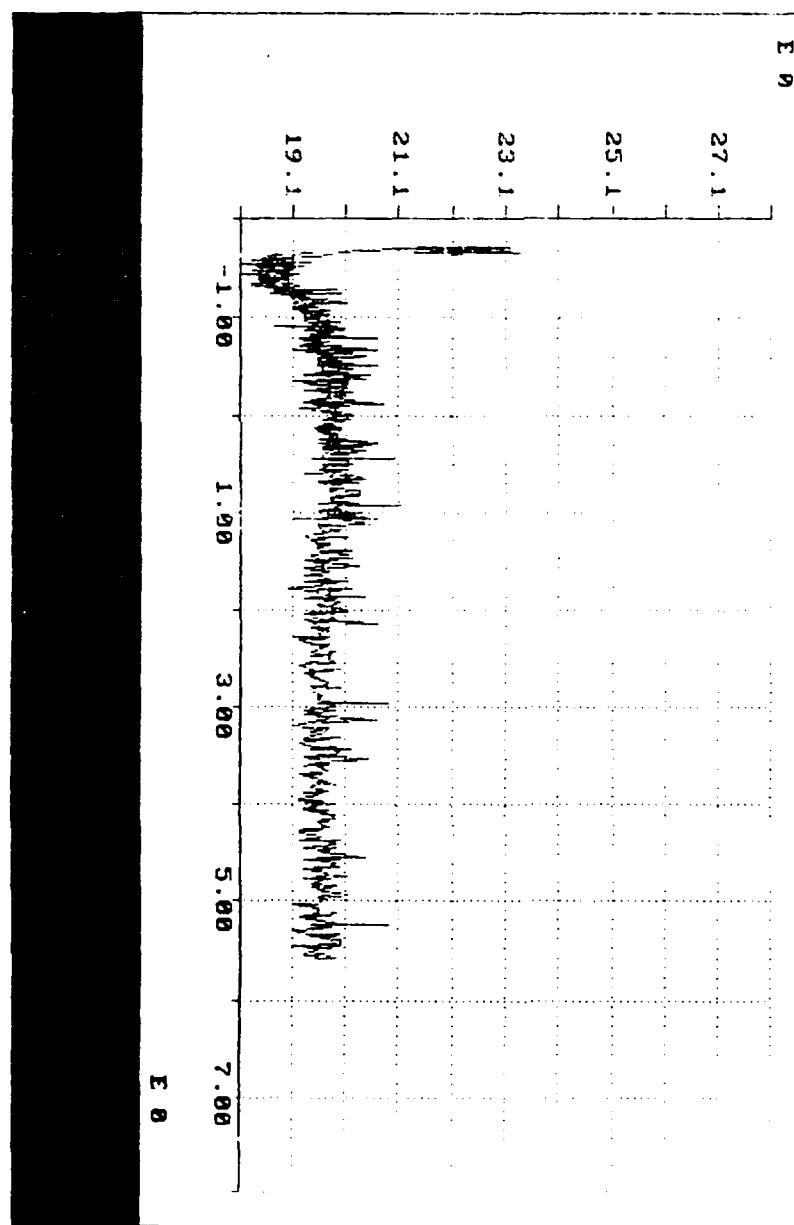
: Subtitle

; Row (0=ali):

Start Column: 1



File Name: C:\SST\101.10
Comments:
1. CLOUD ALONE SET
2. TOTAL EXPOSURE TIME
3. DAY/NIGHT
4. DESCRIPTION
5. SNOUT
6. TAIL WELD
7.
8.
9. # of Columns
10. Year/Month/Day
11. Column



File Name: E6.DAT

B Comments

1 BLOOD ALONE 50% CONC
2 THERM EXP GROW 100
3 THERM EXP GROW 100
4 THERM EXP GROW 100

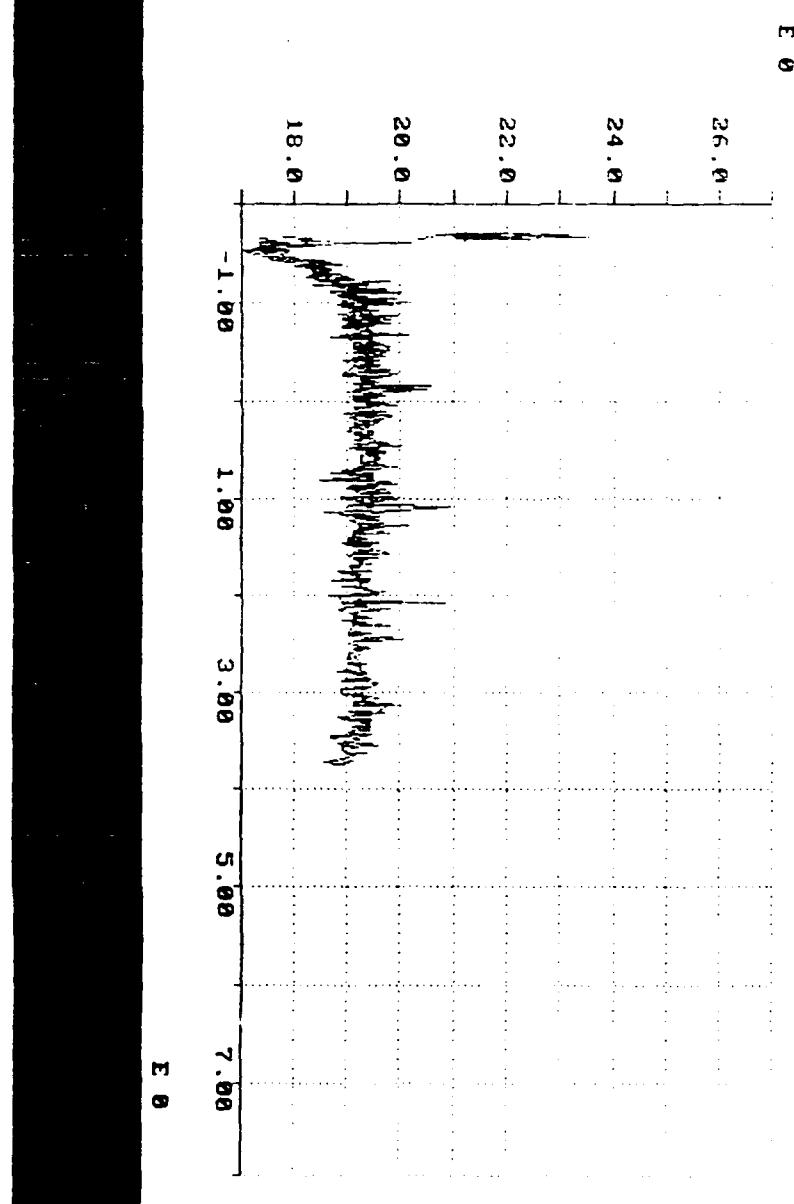
5 1000

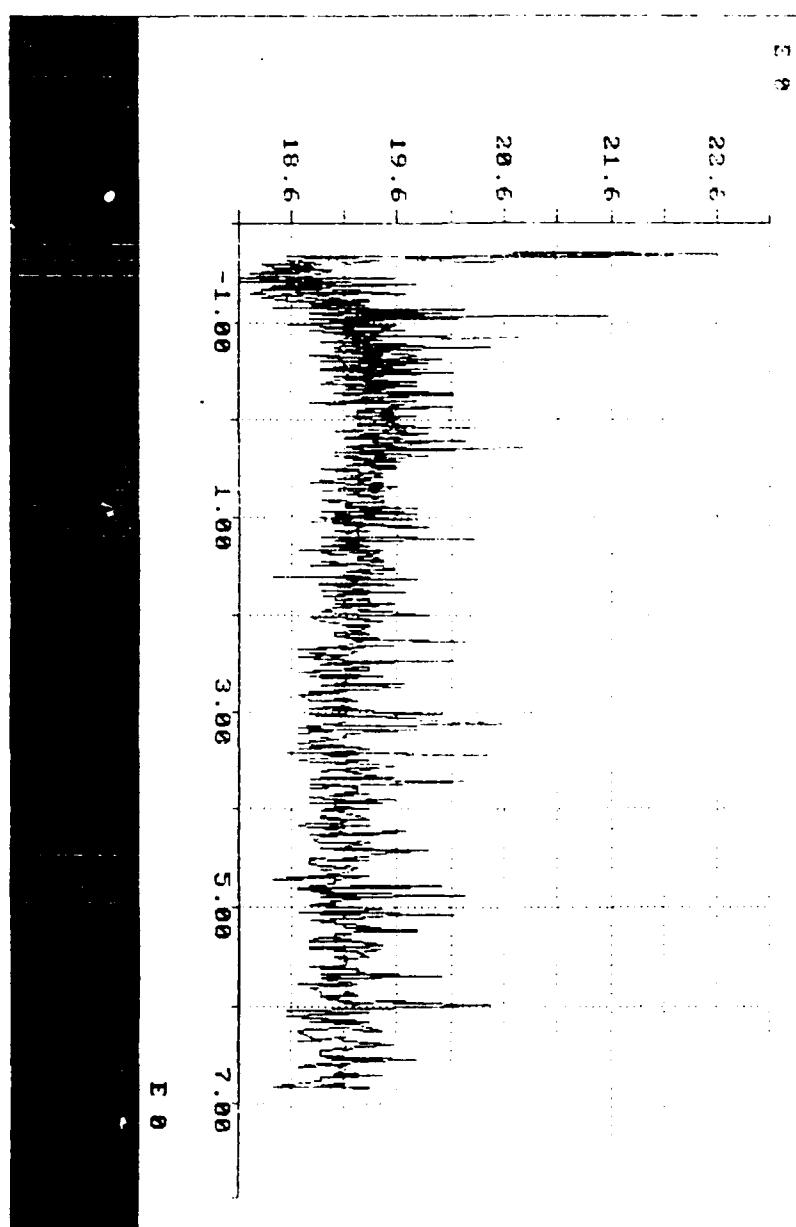
6 1000

7 RightSite 1
8 Row (Overall): 1
9 Start Column: 1
10 End Column: 1000

11 1000 1000 1000 1000 1000 1000 1000 1000 1000 1000

12 1000 1000 1000 1000 1000 1000 1000 1000 1000 1000





SCHÜLERARBEITEN

2. Klassifizierung

- a) UNDIDENGBARKEITSPROBEN UND DABEI ERGEBENDE ERKENNTNISSE
- b) FÜR DEN AUFTRAG VON DER ZEIT PEGE
- c) MÜNDLICHE PRÄSENTATION VON DER ZEIT PEGE

a) UNDIDENGBARKEITSPROBEN
ZUR ZEIT PEGE

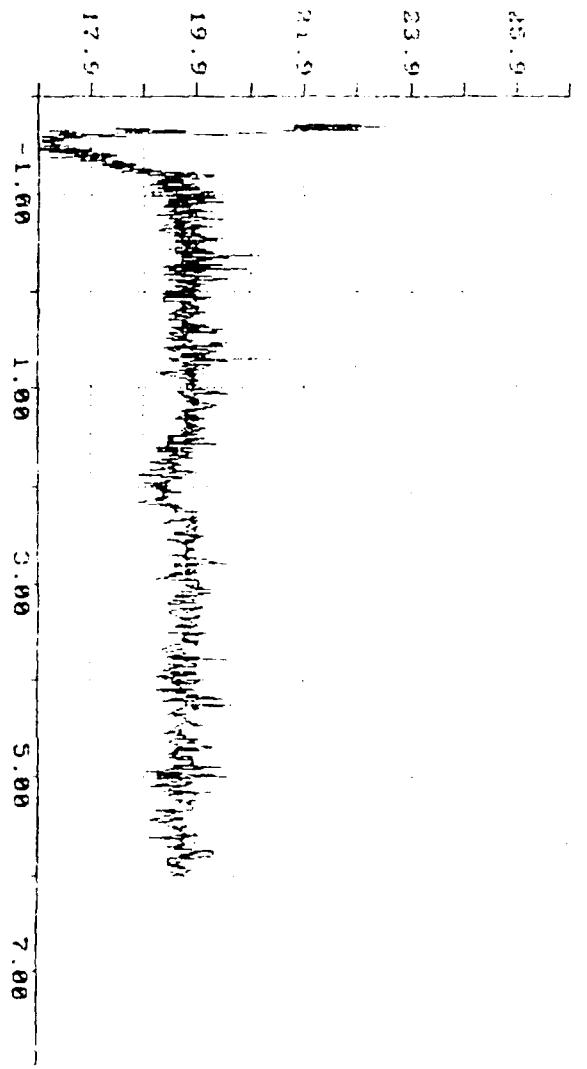
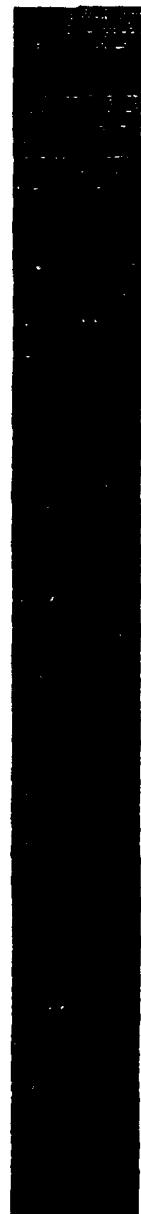
zur Zeit PEGE

ZUR ZEIT PEGE

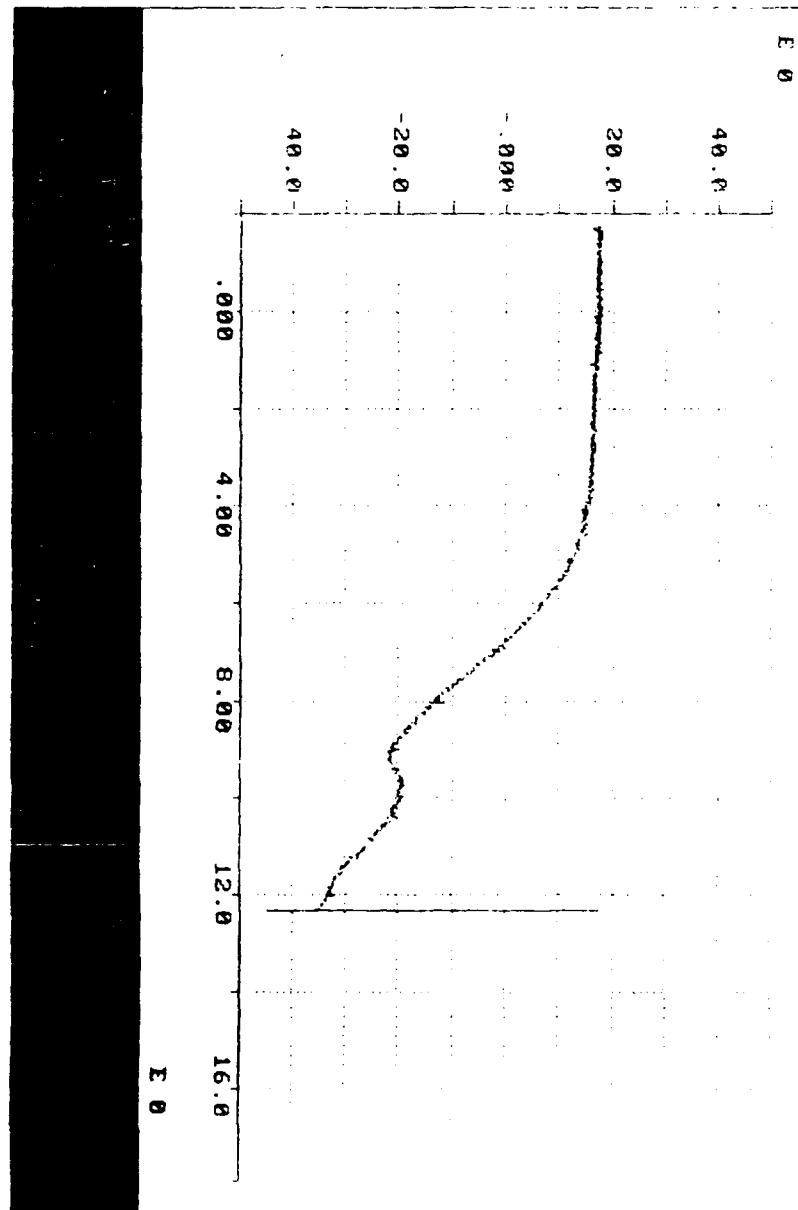
- a) Der Name
- b) Der Name
- c) Die Umgebung

zur Zeit PEGE

zur Zeit PEGE



1.0000000000000000E+000



E 6

16.6

72.0

20.0

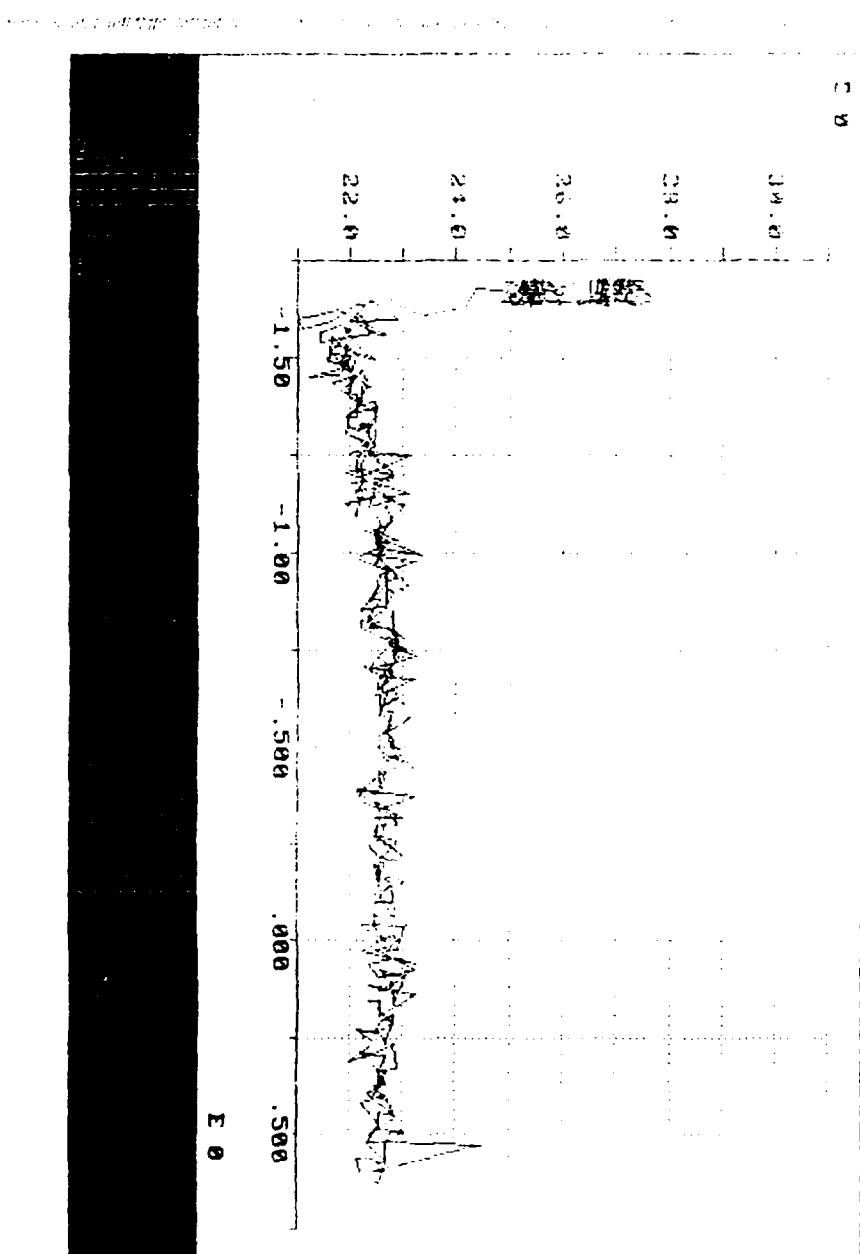
24.6

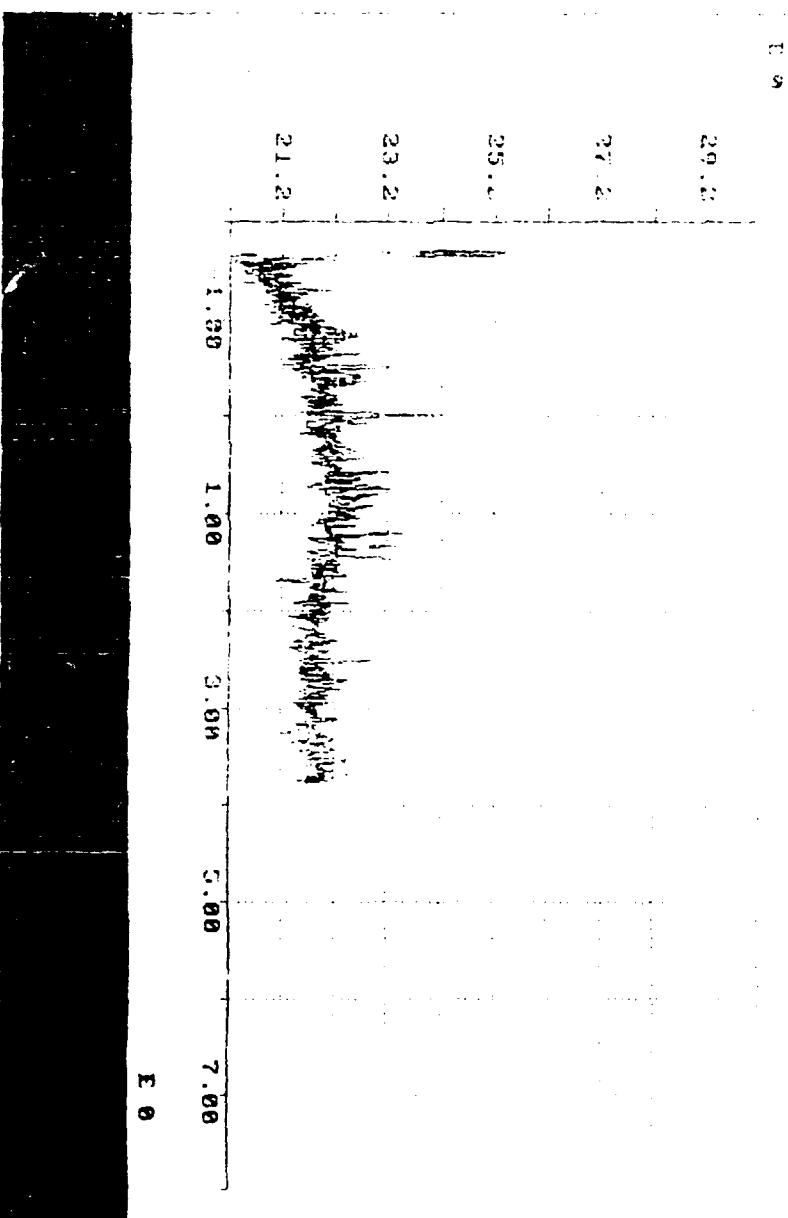
20.2

-1.00	1.00	3.00	5.00	7.00
-------	------	------	------	------

E 6

1.000
0.900
0.800
0.700
0.600
0.500
0.400
0.300
0.200
0.100
0.000





E 0

RECORDED BY DRAFTING TABLE

100% SENSITIVITY

100% LINEARITY

100% ACCURACY

RECORDED BY DRAFTING TABLE

100% SENSITIVITY

100% LINEARITY

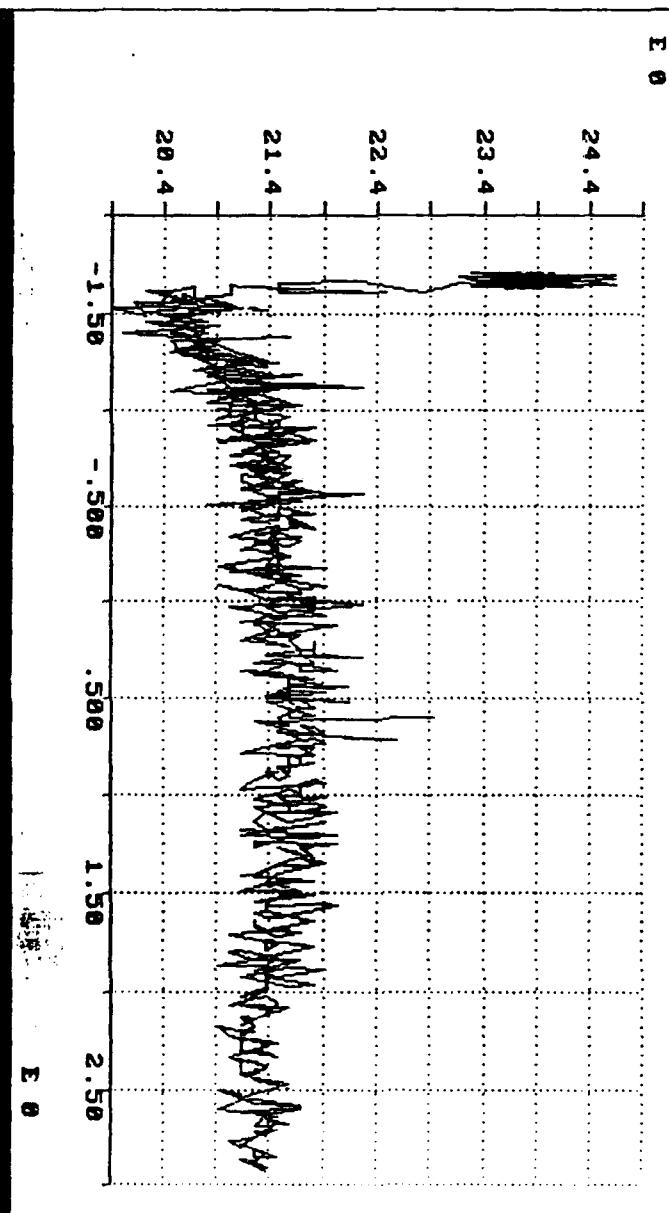
100% ACCURACY

RECORDED BY DRAFTING TABLE

100% SENSITIVITY 100% LINEARITY 100% ACCURACY



E 0



¹ See, e.g., *United States v. Bivens*, 446 U.S. 713 (1980), and *Ex parte Young*, 283 U.S. 275 (1932).

• miles frequent: 1000-1500 miles per month

2. Measurements

- 1 ONE STRIP UNWOUND FROM THE REEL
• 2 EGGES AT THE END OF THE STRIP

ENRICO PIAZZA - www.enricopiazza.it

¹ See also the discussion of the relationship between the concept of "cultural capital" and the concept of "cultural value" in the introduction.

• Variable (F) = $\frac{S_{\text{between}}}{S_{\text{within}}} = \frac{10.0}{1.0} = 10.0$

• [Sustentabilidade](#) • [Inovação](#) • [Gestão](#) • [Marketing](#) • [Produtos](#)

1998 (Oxford): 1–16.

: Start Column: 1

: # of Columns: 25

- Read/Write/Append/Close file - Python file module

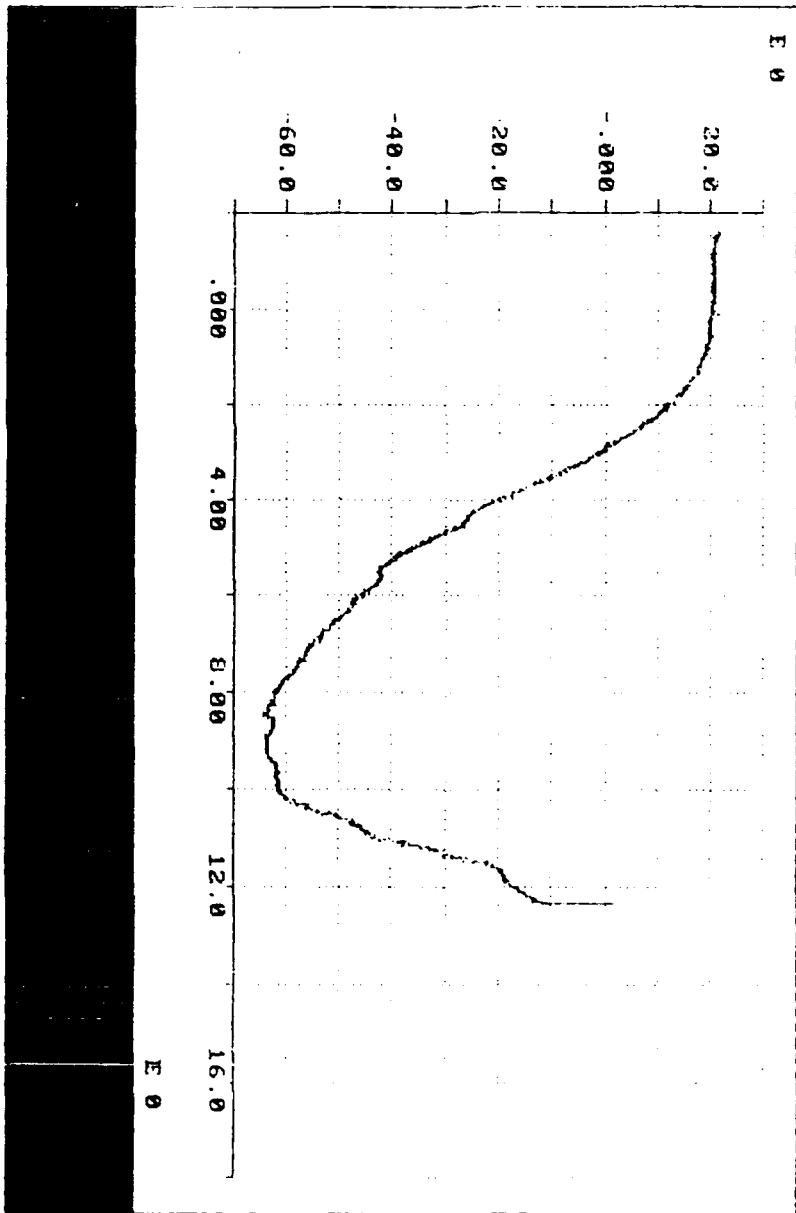
Digitized by srujanika@gmail.com

[View Details](#) | [Edit](#) | [Delete](#)

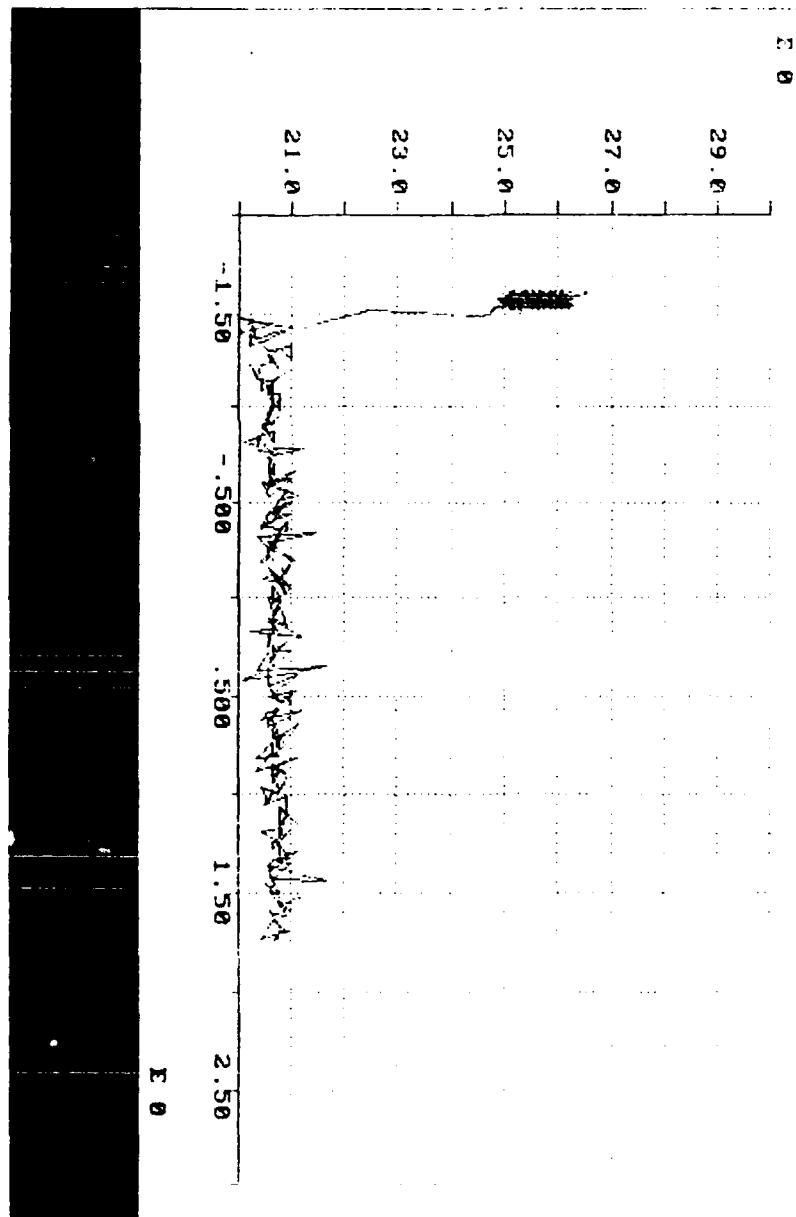
E

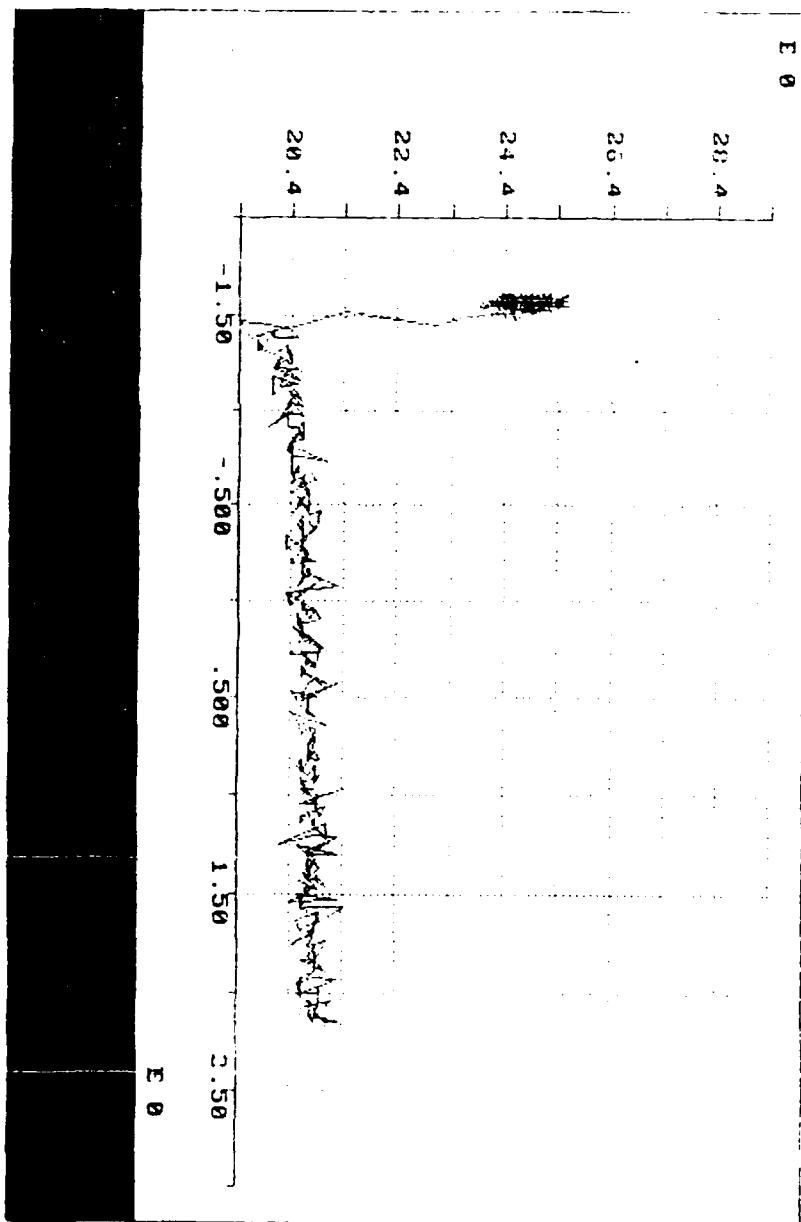
Digitized by srujanika@gmail.com

Digitized by srujanika@gmail.com

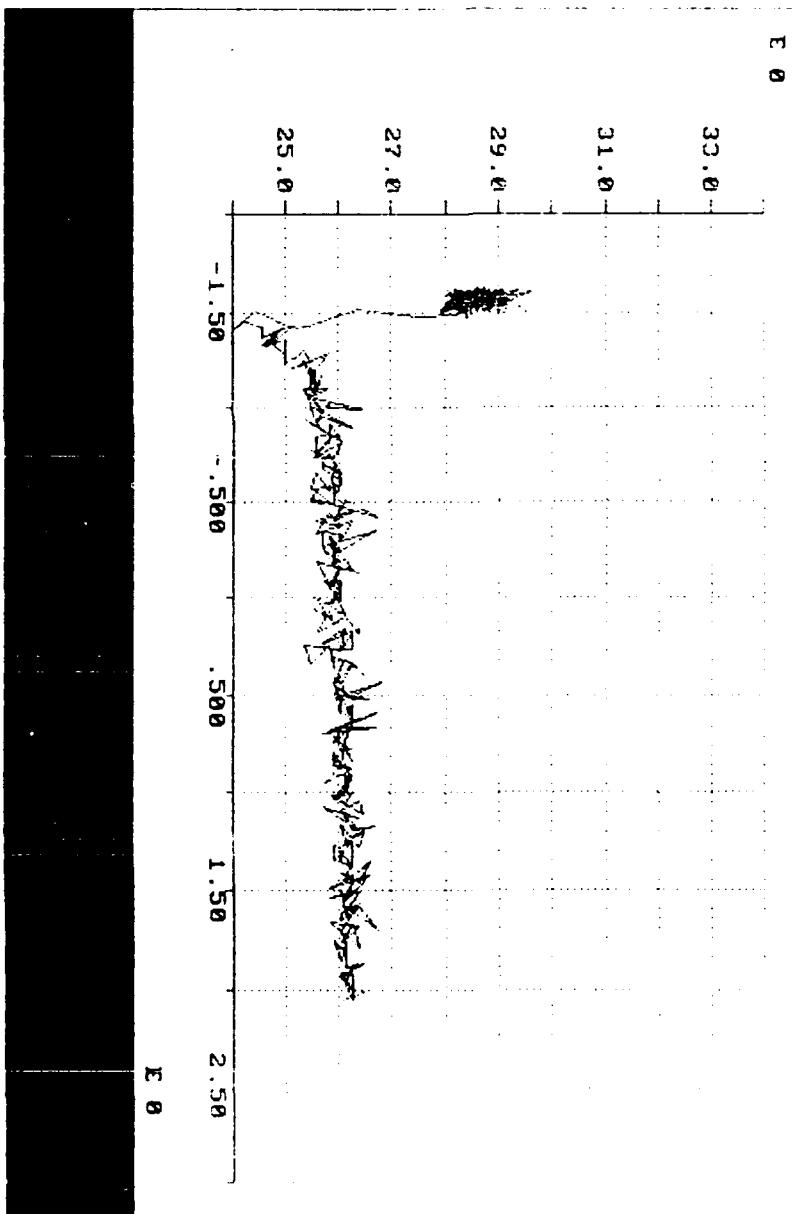


```
# File Name: C:\DE\1.FLT
# Version: 1.0
# Subtitle: 1
# Start Column: 1
# End Column: 200
# Read/Write/Append: R
# I/O Variable: C
```

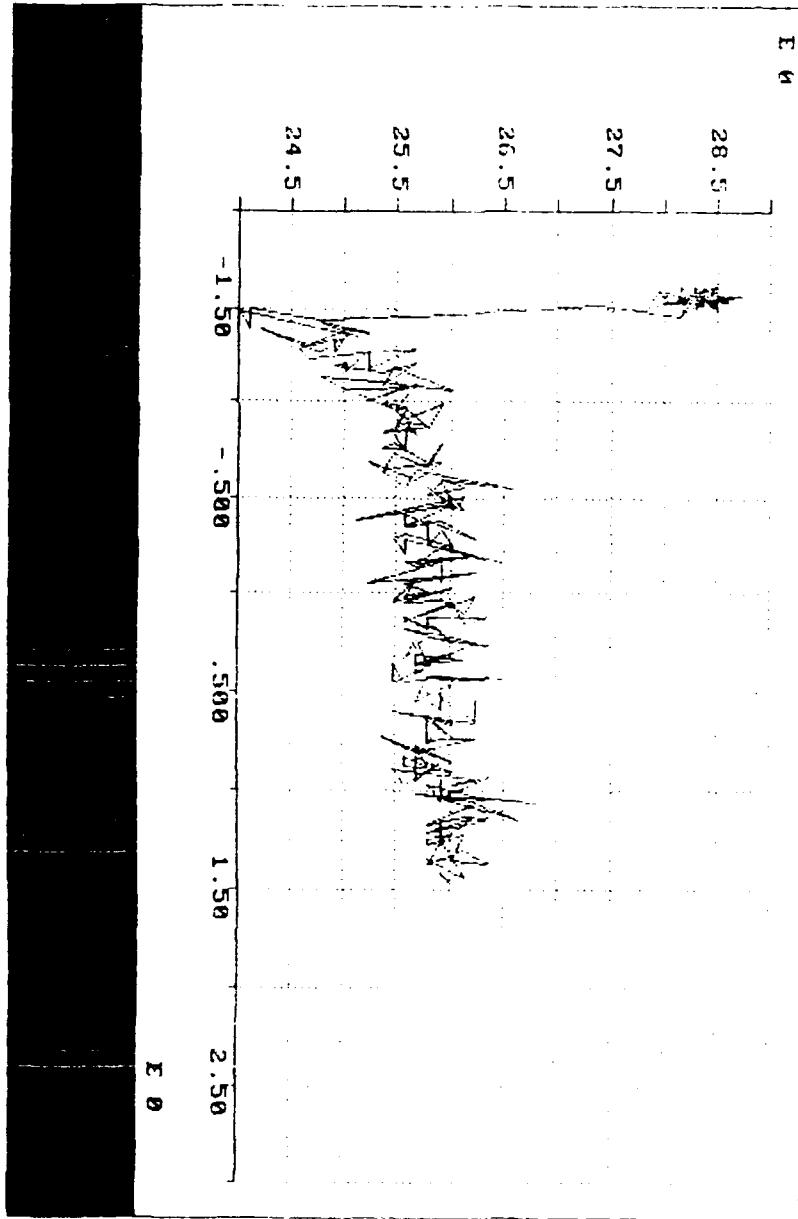




```
File Name: E001  
B Comments:  
• 1) ICD CONVENTIONAL  
• 2) VISIBLE  
• 3) GMORE  
  
B) NO WELD  
  
C)  
I/O Variable (E001)  
Subfile: 1  
Rows (Data):  
Start Column:  
End Column:  
Format/Width/Type: 10.3 10.3 10.3 10.3 10.3 10.3 10.3 10.3 10.3 10.3
```



File name: E01.DAT
Number of Elements: 1000000000
X & Y Dimensions: 1000000000
X & Y Visible Range:
X Min: 0.0000000000000000E+000
X Max: 1.0000000000000000E+000
Y Min: 0.0000000000000000E+000
Y Max: 1.0000000000000000E+000
Y Variable: 1.0000000000000000E+000
Section: 1
Row ID=1111
Start Column: 1
of Columns: 1000
Mean White Apprend: 1.0000000000000000E+000



XXXXXXXXXXXXXXXXXXXXXXXXXXXXXX

: File Name: C:\101.DAT

: Comment:

: FID 2X SENSITIVITY

: NO VIBRATIONS

: NO SHOCKS

: NO VELVET

: NO DUST

: NO VIBRATION DURING RECORDING

: NO DUST

: No spikes

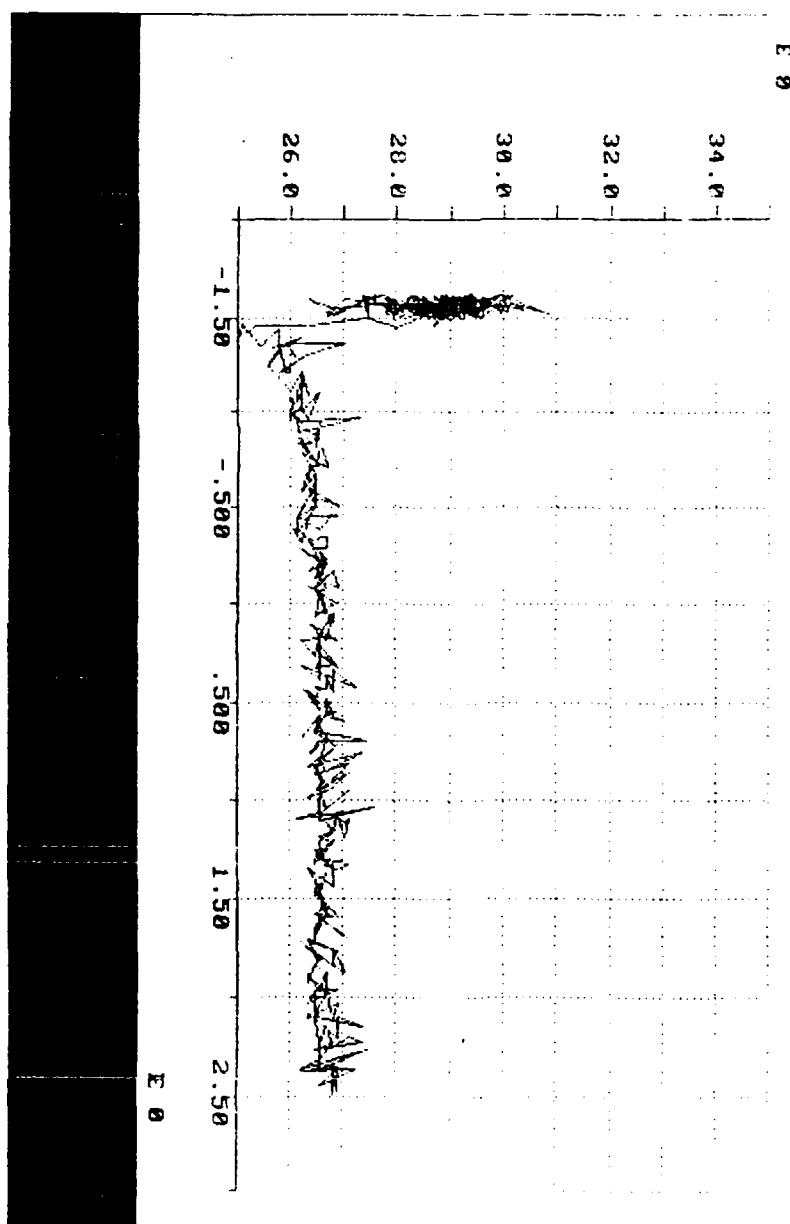
: Low noise

: Start column: 1

: # of Columns: 1000

: Read/Write/Append/Replace/Editor/Save/Print/Exit

XXXXXXXXXXXXXXXXXXXXXXXXXXXXXX



Journal of Health Politics, Policy and Law, Vol. 35, No. 4, December 2010
DOI 10.1215/03616878-35-4 © 2010 by The University of Chicago

• 14 • 長治縣志

Comments

ICG CY SET=100% BRIGHTNESS

— 1 —

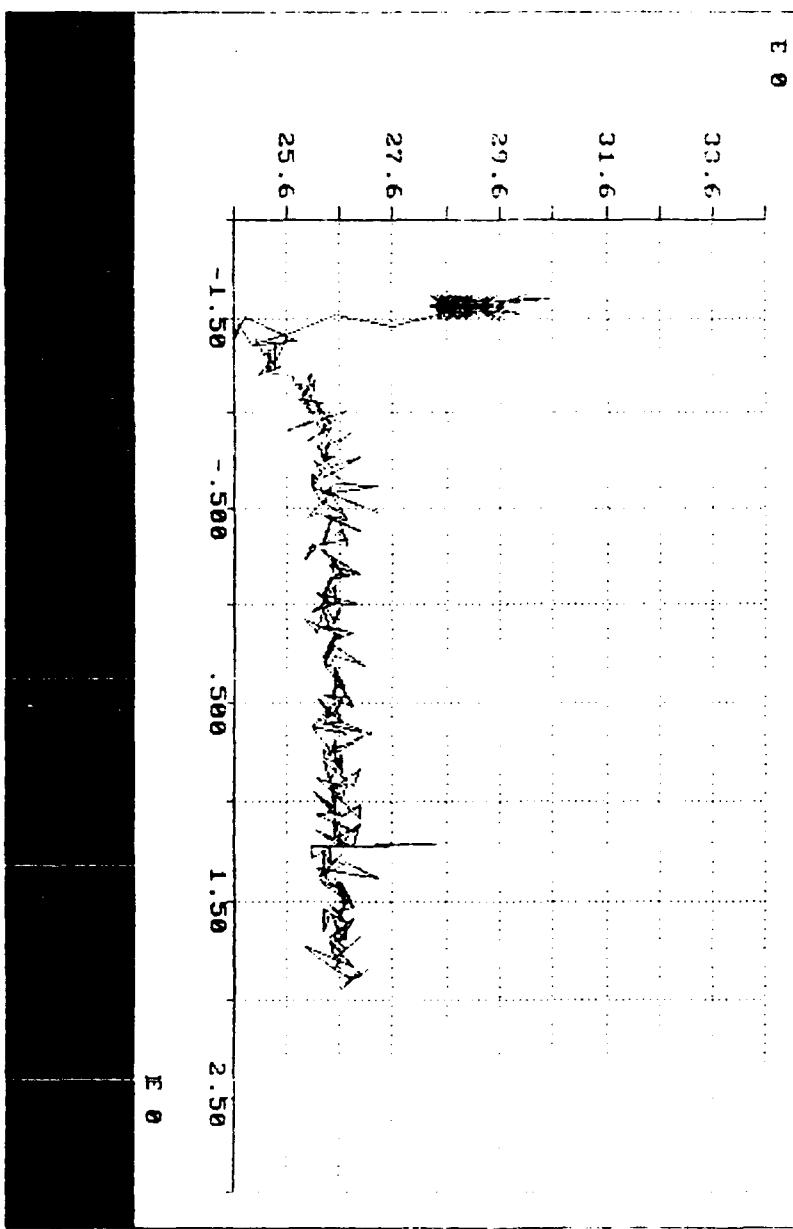
2. *Constitutive expression of the *hsp70* gene in *S. pombe* is controlled by the heat shock factor Hsf1.*

— 20 —

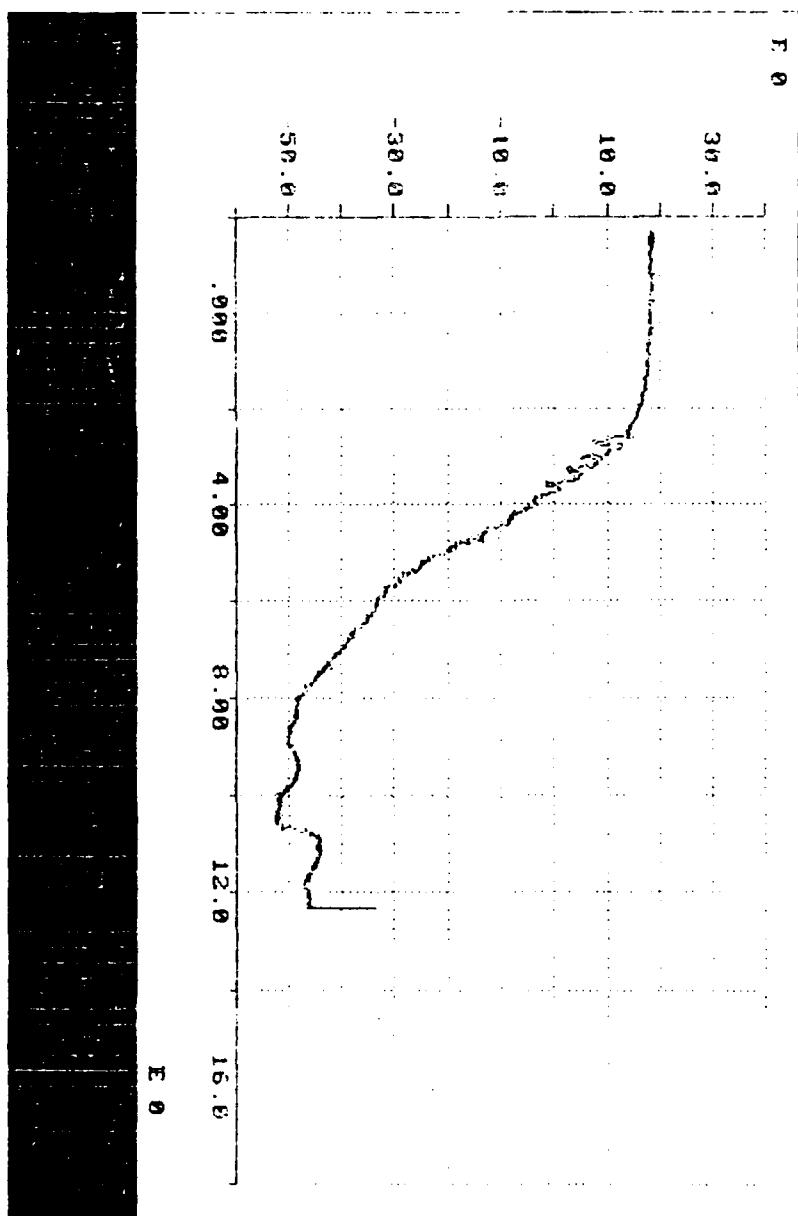
For more information about the study, please contact Dr. Michael J. Hwang at (319) 356-4000 or email at mhwang@uiowa.edu.

Plant Calwensis

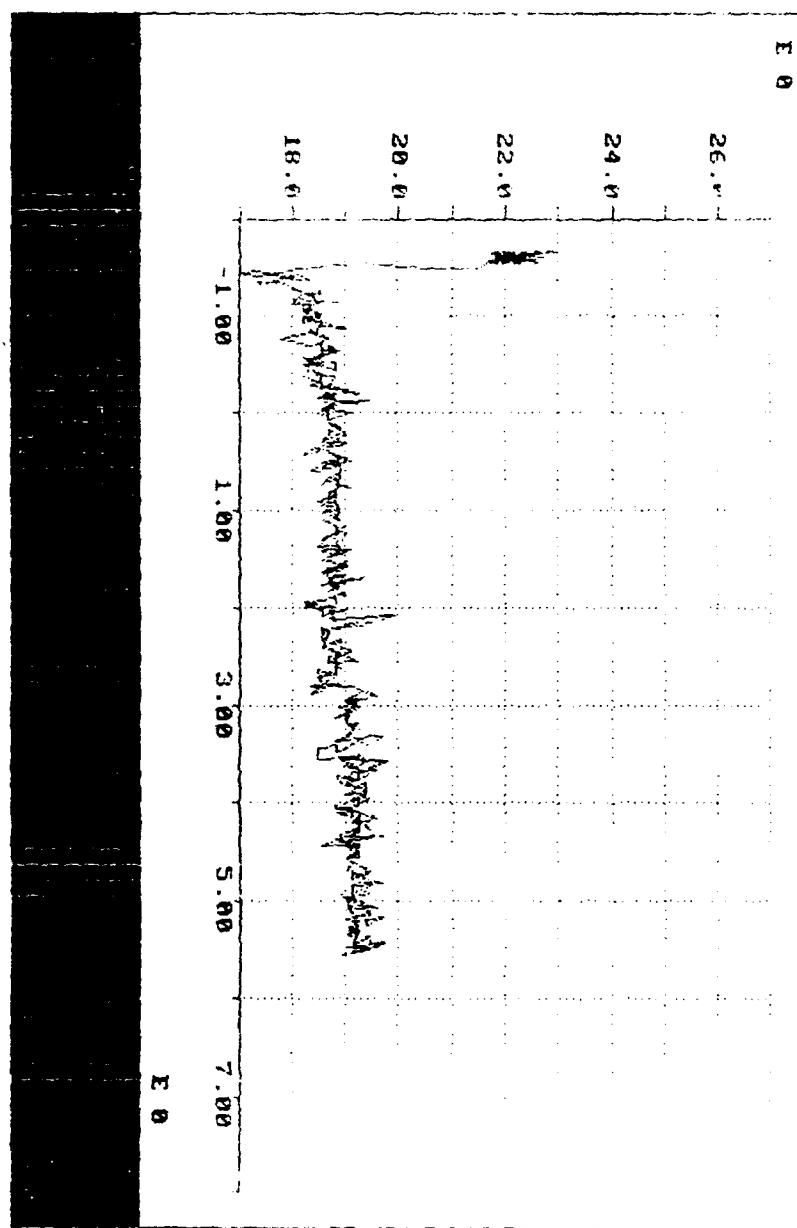
• 1996 年 1 月 1 日起，新規例將適用於所有在港營業的保險公司。



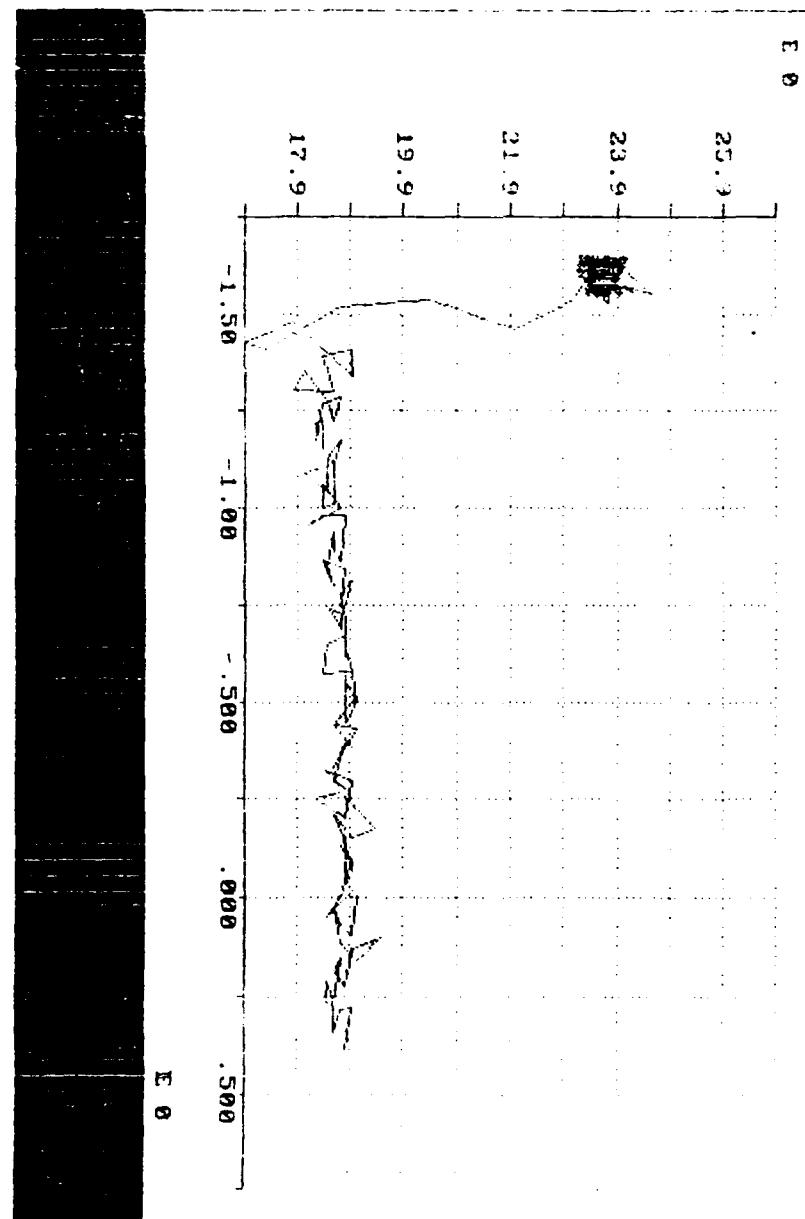

```
File Name: DATA.DAT
# Comments
# ONE STRIP CONVENTIONAL
# 2 SMOKE AT NO TIME POINTS
# 3
# 4
# 5
# 6
# 7
# 8
# 9
# 10 Variables
# 11
# Subfile:
# Row (0=all): 0-0
# Start Column: 1
# # of Columns: 100
# Read/Write/Appending: R
#
```



```
: File Name: EXEC.MAC
: Line Number: 1
: S Comments
: C
: C Two STEUB field
: L
: I
: J
: K
: L
: M
: N
: O
: P
: Q
: R
: S
: T
: U
: V
: W
: X
: Y
: Z
: I/O Variable (Type)
: Subfile:          Subfile number - 101
: Row (0=all):      0
: Start Column:     1
: # of Columns:     1
: Read/Write/Append: R
```



File Name: E0.DAT
Comments:
1 UNWELDED PI
I U Variable:
Subfile:
PCW (0=all):
Start Column:
of Columns:
Read/Write/Append: R



: File Name: K04SF021D.DAT

:
: 8 Comments

: 1> BLOOD CONTROLLED BY: : Sterile
: 2> NO VISIBLE CLEAVAGE
: 3> NO SMOKE
: 4>
: 5> NO WELD
: 6>
: 7>
: 8>

: I/O Variable (Type):

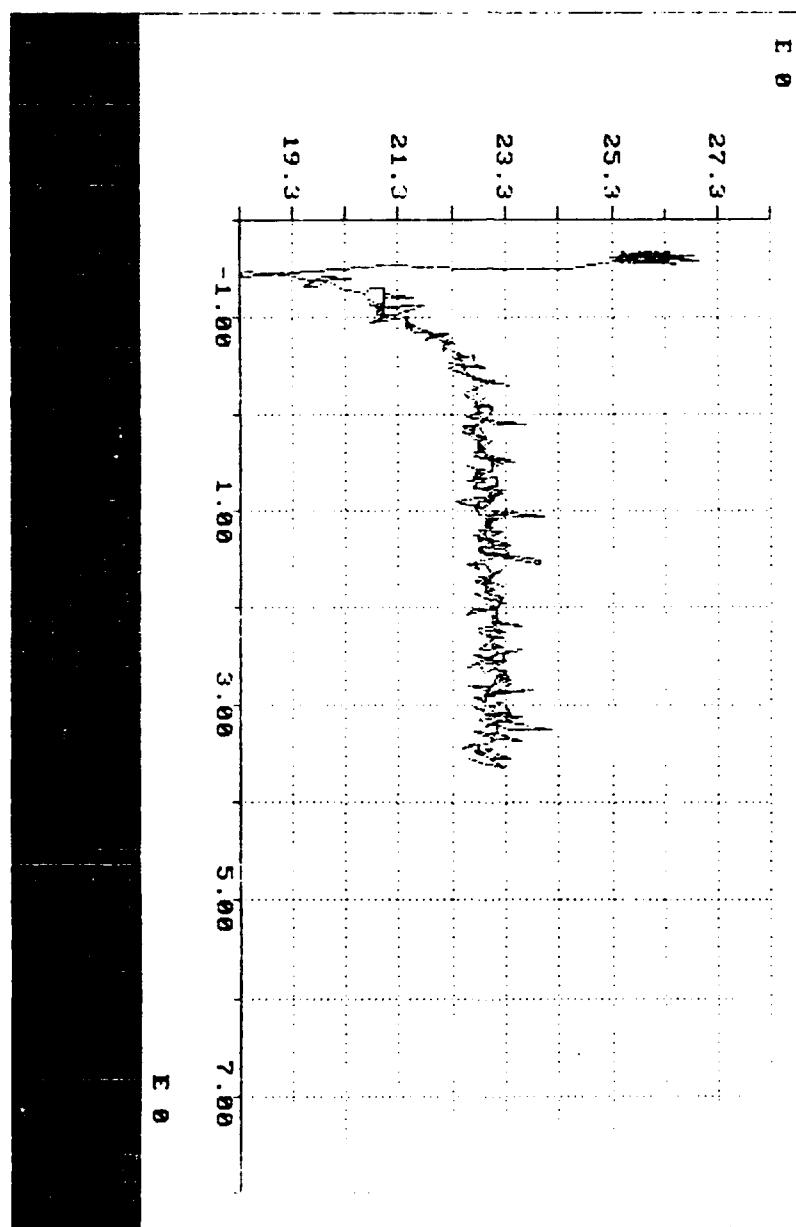
: Subfile: 1

: Row (0=all): 0

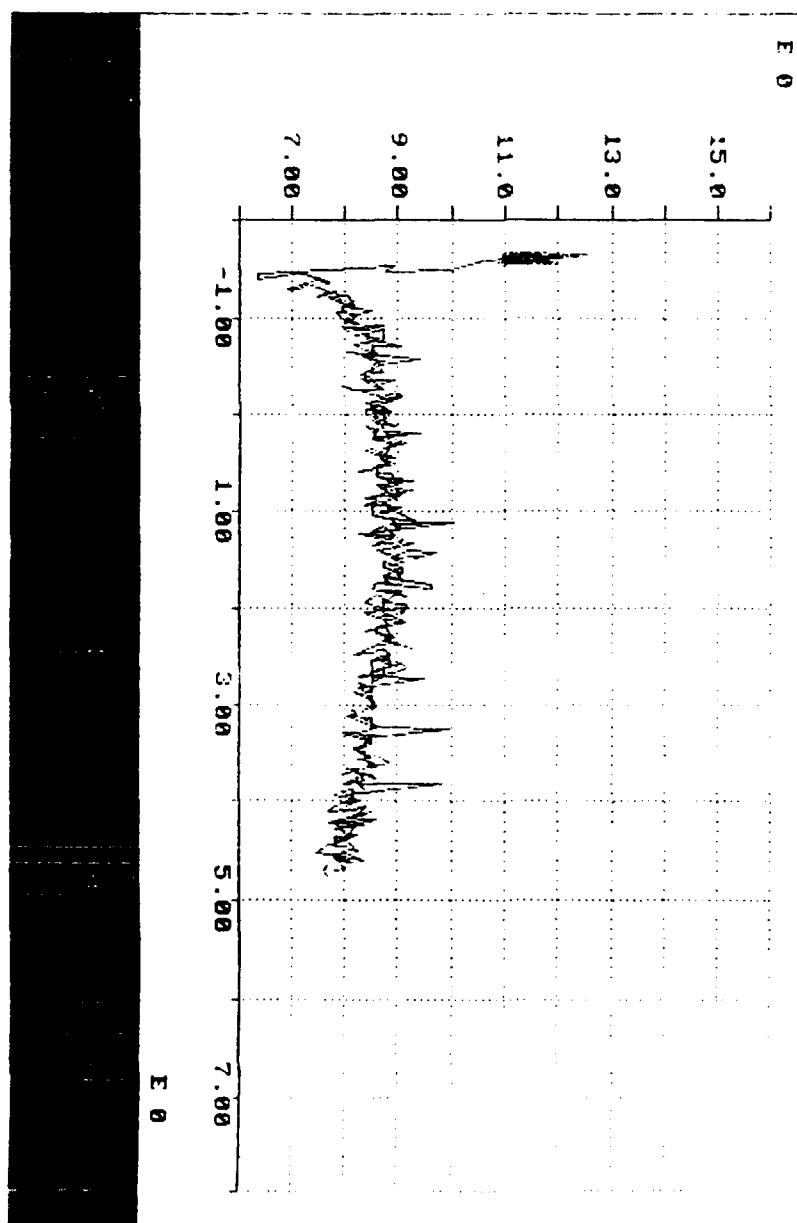
: Start Column:

: # of Columns:

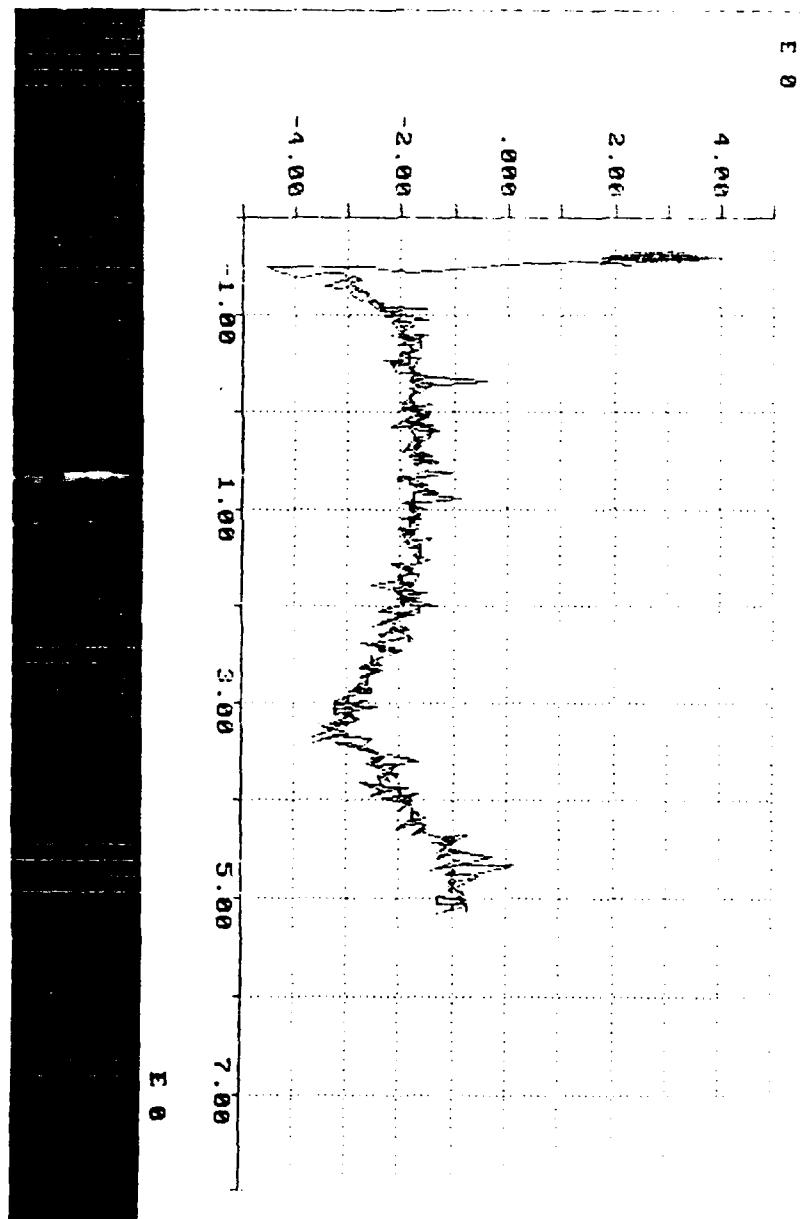
: Read/Write/Append: R



```
: File Name: C:\TEMP\TEST.DAT          Date: 10/10/97
:
: 8 Comments
:
: 1 BLOOD CONTROLS
: 2 BLANCHING AND SWELLING
: 3 MIGHT BE A WELD
: 4
: 5 A WEAK WELD
: 6
: 7
: 8
:
: I/O Variable (Type)          Date: 10/10/97
:
: Subfile:
: Row (0=All):
: Start Column:
: # of Columns:
:
: Read/Write/Append/Replace/First/Last/More/Delete *
```



: File Name: 10-10-00.DAT
:
: 8 Comments
: 1 BLOOD CONTINUOUSLY
: 2 BLANCHING OCCURS
: 3 NO SMOKING
: 4 LOOSES LIKE A
: 5
: 6 MODERATE W.
: 7 TORE AT SEEM
: 8
:
: I/O Variable (E.g., X)
:
: Subfile: 1
: Row (0=all): 0
: Start Column: 1
: # of Columns: 100
:
: Read/Write/Append: R
:
:



File Name: E0

Comments:

1. EOD CUTTING
2. ELANCING AND TORN
3. SMALL AMOUNT OF METAL
4. LOOKS LIKE A WELD
5.
6. MODERATE WELD
7. TORE AT SEVERAL PLACES

I/O Variable (E0)

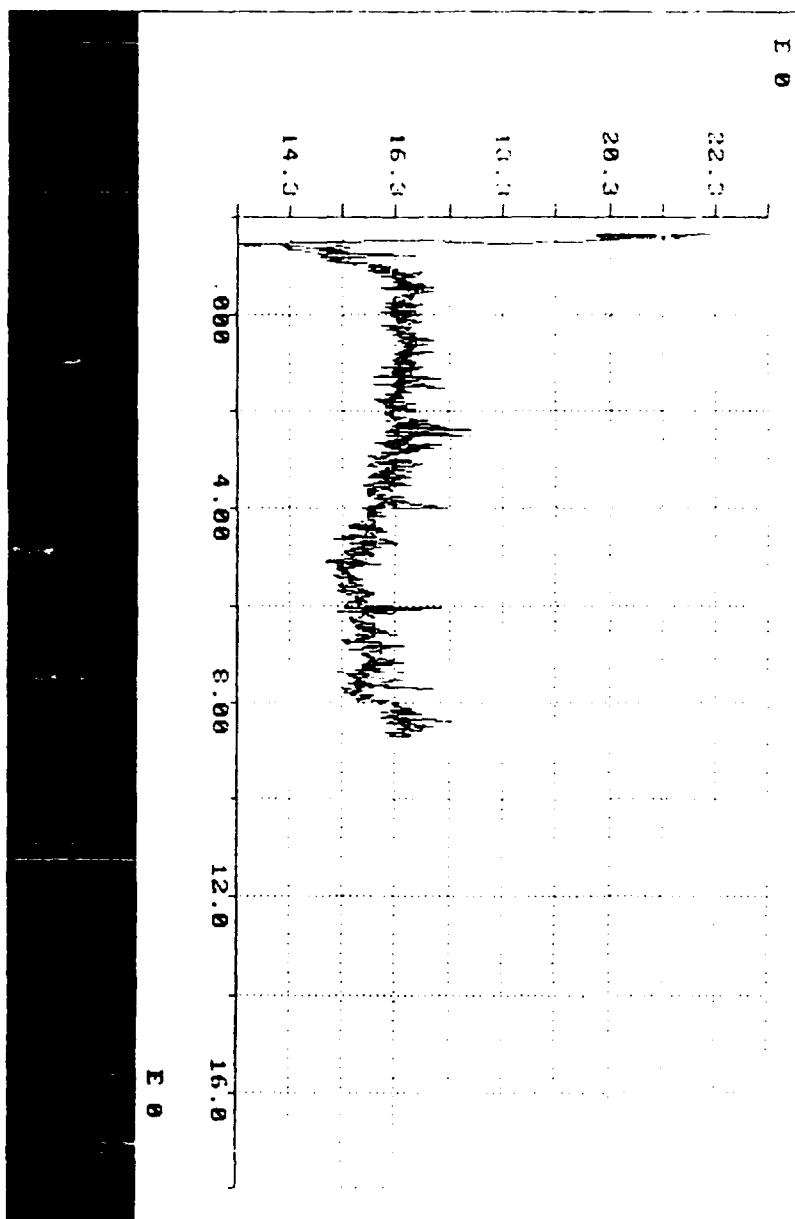
Subfile:

Row (0=all):

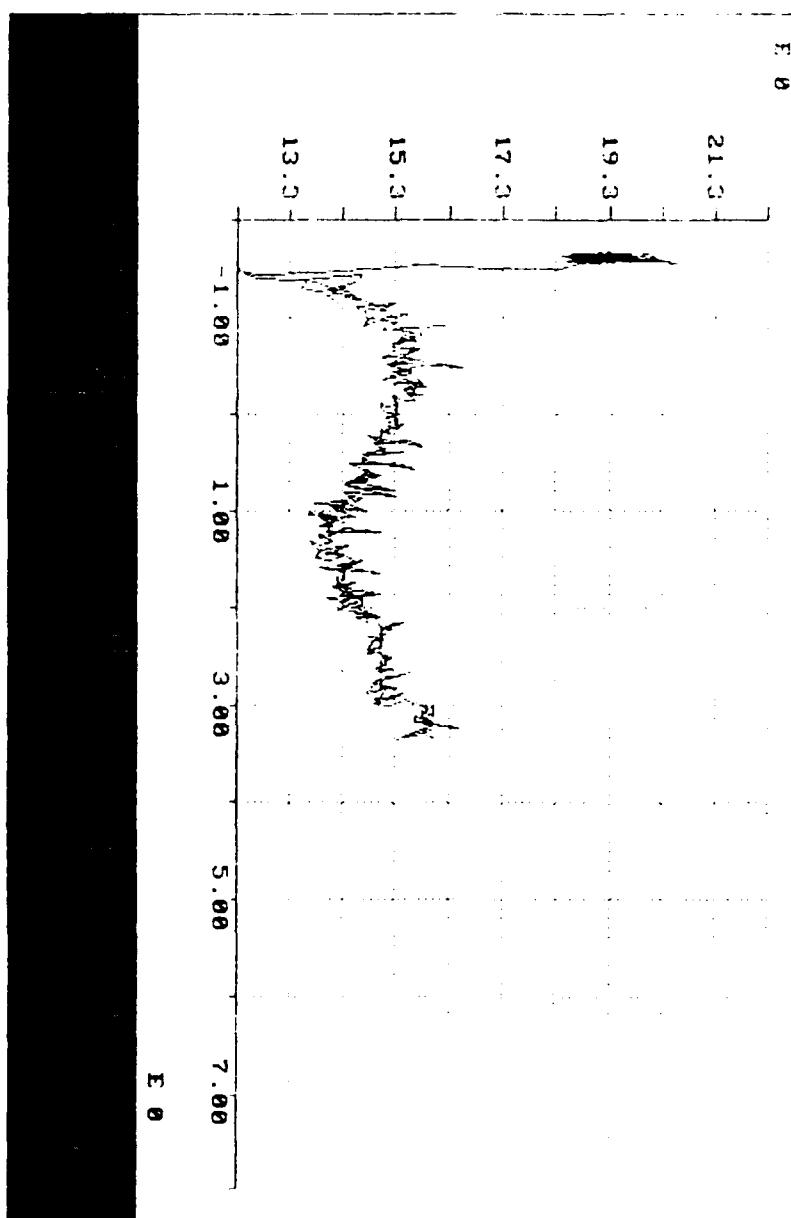
Start Column:

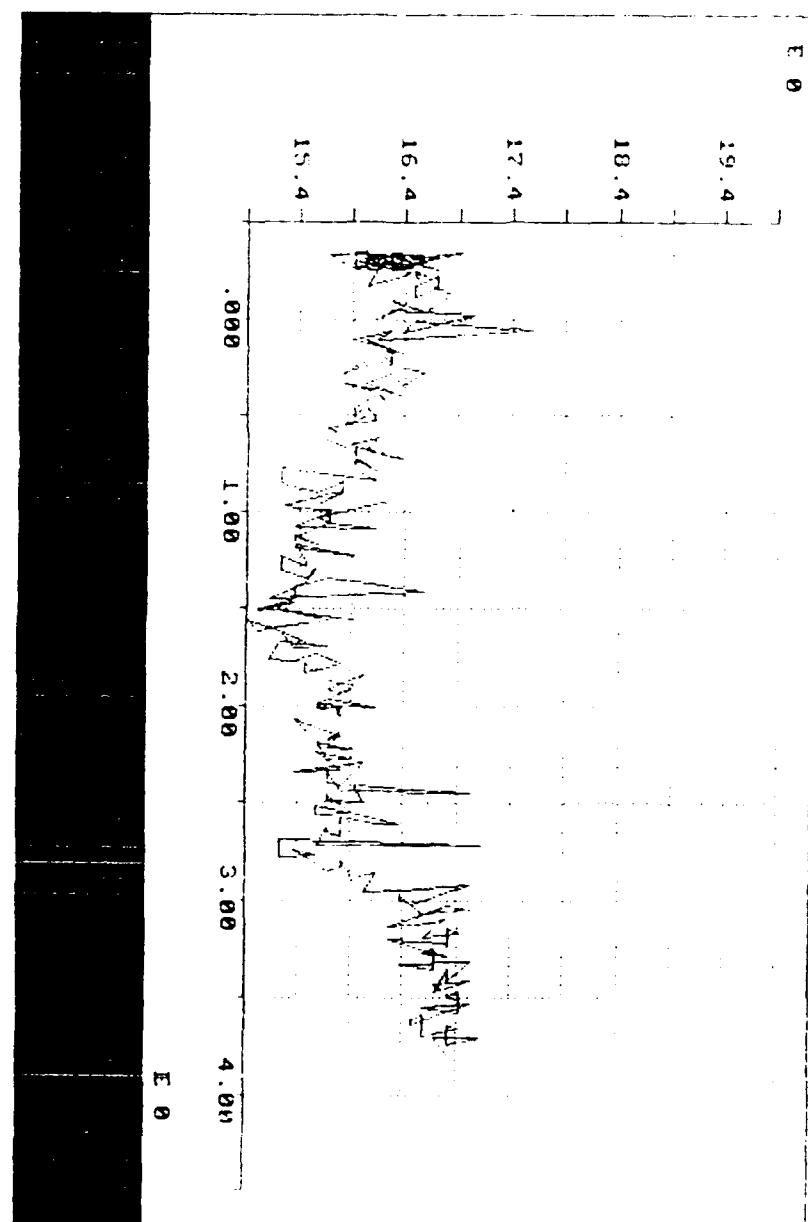
of Columns:

Read/Write/Append (read/write/append) (0=append to file name)

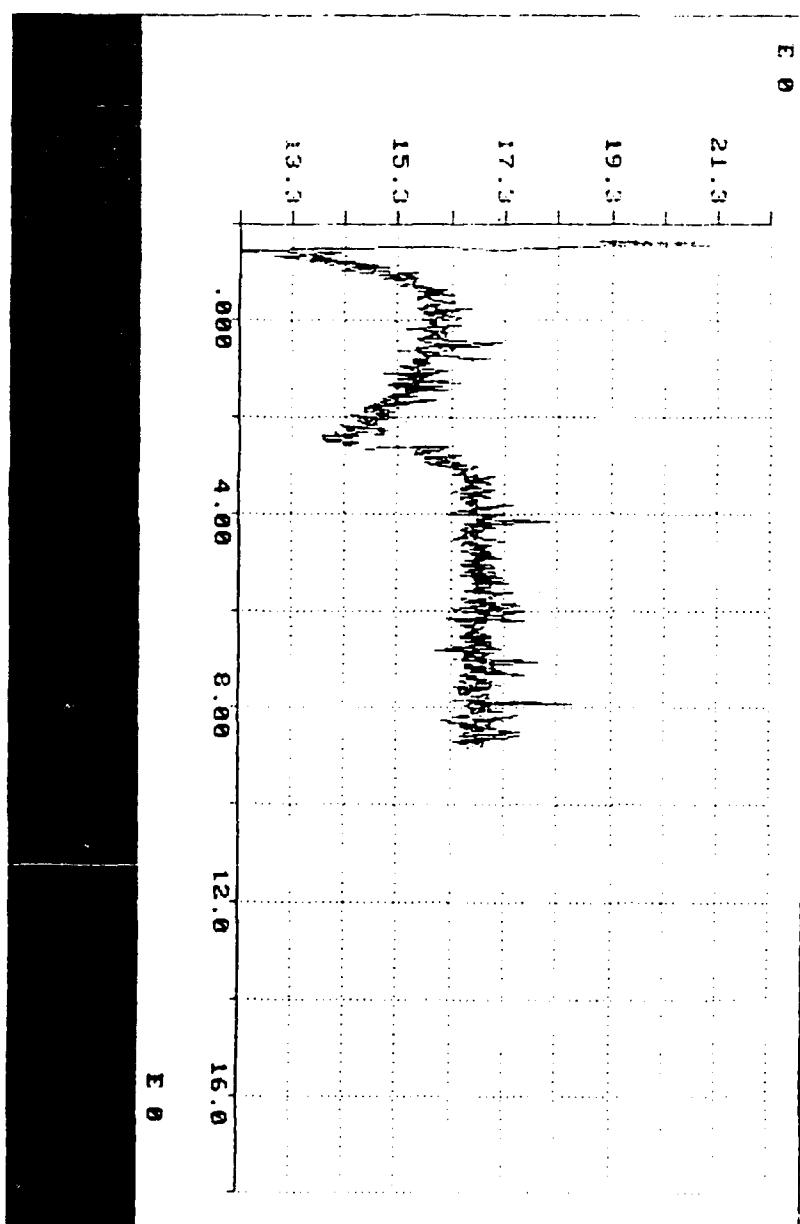


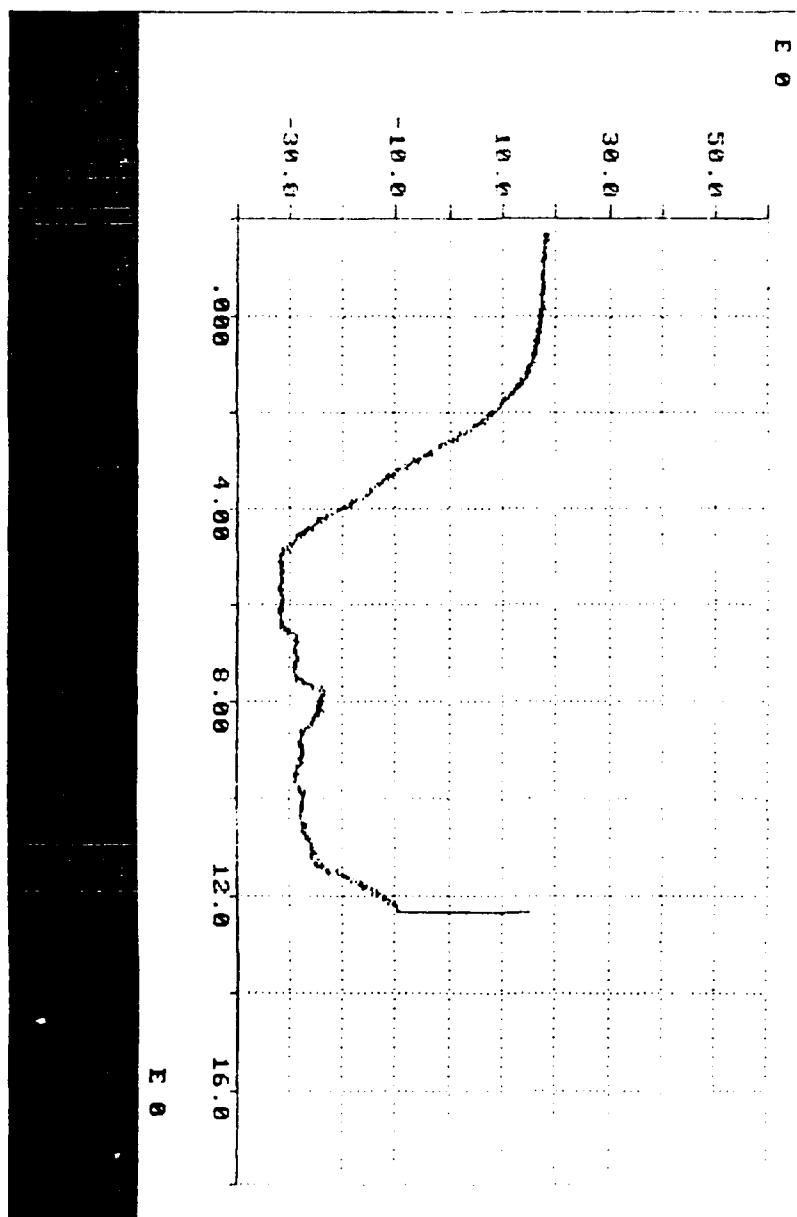
```
: File Name: C:\TEMP\TEST.DAT
: B. Comments:
: E1 BLOOD LOSS
: E2 FLANCHING NO
: E3 MODERATE SWELLING
: E4 PEARL WOUND
: I/O Variable (I=Input, O=Output)
: Subfile:
: Row (0=all):
: Start Column:
: # of Columns:
: Read/Write/Append (R=Read, W=Write, A=Append)
```

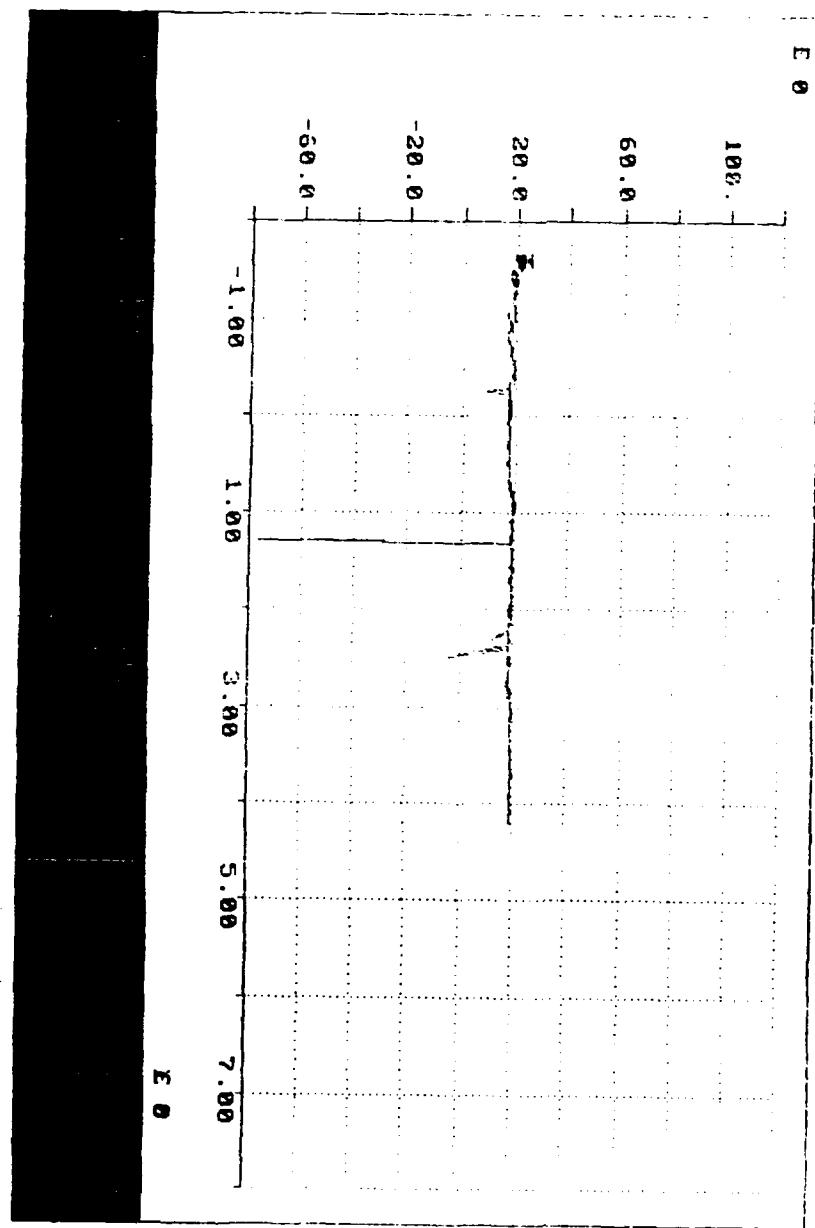




```
: File Name: C:\TEST.DAT
: Subfile:
: 8 Comments
: 1 BLOOD CONTROLS
: 2 BLANCHING AND WELDING
: 3 NO CHARGING
: 4 NO SMOKE
: 5 LOOKS WELDED
: 6 BLOTTED SEMI-SMOOTH
: 7 BLOOD CLOTH
: 8 PROBE AT SIDE OF HEAD
: I.O Variable (1)
: Subfile:
: Low (0=all):
: Start Column: 1
: # of Columns: 1
: Read/Write/Append (r=w=rw) Edit/Get/Print/Find/Replace
```







E 0

36.0

22.0

20.0

12.0

4.00

-1.00

3.00

5.00

7.00

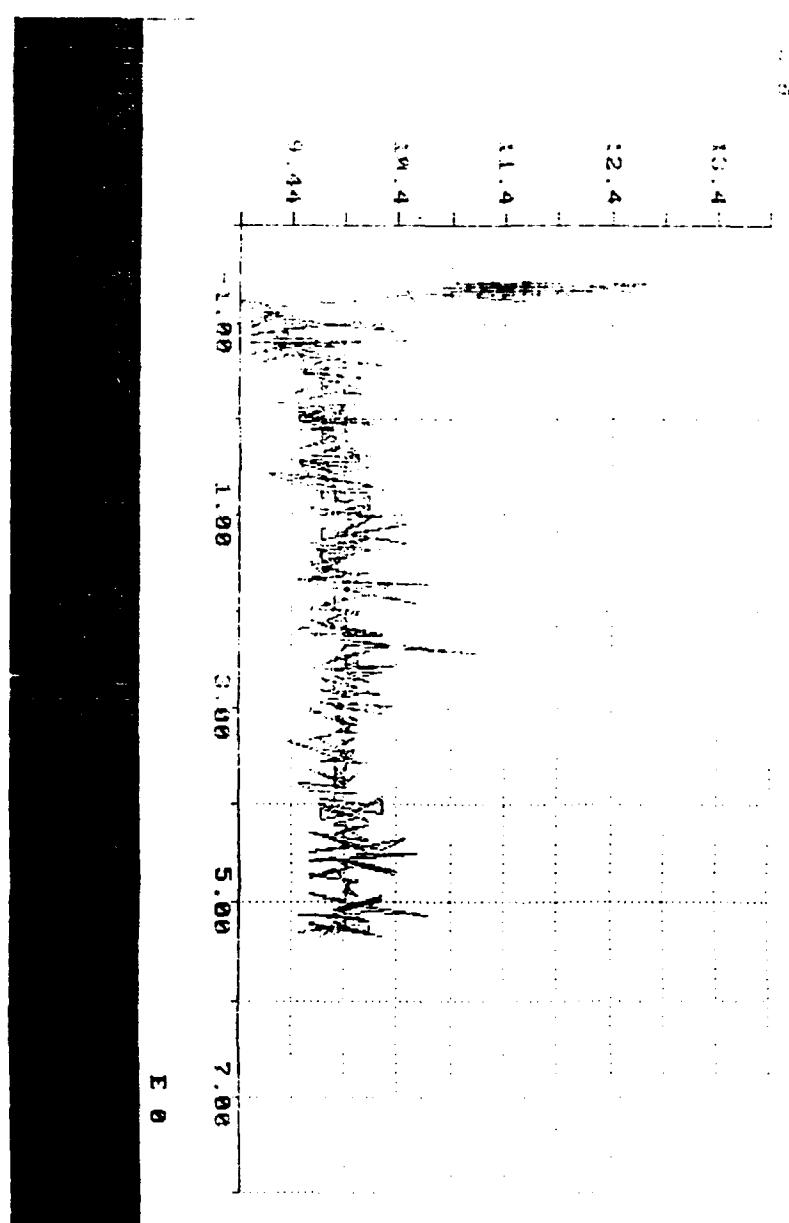
E 0

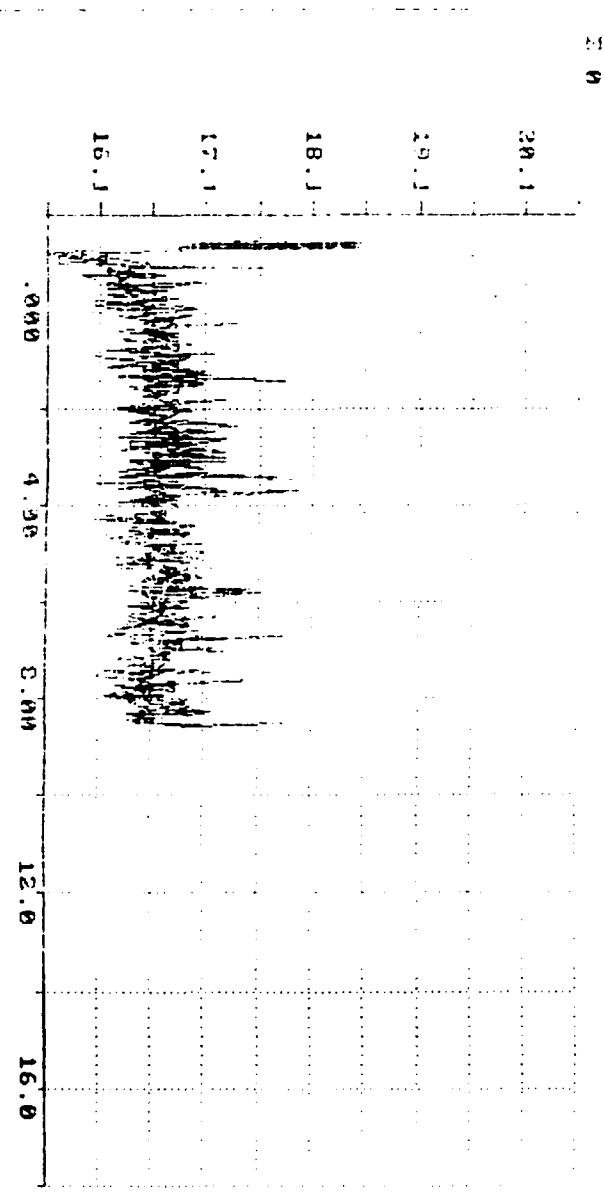
File Information:

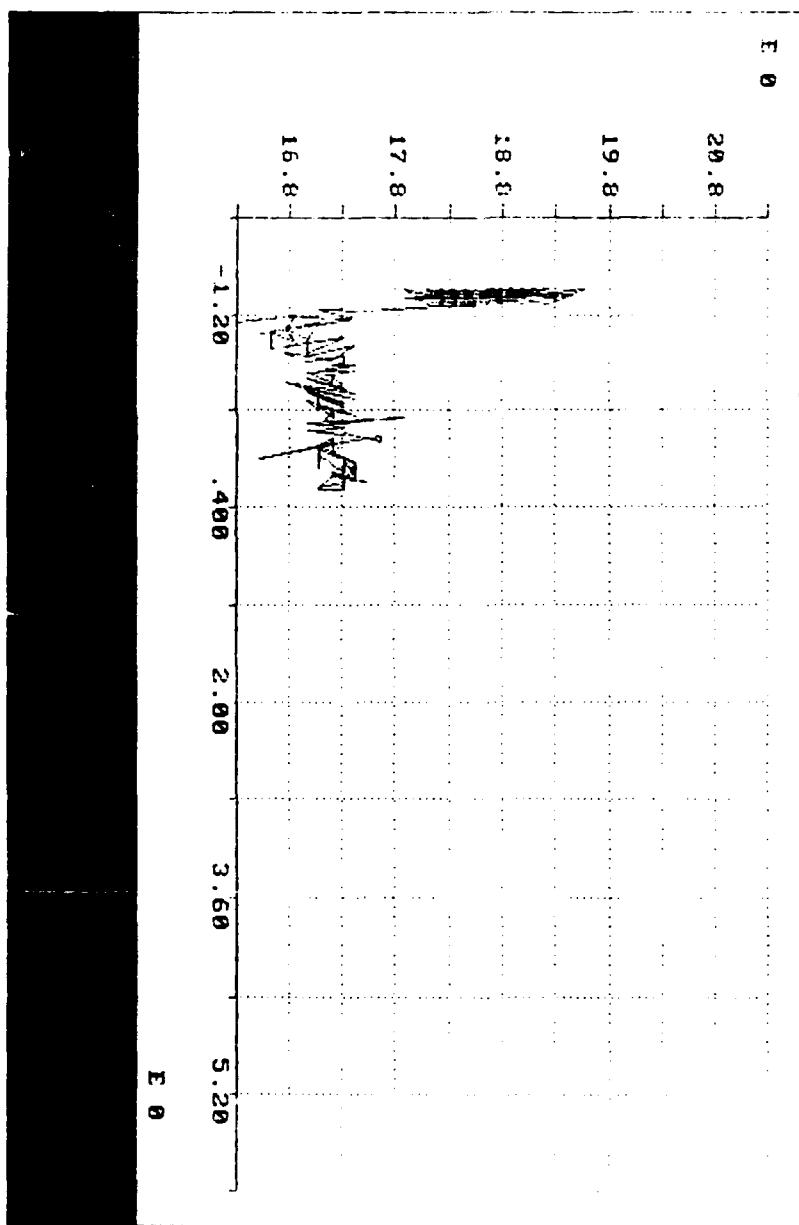
: File Name: DATA
:
: Comments:
Data Information:
: 1. LOG COUNT: 0.000000
: 2. N. VISIBLE CARS: 0
: 3. NO. SNAPS:
:
: 4. FIELD

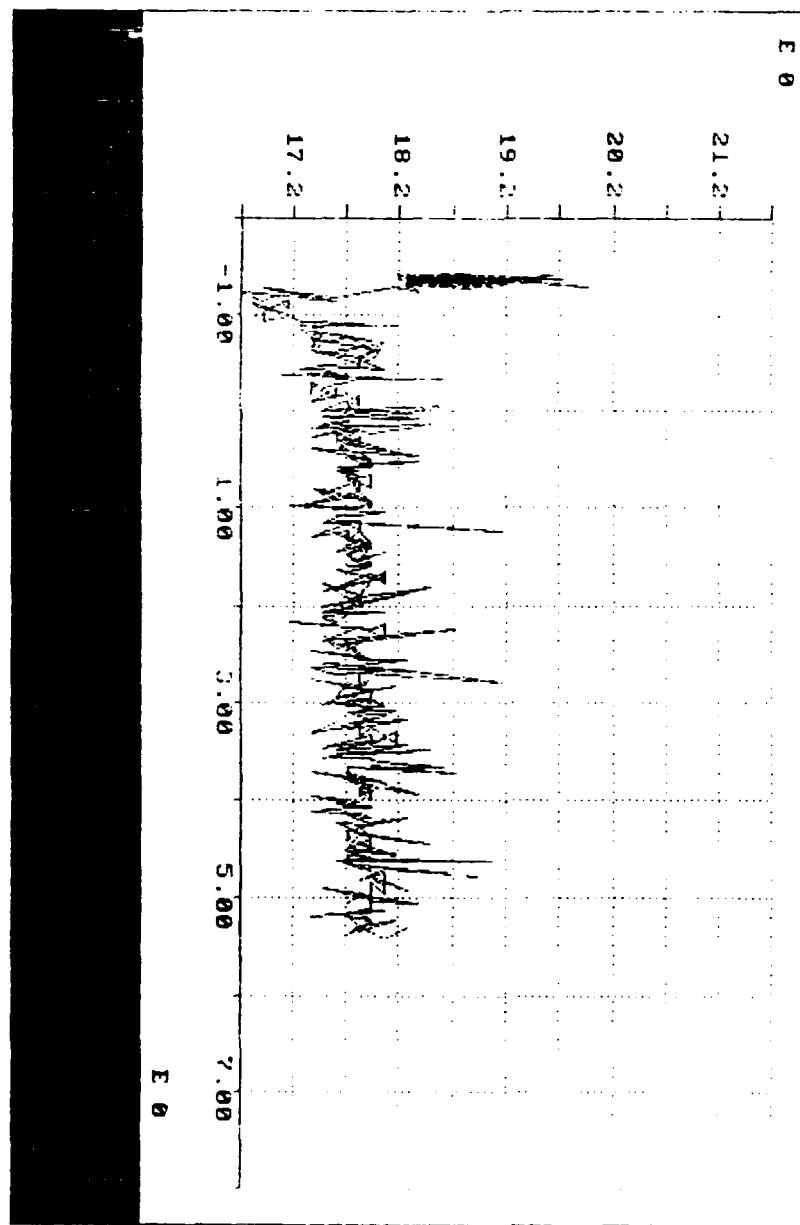
Variable Information:

: Variable 1
:
: 1. Title: HEALTH
: Start Column: 1
: # of Columns: 1
:
: Read/Write/Setup:
:
: End









File Name: (*.*)

S Comments

INDIA INK UNWFLIN P (1024, 0)

1

2

3

4

5

6

7

8

I/O Variable (1 to 1):

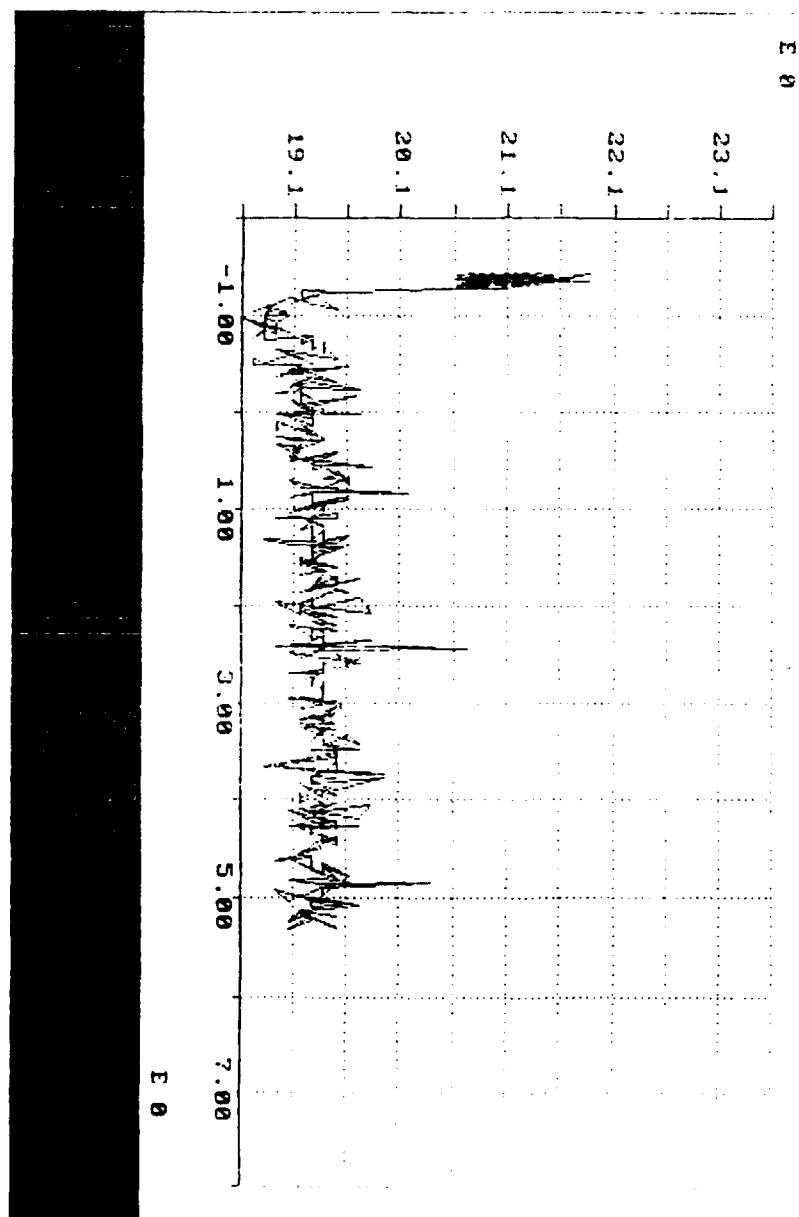
Subfile:

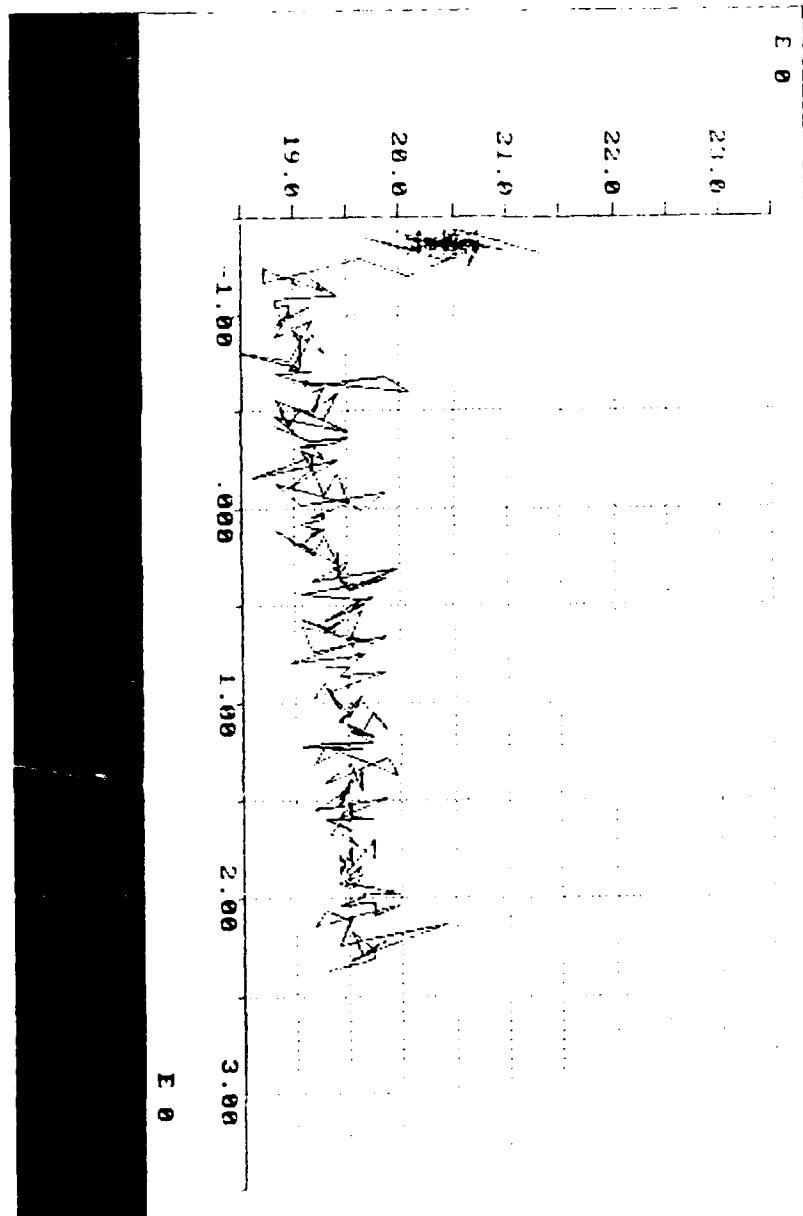
Row (0=all): 0

Start Column: 1

of Columns: 128

Read/Write/Append (String): Read





IMMMIMMMIMMMIMMMIMM

: File Name: .00

: 0 Comments

LMIMIMIMIMIMIMIMIMIM

: 1) INDIA INK CONT.

: 2) NO VISIBLE CHAM

: 3) NO SMOKE

: 4) NO DUST

: 5) NO WELD

: 6) NO

LMIMIMIMIMIMIMIMIMIM

: I/O Variable (

: Subfile:

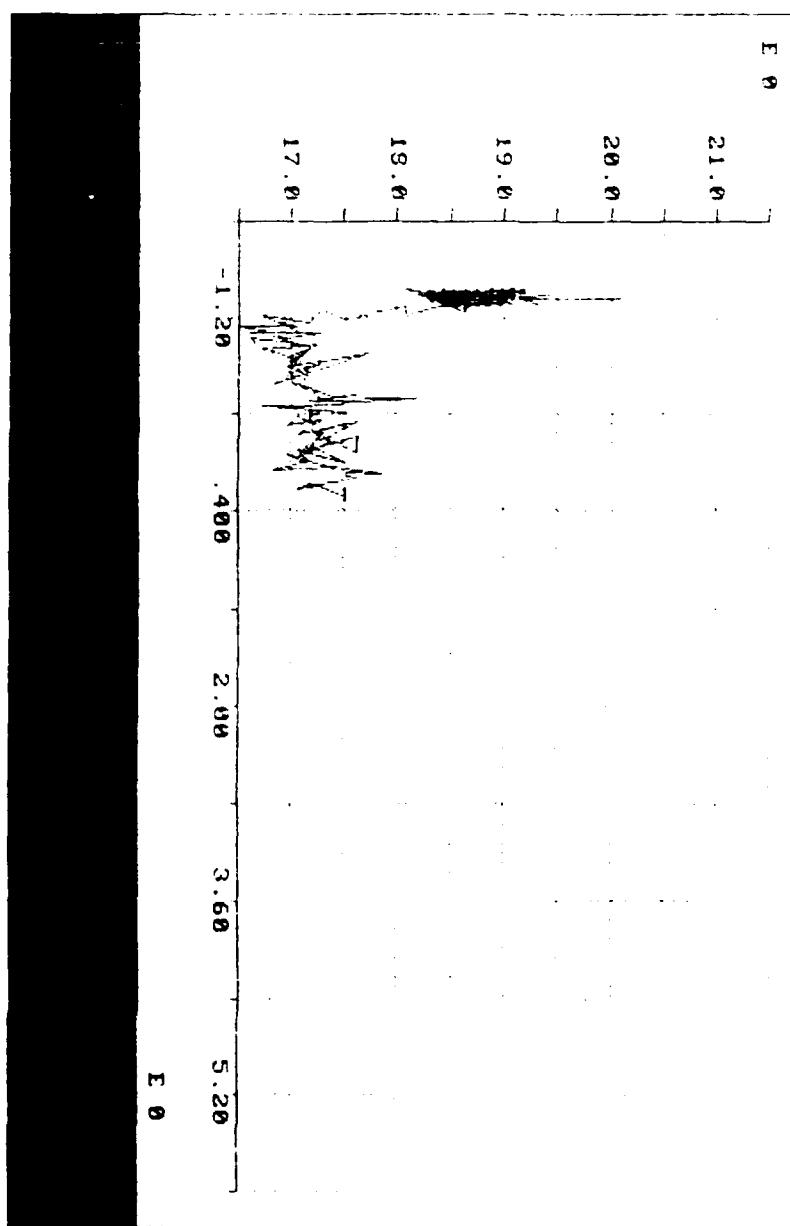
: Row (0=all): 00

: Start Column: 00

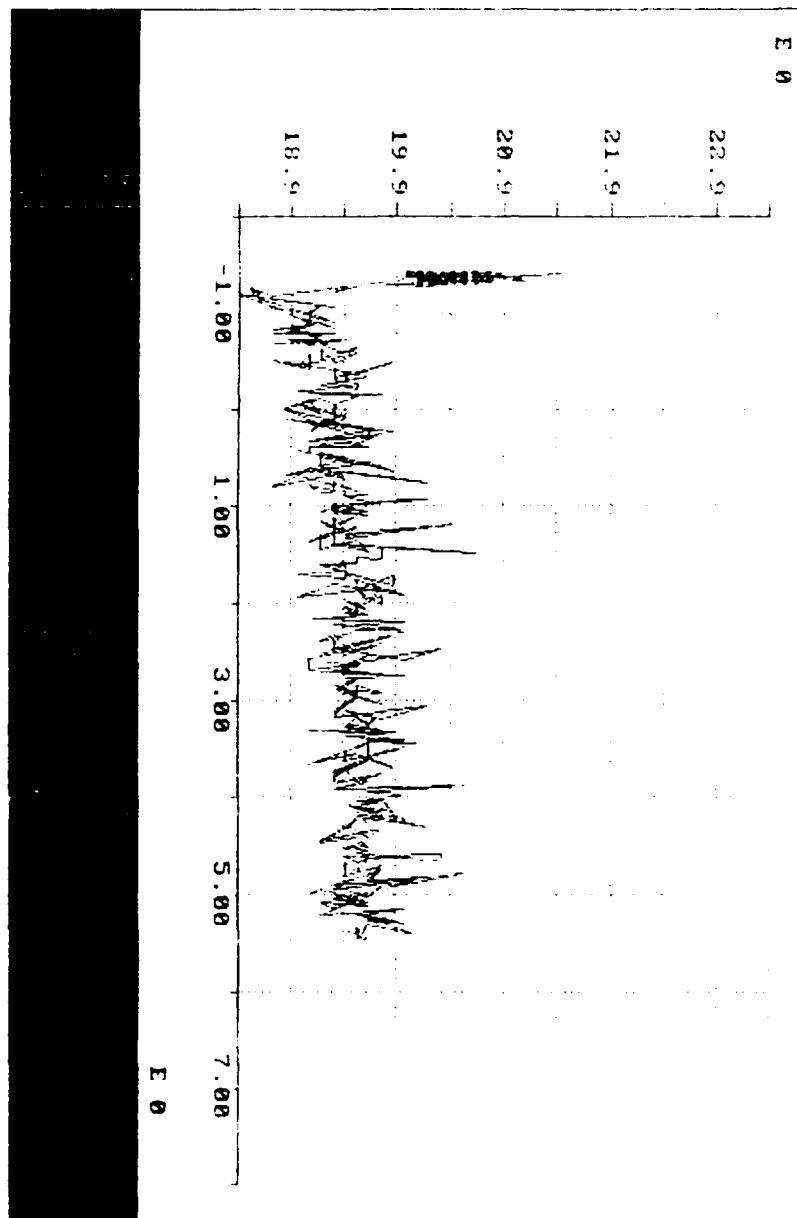
: # of Columns: 256

: Read/Write/Append:

: E



```
: File Name: C:\T\T  
:  
: # Columns:  
LMMMMMMMMMM  
: 1 DATA IN CSV  
: 2 LIGHT DEFECTS  
: 3 ALUMINUM FLAME  
: 4 NO SMOKE  
: 5  
: 6 NO WELD  
:  
:  
: 8  
LMMMMMMMMMM  
: IVD Variable (1)  
:  
: Subfile:  
: Row ("all"):  
: Start Column:  
: # of Columns:  
:  
: Read/Write/Append (r):
```



File Name: INDIA.DAT

B Comments:

INDIA INK, COULD BE INK, SLIGHT DESIGNS, NO SMOKE, NO CHARTS, NOT MUCH OF A

B

I/O Variable C

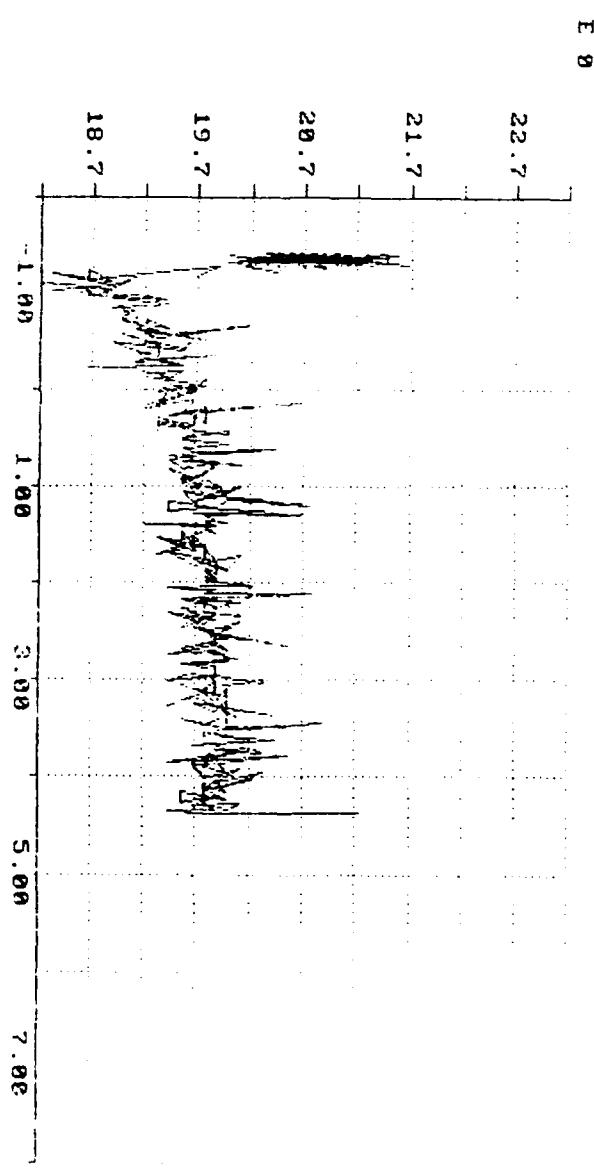
Subfile: 1

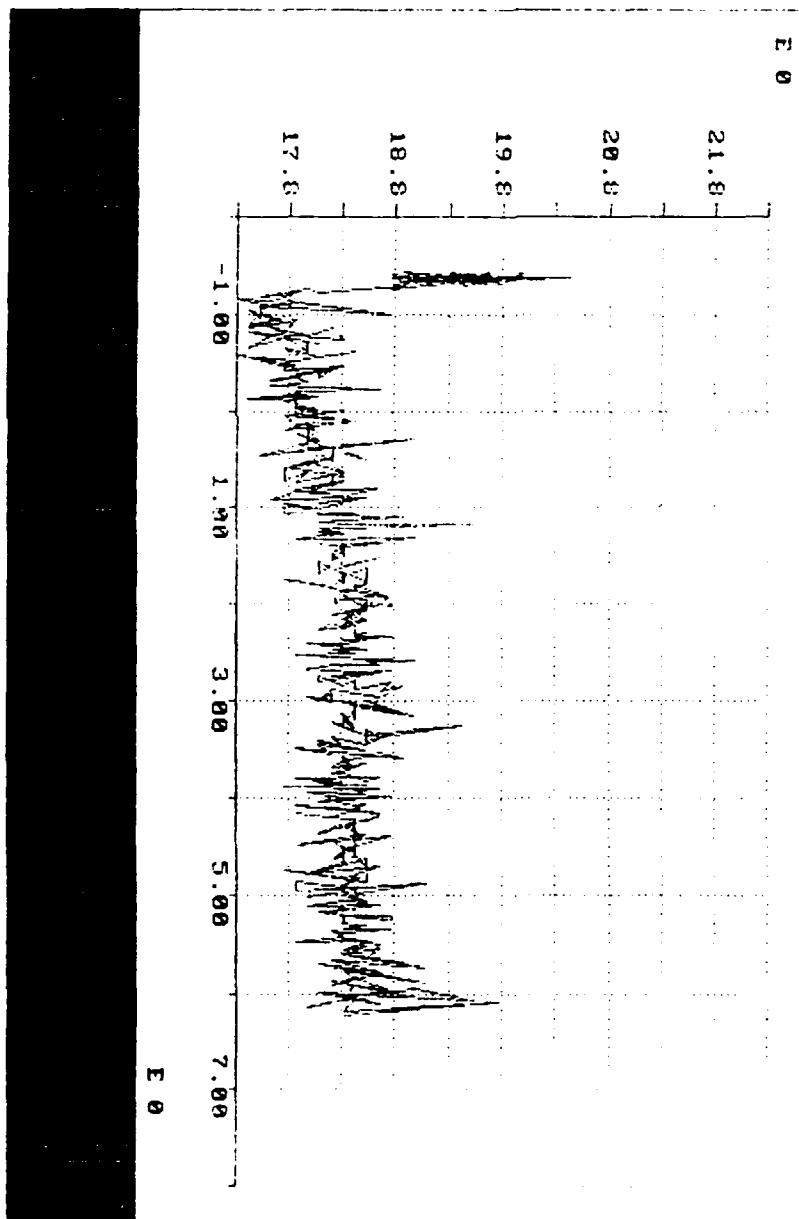
Row (0=all): 0

Start Column: 1

of Columns: 15

Read/Write/Append: R





Appendix C- Supplemental Figures and Illustrations

Figure 1- Thermocouple Placement

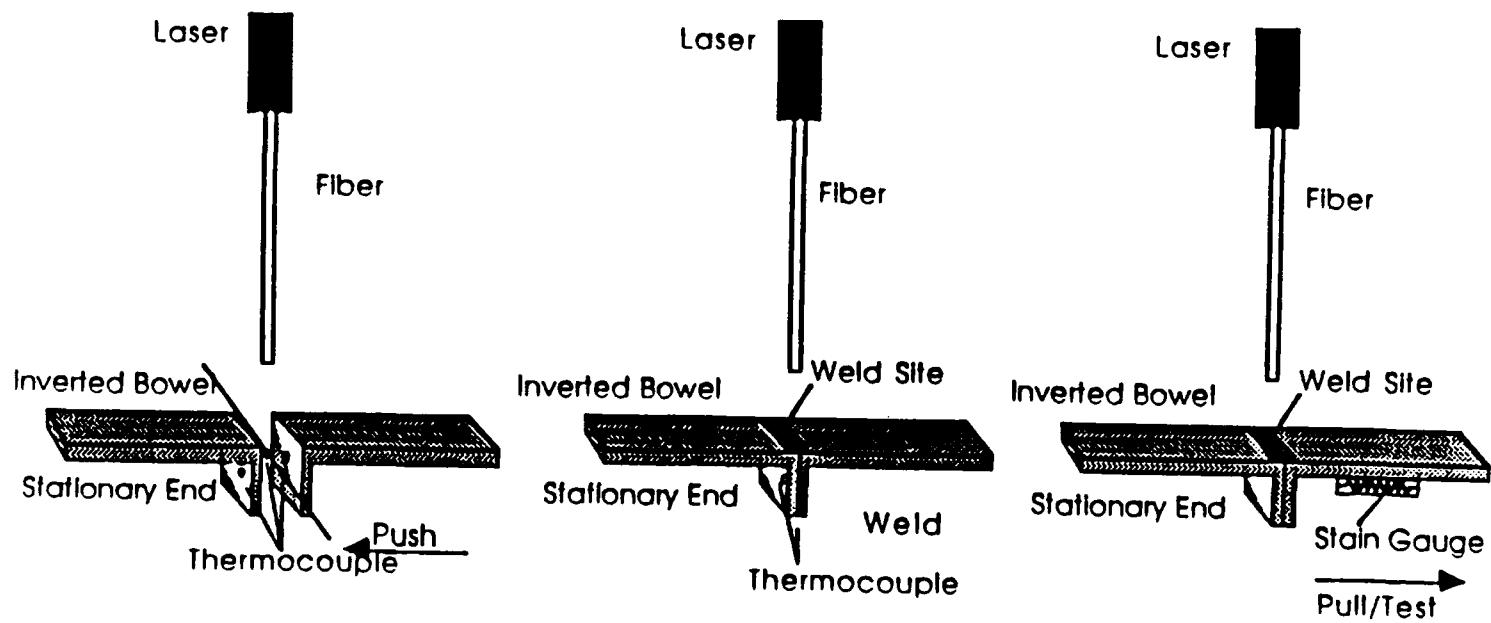


Figure 2- Experimental Protocols

LASER TISSUE WELDING OPTIMIZATION PROTOCOL

<u>Time (s)</u>	<u>Aperture Size (mm)</u>	<u>Power (W)</u>	<u>Chromophore</u>
2	0.5 x 4.0	0.05	None
10	0.5 x 4.0	0.05	None
2	2.0 x 4.0	0.05	None
10	2.0 x 4.0	0.05	None
2	0.5 x 4.0	2.0	None
10	0.5 x 4.0	2.0	None
2	2.0 x 4.0	2.0	None
10	2.0 x 4.0	2.0	None
2	0.5 x 4.0	5.0	None
10	0.5 x 4.0	5.0	None
2	2.0 x 4.0	5.0	None
10	2.0 x 4.0	5.0	None
2	0.5 x 4.0	0.05	India Ink
10	0.5 x 4.0	0.05	India Ink
2	2.0 x 4.0	0.05	India Ink
10	2.0 x 4.0	0.05	India Ink
2	0.5 x 4.0	2.0	India Ink
10	0.5 x 4.0	2.0	India Ink
2	2.0 x 4.0	2.0	India Ink
10	2.0 x 4.0	2.0	India Ink
2	0.5 x 4.0	5.0	India Ink
10	0.5 x 4.0	5.0	India Ink
2	2.0 x 4.0	5.0	India Ink
10	2.0 x 4.0	5.0	India Ink
2	0.5 x 4.0	0.05	ICG
10	0.5 x 4.0	0.05	ICG
2	2.0 x 4.0	0.05	ICG
10	2.0 x 4.0	0.05	ICG
2	0.5 x 4.0	2.0	ICG
10	0.5 x 4.0	2.0	ICG
2	2.0 x 4.0	2.0	ICG
10	2.0 x 4.0	2.0	ICG
2	0.5 x 4.0	5.0	ICG
10	0.5 x 4.0	5.0	ICG
2	2.0 x 4.0	5.0	ICG
10	2.0 x 4.0	5.0	ICG
2	0.5 x 4.0	0.05	Blood
10	0.5 x 4.0	0.05	Blood
2	2.0 x 4.0	0.05	Blood
10	2.0 x 4.0	0.05	Blood
2	0.5 x 4.0	2.0	Blood
10	0.5 x 4.0	2.0	Blood
2	2.0 x 4.0	2.0	Blood
10	2.0 x 4.0	2.0	Blood
2	0.5 x 4.0	5.0	Blood
10	0.5 x 4.0	5.0	Blood
2	2.0 x 4.0	5.0	Blood
10	2.0 x 4.0	5.0	Blood

Rabbit Animal Prep Protocol

Supplies Needed:

Alcohol Preps

1 ml Tuberculin Syringe

3 ml Syringe

#18 Needle

#22 Needle

Gemini Xylazine (Rompun)

Ketaset (Ketamine)

100 ml. bag of 0.9% NaCl or 5% Dextrose in Water for intravenous use

Microdrip (60 drops per ml) dispenser and tubing

Animal Clippers

Manual (non-electric) Razor

Artificial Tears Ointment

#22 JELCO IV Catheter Placement Unit

Paper Tape

Beuthanasia (sodium pentobarbital or formerly known as Sleepaway)

Procedure:

1. One day prior to desired surgery, remove food container from rabbit's cage. Leave water bottle.
2. Prepare tray of surgical instruments for autoclaving. Place blue cloth drape under and over instruments. Add hemoclips, needle holders, atraumatic clamps, and other specialized instruments as needed. Autoclave instruments.
3. Collect all supplies listed above. Place in rabbit room.
4. Close door to rabbit room.
5. Determine amount of Rompun injection. A chart which indicates the appropriate amount to inject is appended to this document.
6. Prepare Rompun injection. Wipe the septum of the Rompun bottle with an alcohol prep. Remove tuberculin syringe from packaging. Insert syringe into bottle and carefully dispense correct amount.
7. With syringe in hand, gently and quietly open door to selected rabbit's cage. Pet rabbit gently to reassure. After calming rabbit, obtain control over the animal by grasping firmly at the neck. Hold down rabbit and inject Rompun into the muscle in the hind quarter.
8. Close rabbit cage. Wait approximately 10 minutes for rabbit to become sedated. Rabbit may slouch or turn head to side.
9. Remove rabbit from cage, supporting head and neck carefully.
10. Transfer rabbit to sink.
11. Using chart in appended to this document and the actual weight of the rabbit, determine the appropriate amount of Ketaset to inject.
12. Prepare injection of Ketaset. Remove the 3 ml. syringe from its packaging and place the #18 needle on the end of the syringe. Wipe the septum of the Ketaset bottle with an alcohol prep, and insert syringe. Remove desired amount of Ketaset and then carefully change needles from the #18 to the #22. Grasp the rabbit firmly and inject Ketaset, using the #22 needle, into the hind quarter.
13. Wait approximately 10 minutes for the rabbit to become completely anesthetized. The rabbit is completely anesthetized when it can be picked up and placed on its back and does not attempt to turn itself over.
14. While waiting, prepare intravenous anesthetic cocktail. To the 100 ml. 0.9% NaCl bag add 4.0 ml. Rompun and 8.0 ml. Ketaset. Invert bag to mix. Hang bag on IV pole. Insert microdrip and tubing into bag and then remove plastic cover from distal end of tubing. Place three or four strips of paper tape on the IV pole for later use.
15. Place three or four paper towels on the countertop next to the sink and two paper towels inside the sink. Place rabbit, belly up, on the countertop.

16. Apply Artificial Tears to the rabbit's eyes to prevent painful drying.
17. Using a marker, number the rabbit by writing its identification number from the experimental protocol inside its ear
18. Use electric clippers to shave appropriate area of rabbit, usually abdomen and groin. Push excess fur onto the paper towels. Quickly remove paper towels and fur as soon as clipping is completed.
19. With non-electric razor, gently remove hair from the ears, particularly along the external vein. This vein will be used for the intravenous anesthetic.
20. Transfer the rabbit to the surgical site.
21. Wipe the external ear vein region with an alcohol prep. This should improve visualization of the vein.
22. Remove JELCO catheter from its packaging. Insert catheter into the external vein. Use needle to enter vein and then pull back on the needle so that the catheter slides along the inside of the vein. Watch catheter slide inside the vein. Look for a backflow of blood into the top part of the catheter placement unit. Remove the needle completely when the catheter has been correctly positioned.
23. Attach the tubing from the IV bag to the top part of the catheter placement unit. Test catheter placement by slowly opening the anesthetic flow regulator and watching for anesthesia flow. Alternatively, open the saline regulator if one has been piggy-backed, and watch its flow. When adequate placement is verified, carefully tape the tubing and the catheter placement unit to the rabbit's ear. Tape excess tubing to the table.
24. Adjust the anesthetic flow rate to 1 drop every 4 seconds. Watch the rabbit's chest rise and pupil size to monitor anesthesia. If rabbit begins to come out of the anesthesia, increase rate to 1 drop every 2 seconds for about 1 minute. Return to 1 drop every 4 seconds, and wait until rabbit is sedated (1-2 minutes).
25. If rabbit is to be euthanized, inject 4.0 ml. of 1:1 diluted Beuthanasia into the septum of the IV line. The bottle will indicate if it has already been diluted. If it has not been diluted, mix equal parts of concentrated Beuthanasia and 0.9% NaCl, and inject this solution into the IV line. Do not inject concentrated Beuthanasia into the IV line because it is very viscous, and therefore, it is difficult to force it out of the tubing and into the rabbit.
26. After rabbit has died, remove IV line and place rabbit remains into a large brown plastic bag.
27. Carefully wash instruments. Alconox or similar detergent should be used. Never leave instruments soaking in water. Rinse instruments and place them on a towel to dry.

RABBIT ANESTHESIA

INITIAL INTRAMUSCULAR DOSE OF XYLAZINE GIVEN, FOLLOWED
10 MINUTES LATER BY AN INTRAMUSCULAR DOSE OF KETAMINE.
DOSAGE BASED ON WEIGHT OF RABBIT.

WEIGHT(LBS.)	XYLAZINE 20 MG/ML ML	KETAMINE 100 MG/ML ML
4	0.5	0.72
4.25	0.53	0.77
4.5	0.56	0.81
4.75	0.59	0.86
5	0.63	0.9
5.25	0.66	0.95
5.5	0.69	0.99
5.75	0.72	1.04
6	0.75	1.08
6.25	0.78	1.13
6.5	0.81	1.17
6.75	0.84	1.22
7	0.88	1.26

INTRAOPERATIVE ANESTHESIA CONSISTS OF 4 MG KETAMINE +
2 MG XYLAZINE IN 50 ML DILUENT, INFUSED AT 1 DROP EVERY
4-6 SECONDS FOR A 6-8 LB. RABBIT.

Tissue Preparation for Weld Strength Optimization Study

1. Make a midline incision with a #15 scalpel.
2. Identify and isolate the small bowel.
3. Place an atraumatic clamp across the mesentery at the section to be resected.
4. Place a hemoclip at the site of the nearest vessels to occlude blood flow.
5. Resect a small segment of bowel measuring approximately 5 cm.
6. Return the remaining *in situ* bowel to the abdomen.
7. Make a longitudinal cut in the bowel segment, and open it up. Carefully rinse the bowel segment in physiological saline to remove succus entericus.
8. Using the specialized cutters, cut two strips measuring 4.0 x 20.0 mm. Place the two strips into the clamps, making sure that the strips are aligned relative to the marks on the clamps.
9. Apply a barely visible amount of chromophore with a cotton-tipped applicator. The chromophore should be applied to both tissue strips.
10. Position the thermocouple between the two tissue strips.
11. Slowly bring the two tissue strips into apposition using the computer-controlled piezoelectric motor.
12. Verify appropriate thermocouple placement (1 mm below surface of weld).
13. Set laser welding parameters in laser welding control program. Weld tissue.
14. Switch to the tensiometry data acquisition program. Pull tissue apart and measure the load required to rupture the weld.
15. Repeat steps 5-14 to prepare additional tissue strips.
16. Import data into Lotus or Symphony to generate load versus distance plots.